

Hydrogen Sulfide in Biochemistry and Medicine

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Abstract

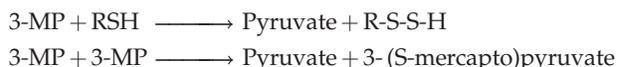
Significance: An abundance of experimental evidence suggests that hydrogen sulfide (H₂S) plays a prominent role in physiology and pathophysiology. Many targets exist for H₂S therapy. The molecular targets of H₂S include proteins, enzymes, transcription factors, and membrane ion channels. **Recent Advances:** Novel H₂S precursors are being synthesized and discovered that are capable of releasing H₂S in a slow and sustained manner. This presents a novel and advantageous approach to H₂S therapy for treatment of chronic conditions associated with a decline in endogenous H₂S, such as diabetes and cardiovascular disease. **Critical Issues:** While H₂S is cytoprotective at physiological concentrations, it is not universally cytoprotective, as it appears to have pro-apoptotic actions in cancer cells and is well known to be toxic at supraphysiological concentrations. Many of the pleiotropic effects of H₂S on health are associated with the inhibition of inflammation and upregulation of prosurvival pathways. The powerful anti-inflammatory, cytoprotective, immunomodulating, and trophic effects of H₂S on the vast majority of normal cells seem to be mediated mainly by its actions as an extremely versatile direct and indirect antioxidant and free radical scavenger. While the overall effects of H₂S on transformed (*i.e.*, malignant) cells can be characterized as pro-oxidant and pro-apoptotic, they contrast sharply with the cytoprotective effects on most normal cells. **Future Directions:** H₂S has become a molecule of great interest, and several slow-releasing H₂S prodrugs are currently under development. We believe that additional agents regulating H₂S bioavailability will be developed during the next 10 years. *Antioxid. Redox Signal.* 17, 119–140.

Introduction

ORGAN AND CELL FUNCTION ARE REGULATED by a myriad of signaling chemical species. Among them, only three are diatomic or triatomic molecules: nitric oxide (NO), carbon monoxide (CO), and hydrogen sulfide (H₂S), the so-called “gaseous signaling molecules, or gasotransmitters” whose production and metabolism are primarily enzymatically regulated. These small molecules freely diffuse through cell membranes to elicit various responses independently of transporters or membrane receptors or second messenger systems (216), and they modulate many cellular functions through an array of intracellular signaling processes.

An abundance of recent experimental evidence suggests that H₂S plays a prominent role in normal physiology and pathophysiology, and many therapeutic targets exist for H₂S therapy (Fig. 1). The molecular targets of H₂S include proteins, enzymes, transcription factors, and membrane ion channels. Cysteine is the major source of H₂S in mammals, catalyzed by the enzymes: cystathionine beta-synthase (CBS), cystathionine gamma-lyase (CSE), and 3-mercaptopyruvate sulfurtransferase (3-MST) (Fig. 2). Whereas 3-MST is mainly localized in mitochondria, CBS and CSE exist in the cytosol.

CBS and CSE generate H₂S by using many different substrates (71). 3-MST catalyzes only sulfur transfer reactions from 3-mercaptopyruvate (3-MP) to various donors, for example:



The enzymatic sulfur transfer yields a hydropersulfide, not H₂S (10). Release of H₂S requires a further redox reaction between RSSH and a biological thiol such as glutathione (GSH):



Recently Kimura *et al.* demonstrated that 3-MST depends on a biological dithiol-thioredoxin (Trx) or dihydrolipoic acid- for the production of H₂S from 3-MP (128).

H₂S is enzymatically generated in the vasculature, heart, liver, kidney, brain, nervous system, lung, airway tissues, upper and lower GI tract, reproductive organs, skeletal muscle, pancreas, synovial joints, connective tissues, cochlea, and adipose tissues (105, 112). The key role of H₂S in health

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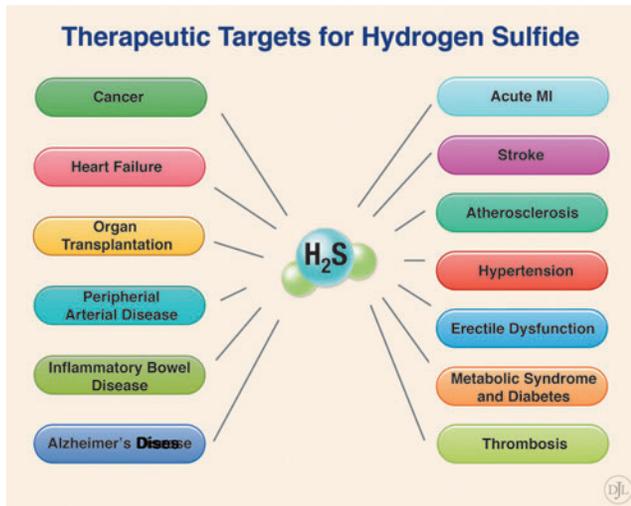


FIG. 1. Therapeutic targets for hydrogen sulfide (H_2S). An abundance of experimental evidence suggests that H_2S plays a prominent role in normal physiology and pathophysiology. Therefore, many therapeutic targets exist for H_2S therapy, including cancer, heart failure, organ transplant, peripheral artery disease, inflammatory bowel disease, Alzheimer's disease, acute myocardial infarction (MI), stroke, atherosclerosis, hypertension, erectile dysfunction, metabolic syndrome, diabetes, and thrombosis.

and disease is clearly borne out by the correlations found to exist between low levels of plasma/tissue endogenous H_2S /sulfane sulfur and/or H_2S -generating enzymes on the one hand, and on the other the presence and progression of adiposity, marked endothelial dysfunction/insulin resis-

tance, hypertension, hyperhomocysteinemia, diabetes, exacerbated cardiac injury following ischemia-reperfusion injury, Alzheimer disease, cirrhosis, chronic kidney disease, GI tract irritation, asthma, wound healing, and cancer (19, 46, 57, 58, 62, 85, 109, 156, 159, 210, 211, 219, 221, 239).

Physiological Actions of Hydrogen Sulfide

Nutrition, metabolism, and homeostasis

The main dietary sources of sulfur compounds in human nutrition are inorganic sulfates in drinking water and proteins derived from plants and animals. Only two of the twenty amino acids normally present in proteins are sulfur-containing amino acids (SAAs), namely methionine and cysteine. Methionine cannot be synthesized by the human body and must be supplied by the diet, whereas cysteine requirements can, in principle, be met by an excess of dietary methionine. However, cysteine is known as a semi-essential amino acid because humans can synthesize it from methionine to a limited extent (51, 143). Furthermore, the enzymes required for conversion of methionine to cysteine decline with age (17, 21). Dietary excess of cysteine and methionine is stored as GSH (17) (a thiolic antioxidant tripeptide) or, once the GSH pool has been replenished, converted to taurine or oxidized to sulfate (169). In fact, the availability of cysteine appears to be the rate-limiting factor for GSH biosynthesis from glutamate, glycine, and cysteine (9).

The "sulfane sulfur" pool (Fig. 7) performs an essential function in the brain, upon neuron excitation the bound sulfane sulfur releases H_2S (80, 194). It is highly likely that H_2S formation from sulfane sulfur requires reduced GSH as both hydrogen and electron donor. In the brain, H_2S is produced mainly in astrocytes, which contain larger amounts of GSH than neurons.

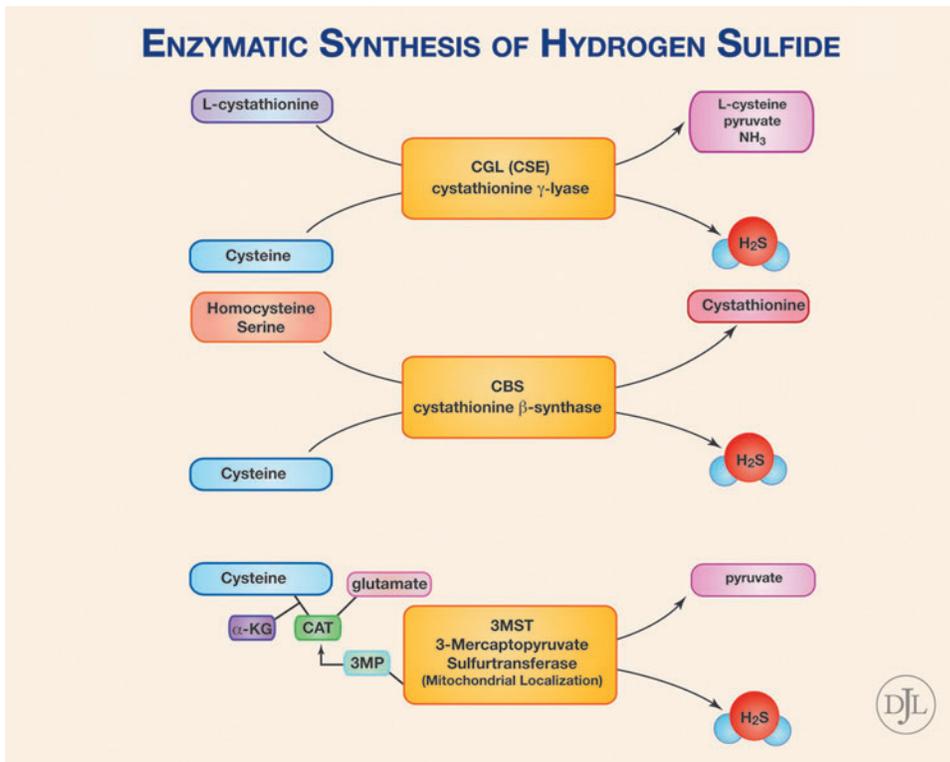


FIG. 2. Enzymatic synthesis of hydrogen sulfide (H_2S). Desulfhydration of cysteine is the major source of H_2S in mammals and is catalyzed by the trans-sulfuration pathway enzymes cystathionine beta-synthase (CBS), cystathionine gamma-lyase (CSE), and 3-mercaptopyruvate sulfurtransferase (3-MST). Cystathionine can be converted by CSE to form H_2S . CBS can form cystathionine from serine and homocysteine, and additionally can form H_2S from cysteine. Cysteine, along with alpha-ketoglutarate (alpha-KG), is converted to 3-mercaptopyruvate (3MP) by cysteine aminotransferase (CAT). 3MP can then be broken down by 3MST to form H_2S .

There is compelling evidence that the reversible formation of mixed disulfides between GSH and low-pKa cysteinyl residues of proteins (*e.g.*, S-glutathionylation) is an important mechanism for dynamic, post-translational regulation of a significant number of regulatory, structural, and metabolic proteins, and signaling pathways (9, 53, 97, 114). Mitochondrial GSH has been shown to act as a "sulfide buffer" when H₂S starts to build up in the cell (208). In mice and humans, ethylmalonic encephalopathy (EE) responds well to treatment with high doses of N-acetylcysteine (NAC, a cysteine/GSH prodrug) (208). This disorder is caused by mutations in *ETHE1*, a mitochondrial matrix sulfur dioxygenase involved in oxidative sulfide catabolism. Mitochondrial GSH can accept the sulfur atom of H₂S through the action of sulfide-CoQ reductase, yielding GSH persulfide (GSSH), which is better tolerated by the cell than H₂S (70, 71). Importantly, thiosulfate is excreted in massive amounts in the urine of mice and humans presenting with EE, with high thiosulfate and H₂S concentrations present in mouse tissues (200). In the five children treated by Viscomi *et al*, serum thiosulfate concentrations consistently decreased during treatment (208).

In spite of the critical role of sulfur in our diet, and especially of an adequate cysteine intake, dietary consumption of cysteine is generally suboptimal (51, 143). On the other hand, homeostatic regulation of cysteine and GSH pools declines with age, with the onset appearing in men at a younger age than in women (17). Since high dietary intakes of methionine have been shown to raise plasma levels of homocysteine (190), despite adequate intake of B vitamins, and since free cysteine can be a prooxidant (8, 75, 167), cysteine supplementation is nowadays achieved by oral administration of NAC, L-2-oxothiazolidine-4-carboxylate (OTC, another cysteine-GSH prodrug) or IMMUNOCAL (an undenatured protein concentrate rich in SAAs) (8). High-dose oral NAC has been shown to counter the intertwined redox and inflammatory imbalances in cystic fibrosis (201), and in several clinical trials, cysteine supplementation improved skeletal muscle function, decreased the body fat/lean body mass ratio, decreased plasma levels of pro-inflammatory cytokines NF- κ B and TNF- α , improved immune function, and increased albumin levels (9, 20, 53, 103, 122, 217). However, Palmer *et al*. found that oral administration of NAC to mice (10 mg/ml in drinking water) daily for 3 weeks led to development of pulmonary arterial hypertension that mimicked the effects of chronic hypoxia (150). These findings raise the concern that chronic NAC therapy might have similar consequences in patients (124).

The H₂S-cysteine-GSH connection has been documented often in the biomedical literature (26, 59, 93, 94, 141, 147, 158). Five factors are currently considered to contribute to the H₂S-stimulated increase in intracellular GSH levels: (i) enhancement of cellular glutamate uptake (194), (ii) a H₂S-induced increase in the level of gamma-glutamylcysteine synthetase and cystine transporter activity in the cell (94) (iii), reduction of cystine into cysteine by H₂S in the extracellular space, and transport of cysteine into cells by the cysteine transporter (93), (iv) H₂S stimulation of nuclear transcription factor Nrf2, which in turn upregulates GSH synthesis and transport (9, 26), and (v) a decrease in the activity of GSH-catabolizing enzymes (184). We believe the H₂S-cysteine-GSH connection to be strongly dependent on the fact that H₂S and L-serine act as co-substrates of cystathionine for CBS to yield L-cysteine (99, 153). This reaction is widely acknowledged to proceed in

the opposite direction, producing H₂S from cysteine, but its ready reversibility is firmly established (76, 153).

In summary, GSH is the most important intracellular thiolic antioxidant, a major determinant of the thiol/disulfide redox state, and a critical regulator of immune function, cell senescence, apoptosis, and vital redox-sensitive signaling pathways. Adequate levels of GSH are essential for effecting detoxification of xenobiotics and endogenously-generated toxins, for the biosynthesis of many essential biomolecules, and for protecting all cells from oxidative stress. Through the H₂S-cysteine-GSH connection, an H₂S prodrug may function not only as a source of H₂S but also as precursor of L-cysteine and GSH.

Inflammation and immunity

H₂S regulates inflammation and cell death, possibly exerting its beneficial effects through action on ATP-sensitive K⁺ channels (K_{ATP}) (196), inhibition of activation of NF- κ B and p38 MAPK, scavenging of oxidants, upregulation of intracellular cAMP, and inhibition of caspase-3 cleavage (212). Chronic inflammation is involved in some of the most common human diseases such as rheumatoid arthritis, tuberculosis, asthma, inflammatory bowel disease, vasculitis, and Crohn's disease. Chronic inflammation is an influential factor in type II diabetes, cardiovascular disease, and tumor development (1, 107, 133, 245). Infiltration of macrophages into the cellular mass is a common characteristic of atherosclerotic lesions and tumors. Since Virchow first showed that the inflammatory process influences atherosclerosis and tumor development, a growing body of evidence supports the hypothesis that macrophages play an important role in initiating and promoting both pathologies. In both cases, the combined effects of reactive oxygen species (ROS), cytokines, chemokines, and angiogenic factors, produced by tumor-associated macrophages and other inflammatory cells, explain the abnormal growth of cells: once a cellular mass becomes infiltrated by macrophages, the ability of tumor and atherosclerotic tissue to survive the immune response increases exponentially (163).

Ischemia-reperfusion (I/R) injury is regarded as a form of acute inflammation in which leukocytes play a key role. Experimental studies carried out during the last 20 years contributed to develop the concept that oxidant-induced leukocyte-endothelial tissue interactions are largely responsible for the microvascular dysfunction induced by reperfusion. Recognition of the vital role of the inflammatory process in I/R injury has provided the impetus for an intensive research effort aimed at preventing leukocyte infiltration into post-ischemic tissue (110, 242).

In atherosclerosis, monocyte adhesion to endothelial cells is stimulated by an oxidized cysteine/cystine redox status. The specific mechanism involves intracellular generation of hydrogen peroxide, activation of NF- κ B, and transcriptional activation and increased cell surface expression of cell adhesion molecules (CAM's) (83). H₂S is an extremely potent inhibitor of leukocyte adherence to the vascular endothelium (243). H₂S might interfere with inflammatory processes by diminishing the tissue injury induced by neutrophils via induction of apoptosis and/or scavenging of neutrophil-derived HOCl (220). Importantly, H₂S exerts opposite effects on the viability of lymphocytes and granulocytes, which is

probably the reason for the potentiation of the acute inflammatory and bactericidal responses and the depotentiation of the chronic inflammatory cellular response (243).

ROS/reactive nitrogen species (RNS) are mediators of NF- κ B activation and this process can be blocked by antioxidants, in particular, cysteine and GSH (83). H₂S has been shown to downregulate several pro-inflammatory cytokines including NF- κ B, TNF- α , IL-1 β , IL-6, and IL-8 (55, 98, 144, 151), to modulate leukocyte adhesion and leukocyte-mediated inflammation (55,181), to mediate the cardioprotection induced by ischemic postconditioning (241) and to protect from NF- κ B and TNF- α mediated endotoxic shock (113). The powerful reducing/antioxidant/free radical scavenging properties of H₂S can explain its wide-ranging anti-inflammatory and cytoprotective effects, including protection against: ischemia-reperfusion injury in heart, brain, retina, liver, and intestine; endothelial dysfunction; hydrogen peroxide-induced damage in rat gastric epithelial cells; hyperhomocysteinemia in rats; methionine- and homocysteine-induced oxidative stress; and hemin-mediated oxidation of low-density lipoprotein (112).

Cytoprotection and pharmacological conditioning

Cardiovascular system. H₂S strongly influences the body's redox status through various mechanisms, such as increasing GSH levels in the cytosol, mitochondria, and nucleus of cells, increasing the GSH/GSSG ratio, activating the reperfusion injury salvage kinase (RISK) pathway with up-regulation of protective heat-shock proteins, and acting as "master switch" of Nrf2 nuclear translocation, resulting in persistent activation of the antioxidant responsive elements (AREs) of antioxidant genes and concomitant overexpression of antioxidant and phase II enzymes (151, 165). H₂S not only exerts anti-apoptotic and anti-inflammatory effects but also anti-nociceptive and blood pressure-lowering effects by activating K_{ATP} channels (196). The cardioprotective effect of H₂S also involves activation of cardiac extracellular signal-regulated-kinase and/or Akt pathways (196).

Evidence on the cardioprotective effects of H₂S has been obtained by many researchers. It has been shown that H₂S has profound protective effects on the heart in murine models and that genetic overexpression of CSE in the heart is highly protective from I/R injury (26, 55). Exogenous administration of H₂S and its donors in the settings of atherosclerosis, myocardial I/R injury, chronic heart failure, and cardiopulmonary resuscitation shows significantly improved outcomes in small animal models (25, 26, 119, 131, 156). These results are being translated into large animal models (155, 179, 181). The observed protection is associated with improved heart mechanics, reduced myocardial inflammation, preserved mitochondrial function, Nrf2 activation, and reduced cardiomyocyte apoptosis (25, 26, 55, 131, 156).

Yusof *et al.* reported the first evidence that preconditioning by exposing the small bowel of rats to NaHS induces an anti-inflammatory phenotype, such that postcapillary venules fail to support leukocyte rolling and adhesion when subjected to I/R injury 24 hours later (242). I/R injury is a major source of morbidity and mortality, not only in myocardial infarction, but also in many other clinical settings, including solid organ transplantation and ischemic cerebral and retinal vascular episodes. It is also a cause of irreversible damage to skeletal muscle made ischemic either as the result of pathologic hy-

poperefusion or of a planned surgical intervention. On the basis of results of both *in vitro* and *in vivo* experiments, it was recently concluded that the preischemic or postischemic delivery of NaHS limits I/R-induced cellular damage and confers significant long-term protection, that intravenous or even intra-arterial delivery of an H₂S donor would provide more focused treatment of target tissue and, when administered in appropriate doses and within the proper time frame, H₂S holds significant promise as a cytoprotective agent (65).

Peripheral arterial disease (PAD) affects over 5% of the older population (>60 years). PAD is considered a marker for systemic atherosclerosis and is frequently complicated by coronary and cerebral events (116). In PAD, oxidative stress is implicated in the correlation of a reduction in flow-mediated dilation (FMD) with a higher risk of developing CV complications. Therefore, treatment with antioxidants, aimed at improving peripheral arterial dilatation, is being investigated (116). In a rat unilateral hind limb ischemic model, treatment with NaHS (50 μ mol kg⁻¹ day⁻¹) promoted significant angiogenesis and improved regional blood flow. These effects are associated with an increase in vascular endothelial growth factor (VEGF) expression in skeletal muscle and VEGF receptor 2 (VEGFR2) phosphorylation in neighboring vascular endothelial cells. In addition, Akt phosphorylation is increased in ischemic muscles following NaHS treatment. However, treatment with 200 μ mol kg⁻¹ day⁻¹ has no angiogenic effect (215).

Angiogenesis is triggered when the effects of pro-angiogenic factors, such as hypoxia inducible factor (HIF) and tumor growth factor (TGF), present in the tissue overcome those of the anti-angiogenic factors. It is possible that, at the higher dose, H₂S/HS⁻ inhibits nicotinamide adenine dinucleotide phosphate (NADPH) oxidase (27) and/or binds to multiple cellular targets evoking mechanisms that counteract the pro-angiogenic effects (24, 215). In 2007 Isenberg *et al.* presented the results of a study on modulation of angiogenesis by certain simple dithiolethiones (DTTs), certain dithiolethione-modified nonsteroidal anti-inflammatory drugs (S-NSAIDs) and valproic acid, and H₂S (79). Simple DTTs, S-NSAIDs and S-valproate demonstrated significant anti-angiogenic activities, inhibiting endothelial cell proliferation and vascular cell outgrowth and invasion of extracellular matrix. H₂S, on the other hand, dose-dependently inhibited vascular cell outgrowth (at concentrations between 0.1 and 1000 μ M) while stimulating endothelial cell proliferation in a dose-dependent manner within the same concentration range. Importantly, vascular outgrowth from muscle tissue was completely abrogated by H₂S at a concentration of 0.01 μ M, whereas endothelial cell proliferation increased by a factor of less than two between 0.1 and 1000 μ M (79). According to Sparatore *et al.*, H₂S-donating hybrids-containing a DTT moiety inhibit angiogenesis and cell proliferation, these effects being related to their ability to slowly and gradually release H₂S (184).

Nervous system. 1-Methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) is a neurotoxin that can induce Parkinson's disease (PD)-like symptoms and biochemical changes in animals and humans. Inhaled H₂S has been shown by Kida *et al.* to prevent MPTP-induced movement disorder, neuron degeneration, and neuron apoptosis and gliosis in mice (91). These effects were attributed to upregulation of genes encoding anti-inflammatory and antioxidant proteins, including

heme oxygenase-1 (HO-1) and glutamate-cysteine ligase. Levodopa (L-DOPA) is widely used in PD therapy, but it does not prevent loss of substantia nigral dopaminergic neurons. The main factors responsible for this loss are oxidative stress and inflammation, which can be controlled by L-3,4-dihydroxyphenylalanine (L-DOPA) derivatives capable of being converted *in vivo* into L-DOPA and H₂S by chemical and/or enzymatic means such as ACS83, ACS84, ACS85, and ACS86 (Fig. 3).

The four molecules in Figure 3 were synthesized and studied by Sparatore *et al.* (106). ACS83 and ACS84 are [1,2]-dithiole-3-thione derivatives, ACS85 is a [1,3]-dithiole-2-thione derivative, and ACS86 is a disulfide containing an allylmercapto moiety, which is expected to release H₂S upon nonenzymatic reduction by GSH. ACS84 was converted by isolated mitochondria into H₂S. This conversion was also observed *in vivo*, with a large increase in intracerebral dopamine (30% more than with L-DOPA) and GSH after intravenous administration to rats. The four L-DOPA hybrids reduce release of TNF- α , IL-6,

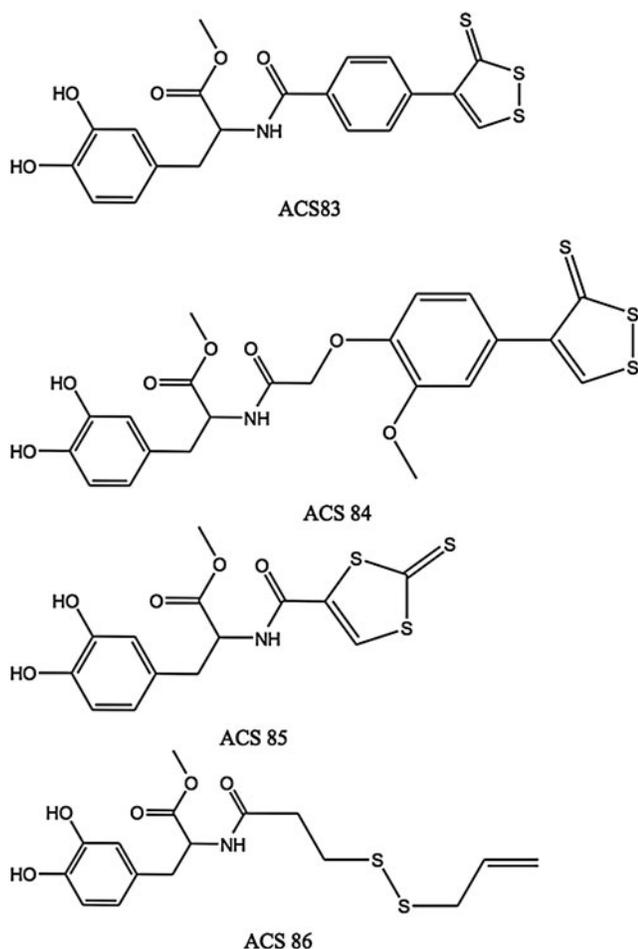


FIG. 3. The L-DOPA derivatives capable of being converted *in vivo* into L-DOPA and H₂S by chemical and/or enzymatic means. ACS83, ACS84, ACS85, and ACS86 were synthesized and studied by Sparatore *et al.* (106). ACS83 and ACS84 are [1,2]-dithiole-3-thione derivatives, ACS85 is a [1,3]-dithiole-2-thione derivative, and ACS86 is a disulfide containing an allylmercapto moiety, which is expected to release H₂S upon nonenzymatic reduction by GSH.

and NO from stimulated microglia and astrocytes. They proved superior to L-DOPA itself as neuroprotectants.

Emerging evidence suggests that H₂S may have therapeutic potential in Alzheimer's patients since it reduces mRNA levels and protein levels of beta-site amyloid precursor protein-cleaving enzyme 1 in nerve growth factor differentiated PC12 cells (105, 248). The depletion of H₂S in the brains of Alzheimer's patients may be due to high levels of myeloperoxidase. Abnormally low brain levels of endogenous H₂S and inflammatory stress are hallmarks of Alzheimer's (219).

Total GSH was shown to be substantially lowered in mitochondria from severely ischemic rat brain tissue (3). Hideo Kimura and colleagues recently showed that exogenous H₂S increases GSH production and suppresses oxidative stress in isolated rat mitochondria (93). These *in vitro* findings were mirrored by *in vivo* observations that H₂S protects ischemic brain by reinstating GSH levels decreased by oxidative stress. H₂S has been shown to protect neurons against hypoxic injury by upregulating expression of heat shock protein (HSP) 90 (127, 199). HSP90 is a ubiquitous molecule that contributes to cell survival by regulating the folding of various cellular proteins, including survival factors, and by binding to apoptotic protease activating factor-1 (Apaf-1), thereby preventing apoptosis.

The slow-releasing H₂S prodrug ACS67 (a latanoprost-dithiolethione conjugate) was shown to attenuate retinal ischemic damage following experimental elevation of retinal pressure in rats, with ACS67 being more potent than latanoprost (147). The same authors found that ACS67 significantly attenuated hydrogen peroxide-induced damage to transformed neural precursor cells known to exhibit a number of characteristics associated with retinal ganglion cells (RGC). It is pertinent to point out that, according to Osborne *et al.* (147), the neuroprotective effect of ACS67 probably involves several mechanisms, prominently including stimulation of GSH formation. Biermann *et al.* (14) recently demonstrated in rats that preconditioning with inhaled H₂S (80 ppm in air) significantly attenuates apoptosis of RGCs after retinal ischemia/reperfusion injury. Their results revealed that H₂S is able to attenuate caspase-3 cleavage and caspase-3 activity and significantly upregulated induction of cytoprotective chaperone HSP90, and strongly suggest that NF- κ B downregulation is one component of this neuroprotective action. Furthermore, Mikami *et al.* demonstrated that H₂S protects the retina from light-induced damage by regulation of intracellular calcium via activation of vacuolar type H⁺-ATPase (129).

Elevated levels of homocysteine (Hcy) in the blood (hyperhomocysteinemia), may cause mental retardation, seizures, and Alzheimer disease (205) via Hcy-induced oxidative stress and increased cerebrovascular permeability. Tyagi *et al.* (205) suggest that H₂S functions as a type of armor in the brain, and could be a beneficial therapeutic candidate for the treatment of hyperhomocysteinemia-associated pathologies, such as stroke and neurologic disorders.

Digestive system. Exogenous administration of H₂S prevents ethanol-induced gastric damage in mice and has a protective role against oxidative stress in rat gastric mucosal epithelium (126, 240), and is effective at preventing damage to the gastric mucosa induced by nonsteroidal anti-inflammatory drugs (NSAIDs) (58) and at promoting resolution of colitis in rats (213). Fiorucci *et al.* point out that, in addition to the firmly established contribution of exogenous

H₂S to gastric mucosal protection, its role in accelerating repair of mucosal injury might soon emerge. The therapeutic dose range of NaSH/Na₂S was found to be very narrow (240). Takeuchi *et al.* present evidence supporting the assumption that endogenous H₂S is involved in regulation of acid-induced bicarbonate ion secretion and mucosal protection in the duodenum (193).

H₂S, as well as precursors containing a dithiolethione moiety, are potent inducers of the antioxidant and cytoprotective enzyme HO-1. This is clinically significant because HO-1 promotes ulcer healing (183). Other mechanisms believed to contribute to the GI-protective effect of H₂S are increased epithelial secretion and mucosal blood flow, activation of K_{ATP} channels and of capsaicin-sensitive afferent nerves, reduction of leukocyte adhesion/infiltration, downregulation of TNF- α /IL-1 β /IFN-gamma expression and scavenging of oxidants (212).

Liver and kidneys. Mounting evidence suggests that H₂S regulates intrahepatic blood flow (microcirculation) in the normal and cirrhotic liver (58), with insufficient production of H₂S in the cirrhotic liver and downregulation of H₂S-producing enzymes in kidney and liver of patients with chronic kidney disease (2). Administration of H₂S donors has been found to protect the liver and kidneys from ischemia-reperfusion damage (82, 115). In kidneys, H₂S has been found to be beneficial to the prevention or treatment of diabetic kidney disease via alleviating renal glycolytic injury (115), increasing renal blood flow, glomerular filtration rate, and urinary sodium excretion (230), and ameliorating hyperhomocysteinemia-associated chronic renal failure (173). In the liver, H₂S effectively attenuates stress-mediated liver injury and hepatic mitochondrial dysfunction in acutely ethanol-exposed mice (244), and markedly alleviates acetaminophen-induced hepatotoxicity in mice (135). The hepatoprotective and nephroprotective effects of H₂S are mostly mediated by the "Nrf2 regulon", (*i.e.*, by activation of the many cytoprotective and lipogenesis-regulating genes controlled by the Nrf2-ARE pathway) (95,96,174). Additionally, H₂S is also of benefit in hyperlipidemic and/or hypercholesterolemic prevention and therapy (75) via both enzymatic and nonenzymatic activities.

Diabetes and metabolism

Diabetes mellitus and its CV complications have been associated with increased production of ROS and perturbations of thiol redox homeostasis. Increased oxidative stress and oxidative damage are considered mediators of vascular injury in CV pathologies, including hypertension and atherosclerosis. In fact, CV disease is the major cause of morbidity and mortality for diabetic individuals. In order to reduce these risks, it is necessary to develop therapies aimed simultaneously at improving energy metabolism, insulin resistance, vascular function, blood pressure, and inflammatory/procoagulant status (125, 161).

The rate of ROS production depends on the metabolic status of the cell, as hyperglycemia increases the steady-state superoxide concentration. The rate of enzymatic reduction of glucose to sorbitol increases as well, with concomitant decreases in NADPH and GSH concentration. This depletion of reducing equivalents results in augmented sensitivity to oxidative stress (123). Thus, oxidative stress from excessive ROS

and depleted mitochondrial GSH (mtGSH) can lead to cardiomyocyte apoptosis in the diabetic heart. Similarly, in diabetic retinopathy, superoxide levels in retinal mitochondria of diabetic mice are twice as high as those in nondiabetic controls, and mtGSH levels in the same retinas undergo a 40% decrease due to hyperglycemia (123).

According to Niki and his colleagues, endogenous H₂S protects pancreatic β cells of mice from apoptosis induced by oxidative stress and/or glucotoxicity. They also found that NaHS was able to suppress ROS production induced by cytokines or hydrogen peroxide, via activation of Akt signaling (90, 197, 198). These findings are consistent with the effect of DATS (a hydrogen sulfide precursor, see below) on the level of blood sugar and oxidative stress markers in rats with type II diabetes mellitus (54).

It is now apparent that H₂S biosynthesis declines as the severity of diabetes increases over time, and that therapies based on administration of different H₂S donors to animals or patients in different stages of type I or type II diabetes may be highly successful (19, 46, 109, 221). Interestingly, it has been reported that plasma H₂S levels are reduced in overweight individuals, with increasing adiposity being a major determinant of said levels (221). On the other hand, emerging evidence points to diminished Nrf2/ARE activity as a major contributor to increased oxidative stress, disrupted lipogenesis, mitochondrial dysfunction in the vasculature leading to endothelial dysregulation, insulin resistance, and the abnormal angiogenesis observed in diabetes (37, 95, 174, 192, 195, 209). Taken together, the aforementioned findings suggested that, in diabetes, blunted H₂S biosynthesis is a major contributor to increased oxidative stress/mitochondrial and endothelial dysfunction and insulin resistance, the causal link being diminished Nrf2/ARE activity.

Last, recent evidence indicates that H₂S (or its donors) exerts an anti-atherogenic effect by counteracting the oxidation of low-density lipoprotein (LDL) via HOCl scavenging, H₂O₂ scavenging, myeloperoxidase inhibition, and inhibiting foam cell formation by downregulating CD36, SR-A (scavenger receptor A) and ACAT1 (acyl-coenzyme A:cholesterol acyltransferase-1) expression via the K_{ATP}/ERK1/2 pathway in human monocyte-derived macrophages (100, 250). Lynn and Austin have reviewed experiments demonstrating that H₂S supplementation ameliorates atherogenic processes, and therefore that such supplementation may be of therapeutic benefit in the prevention and treatment of atherosclerosis (119). For a full discussion of the relationship between H₂S and the metabolic syndrome, please refer to the recent review by Desai *et al.* (48).

Benavides *et al.* proposed that endogenous H₂S production from garlic-derived organic polysulfides provides the basis for the long-term beneficial effects obtained from the habitual consumption of garlic (13), in particular, the reduction in risk factors associated with the metabolic syndrome such as increased oxidative stress, obesity, hypertension, high blood glucose levels, hypercholesterolemia, hyperlipidemia, platelet aggregation, and blood coagulation, that together greatly increase the risk of developing CV disease and type II diabetes (148). Benavides *et al.* stressed endogenous H₂S production from allyl, di-, and polysulfides derived from garlic (13), but did not mention the presence in garlic extracts of significant amounts of S-substituted L-cysteine derivatives (cysteine S-conjugates) which are also important H₂S precursors, such as S-allyl-L-cysteine, S-allylmercapto-L-cysteine, S-

propylmercapto-L-cysteine, and S-(penta-1,3-dienyl)mercapto-L-cysteine. These compounds are substrates of CBS (β -cystathionase, which also possesses beta-lyase activity). The mercapto-substituted derivatives are thereby converted into hydropersulfides (RSSH), which readily yield H_2S upon reduction by GSH (13, 152). Taken together, these findings suggest that most of the organosulfur compounds in garlic preparations are potential H_2S precursors in the body.

H₂S as an antioxidant and free radical scavenger

At 37°C and physiological fluid pH (pH 7.4), about 80% of the H_2S molecules dissociate to yield HS^- (hydrosulfide anion), which is therefore the predominant sulfur-containing species in extracellular fluids and plasma (49), whereas within the cell (pH about 7.2) the amounts of H_2S and HS^- are nearly equal (145). Hydrosulfide anions are powerful one-electron chemical reductants capable of quenching free radicals by hydrogen atom transfer or by single electron transfer usually at or near diffusion-controlled rates. Their reaction with dioxygen is fast when catalyzed by divalent metal ions. They are also strong nucleophiles as evidenced by their reaction with S-nitrosothiols to release NO (202). The oxidation of hydrosulfide anions by biochemically relevant two-electron oxidants (*e.g.*, hypochlorous acid and hydrogen peroxide) yields initially hydrogen disulfide (H_2S_2 , also known as disulfane) which is also a highly reactive oxidizing agent (139, 160) capable of regenerating H_2S by reaction with a thiol (13) or by disproportionation (118, 139). H_2S will readily scavenge ROS and RNS, including hypochlorous acid, hydrogen peroxide, lipid hydroperoxides, O_2^- and peroxynitrite (93, 139). It is also able to scavenge the triplet state of riboflavin (214). However, in the presence of molecular oxygen (dioxygen) autooxidation of H_2S generates free radicals (189).

Under oxidative stress conditions, H_2S may be converted to sulfite by activated neutrophils (132). Mitsuhashi *et al.* found that when NaHS was added *in vitro* to the supernatant of activated neutrophils, a significant amount of sulfite could be detected. Furthermore, a NADPH oxidase inhibitor markedly suppressed the production of sulfite. The chemical production of sulfite from H_2S by neutrophil oxidative bursts is associated with inflammation, which might be responsible for the high levels of serum sulfite found in patients with pneumonia (132).

Although seldom acknowledged, simple species containing an SH group such as H_2S , HS^- , HS-SH, and HSS $^-$ excel at undoing the damage inflicted to biomolecules by free radicals through hydrogen atom donation to carbon-centered radicals (Fig. 4) (157, 191). Although hydrogen atom transfer to carbon-centered radicals is a diffusion-controlled reaction, the extremely low concentrations of H_2S and H_2S_x in blood and tissues limit their efficiency at repairing free radical damage to biomolecules (145).

Typically, carbon-centered free radicals react with oxygen to yield alkylperoxyl radicals:



which react further by abstracting hydrogen atoms from other biomolecules:

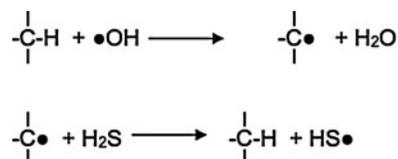


FIG. 4. Hydrogen atom donation to carbon-centered radicals. Simple species containing an SH group such as H_2S , HS^- , HS-SH, and HSS $^-$ excel at undoing the damage inflicted to biomolecules by free radicals through hydrogen atom donation to carbon-centered radicals (157, 191). Although hydrogen atom transfer to carbon-centered radicals is a diffusion-controlled reaction, the extremely low concentrations of H_2S and H_2S_x in blood and tissues limit their efficiency at repairing free radical damage to biomolecules.



An inability to repair oxidized DNA, lipids, and proteins contributes to the damage induced by oxidative stress. Examples of macromolecular repair include DNA repair by base or nucleotide excision, protein repair by thioredoxin and glutaredoxin (162), and lipid repair by GSH peroxidase.

In order to appreciate the importance and uniqueness of the role of H_2S as an antioxidant/free radical shield/cytoprotector, it is essential to recall that single antioxidants as pharmacologically active agents have not been found to exhibit extremely powerful therapeutic effects (177). This rather limited success might seem at first surprising in view of the decreased levels of selected major antioxidants consistently found in a number of disease states, but the limited success of this "single direct antioxidant approach" can be rationalized by recalling that mammals possess highly evolved and well-integrated antioxidant mechanisms that require the concerted and synergistic action of both antioxidant enzymes and low-molecular-weight antioxidants, with different antioxidants operating extracellularly and/or in specific cell compartments and having limited functional overlap: some destroy peroxidic species and/or peroxynitrite, others break free radical chains, and still others quench singlet oxygen (45). In addition, due to their short half-lives, direct antioxidants (vitamins C, E, etc.) must be administered frequently and at relatively high dosages to sustain their physiological efficacy (84). Furthermore, use of high-dose direct antioxidants may elicit pro-oxidant effects (45). However, H_2S is not just another antioxidant to be added to the list of "direct antioxidants", but it is also a powerful cytoprotective agent capable of activating nuclear transcription factor Nrf2 and consequently of inducing the expression of over 200 genes. These Nrf2-dependent genes encode proteins involved in lipid homeostasis, phase 2 detoxifying/antioxidant enzymes, directly acting antioxidant proteins, synthesis of low molecular weight antioxidants, and several P450 enzymes (84, 95, 174).

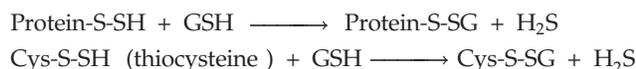
Cell signaling

The interaction of H_2S with nuclear transcription factors has been intensively scrutinized. Many researchers have

shown, using both cells in culture and whole animals that, in most cases, H₂S inhibits NF-κB (112, 222). Slow-releasing H₂S donors such as DATS, GYY4137, and S-diclofenac have also been shown to block NF-κB nuclear translocation in mouse macrophages and rat liver. Administration of GYY4137 to LPS-injected rats resulted in activation of signal transducer and activator of transcription-3 (STAT3), which is known to regulate the expression of many genes that mediate cell survival, proliferation, and angiogenesis (1, 113). H₂S administration induces activation of transcription factor Nrf2 (26). In the nematode, *Caenorhabditis elegans*, H₂S upregulates HIF-1 (23).

Many mechanisms of action of H₂S may be mediated by protein S-sulfhydration (138,172). Sen *et al.* recently showed that S-sulfhydration of NF-κB by H₂S is responsible for its anti-apoptotic actions (171). Mustafa *et al.* pioneered the concept of S-sulfhydration (SHY) as a signaling system (138). They define SHY as a physiological process wherein H₂S attaches an additional sulfur atom to the thiol (-SH) groups of cysteine (Cys) residues within proteins, yielding a hydroper-sulfide group (-SSH). SHY usually activates enzymes (138). S-sulfhydration of GAPDH, for instance, results in a 7-fold increase in catalytic activity (138). Among the 49 proteins that were found to be basally S-sulfhydrated by liver-generated H₂S are albumin, actin, β tubulin, CSE, CBS, several phosphatases, and catalase, and these authors estimate that from 10 to 25 percent of endogenous GAPDH, β tubulin, and actin are S-sulfhydrated *in vivo* (138).

Sulfane sulfur results following sulfhydration, and may also serve as a biological source of H₂S. Operationally, sulfane sulfur was defined by Wood in 1987 (224) as sulfur that reacts, at pH 8.5–10, with cyanide to yield thiocyanate (Fig. 5) (88, 89, 225). From a structural viewpoint, a sulfane sulfur atom in an electrically neutral molecule is always attached to another sulfur atom and is either in an oxidation state of zero, or in an oxidation state of –1, and is attached to a hydrogen atom or to an “activating group” such as allyl, benzyl, phenacyl, etc. The “outer” sulfur atom of a hydroper-sulfide group is highly redox-labile, and is readily converted into H₂S by reducing agents such as dithiothreitol, cysteine, or GSH:



“Activated organic disulfides” such as those shown in Figure 6 are organic sulfane sulfur compounds (12). The molecules of

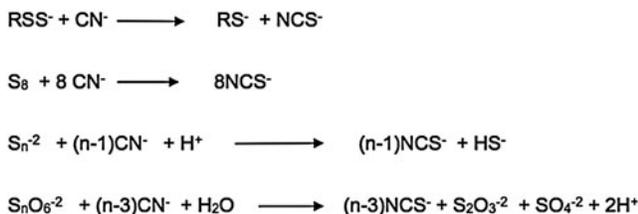
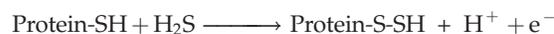


FIG. 5. Sulfane sulfur. Sulfane sulfur was defined by Wood in 1987 (224) as sulfur that reacts, at pH 8.5–10, with cyanide to yield thiocyanate (88, 89, 225). From a structural viewpoint, a sulfane sulfur atom in an electrically neutral molecule is always attached to another sulfur atom and is either in an oxidation state of zero or in an oxidation state of –1 and is attached to a hydrogen atom or to an “activating group” such as allyl, benzyl, phenacyl, etc.

organic hydroper-sulfides, and hydro-polysulfides contain the –S-S-H moiety, and therefore also contain sulfane sulfur. The most important hydroper-sulfides in biology are probably thiocysteine (Cys-SSH) and GSH hydroper-sulfide (G-SSH). “Bound sulfur” was defined by Ogasawara *et al.* as “divalent sulfur that is easily liberated as sulfide by reduction with dithiothreitol” Therefore, the “sulfane sulfur pool” constitutes a major portion of the labile sulfur pool (Fig. 7) present in tissues of plants and animals.

Acid-labile sulfur comprises various metalloproteins, which contain sulfide ions as part of metal/sulfur clusters (mainly Fe/S and Zn/S clusters) (81). Acidification may liberate the S²⁻ ions, which are released as SH⁻ and H₂S. The brain, heart, and liver contain significant amounts of acid-labile sulfur, whereas lung and muscle contain less (87). The labile sulfur pool (206) comprises both inorganic and organic chemical species, the simplest being disulfane (HS-SH), which is present as HS₂⁻ at physiological pH. In this context, it is important to bear in mind that, since sulfane sulfur atoms are in the zero or minus one oxidation state, they must gain electrons (*i.e.*, be reduced) in order to generate S²⁻ or HS⁻.

The conversion of a thiol into a hydroper-sulfide by H₂S requires one equivalent of an oxidant (81, 86, 139):



Although Nagy and Winterbourn recently proposed hypochlorous acid as a candidate (139), we believe hydrogen peroxide and the superoxide radical anion to be much more widely available oxidants in living tissues. Three likely mechanisms for S-sulfhydration are shown in Figure 8.

At physiological pH, most cysteine thiol groups in proteins are protonated (-SH) and hence display low reactivity towards H₂O₂. However, in some proteins where the cysteine residue is flanked by basic amino acids, the cysteine-SH group exists as the highly oxidizable thiolate anion (-S⁻). This introduces an element of specificity in H₂O₂-mediated signaling, suggesting that mainly proteins containing low pKa cysteine residues undergo S-sulfhydration. Hydrogen peroxide is the physiological oxidant of choice because it is constitutively produced inside most cells at various loci such as mitochondria, peroxisomes, and the cytosol mainly via enzymatic processes mediated by SOD, NADPH oxidases, xanthine oxidases, sulfhydryl oxidases, thiol oxidases, and monoamine oxidases (27, 83), and the reactivity of hydrogen peroxide toward thiols and H₂S is high (140). Hydrogen peroxide generation in mammals is probably in the vicinity of 50 μmol kg⁻¹min⁻¹ (83).

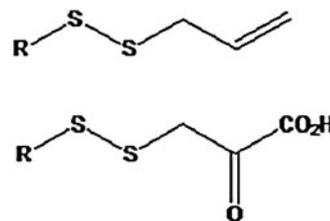
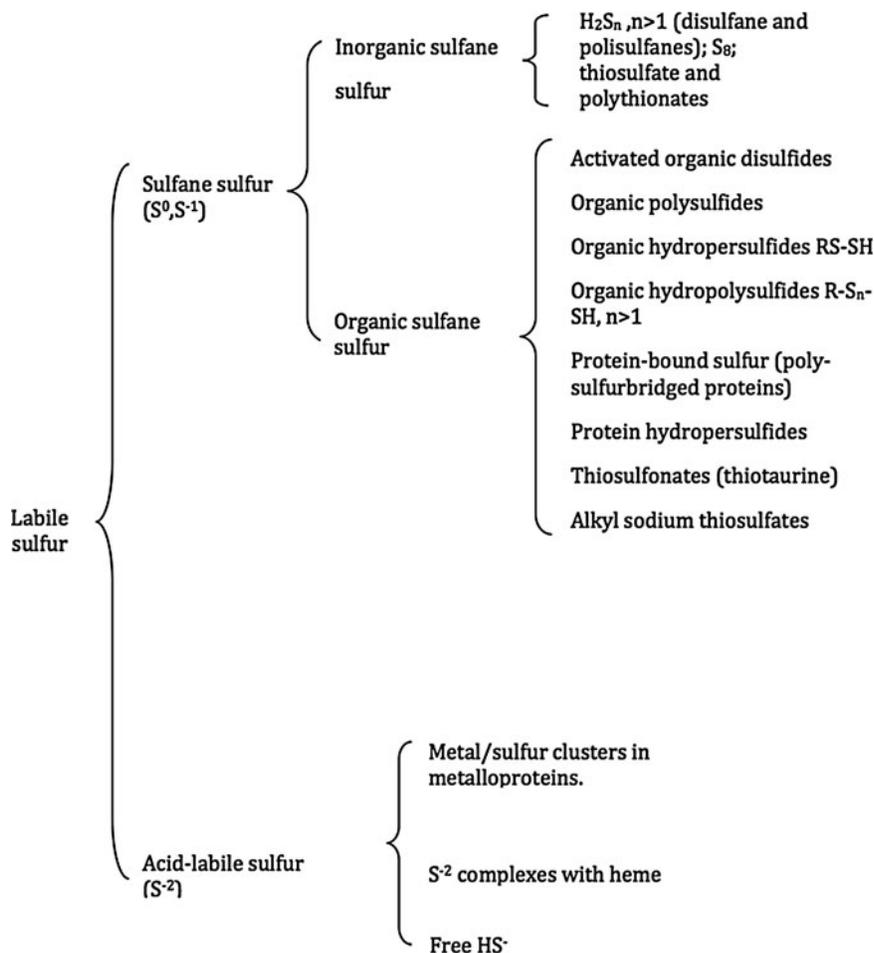
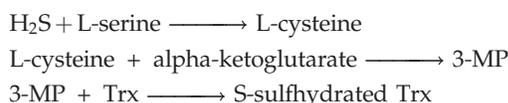


FIG. 6. “Activated organic disulfides.” These compounds contain “cyanolyzable sulfur” (12) and therefore belong in the category of organic sulfane sulfur compounds.

FIG. 7. Labile sulfur. Labile sulfur can be broken down into acid-labile sulfur, consisting of metal/sulfur clusters in metalloproteins, or heme complexes, as well as free HS⁻, or sulfane sulfur. Sulfane sulfur can be inorganic or organic in origin.



S-sulfhydration of an enzyme may be accomplished through interaction with the proper substrate and does not require a discrete oxidation step involving a thiol group at the active site (as in mechanisms A, B or C, Fig. 8). Thus, 3-mercaptopyruvate has been reported to react with Trx, yielding pyruvate and Trx hydropersulfide (218). Therefore, H₂S might S-sulfhydrate Trx through the following enzymatic pathway:



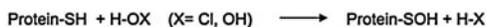
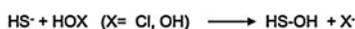
As efficient mitochondrial pathways for H₂S oxidation are available (71), steady state tissue concentration can be held at very low levels and it is possible for H₂S to function as an oxygen sensor (146). Thus, under hypoxic conditions, H₂S catabolism would be blocked, leading to increased H₂S levels with activation of specific responses (146). This hypothesis is consistent with similarities between the effects of hypoxia and H₂S, enhancement of hypoxic signaling by H₂S precursors, and abolishment of hypoxic signaling by H₂S synthesis inhibitors.

Sexual function

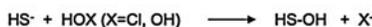
Moore and his co-workers (185—187) have described some pioneering studies that provide evidence for the endogenous formation of H₂S and its pro-erectile relaxant effect on the

corpus cavernosum of mammals, as well as on the effects of H₂S in female sexual function. The first set of results were corroborated in a recently published article (40). There is also evidence that oxidative stress is implicated in erectile dysfunction (ED) in diabetic rodents (15,) and that interventions based on administration of tetrahydrobiopterin (182) and upregulation of antioxidant enzymes may be useful (44). For a discussion of the roles of endogenous and exogenous H₂S in the endocrine and reproductive systems and the possibility of developing new therapies for ED that target this pathway, please see the recent articles by Émanuelle Di Villa Bianca *et al.* (41) and Zhu *et al.* (252).

Sparatore *et al.* have developed an H₂S-donating derivative of sildenafil (ACS6) with possible clinical indications in ED, benign prostatic hypertrophy, and low urinary tract symptoms (184). ACS6 is a hybrid obtained by esterification between a phenolic dithiolethione and a carboxylic acid derived from sildenafil by attachment of a carboxyl moiety (CO₂H) to the N-methyl group joined to the piperazine ring. The H₂S released by S-sildenafil (ACS6) inhibits both PDE5 and NOX expression and activity. Furthermore, H₂S applied *ex vivo* or overexpression of CSE has been shown to increase cGMP levels by phosphodiesterase inhibition in aortic ring preparations (22). Hence, this mechanism may constitute the basis of a new and effective approach to the treatment of patients suffering from ED, benign prostatic hypertrophy, and lower urinary tract symptoms.

Mechanism A.**Mechanism B.**

Please note that HS-OH may be thought of as the simplest sulfenic acid.

Mechanism C.

Please note that HSS⁻ (an inorganic species containing sulfane sulfur) acts here as an oxidant.

FIG. 8. Three likely mechanisms for S-sulfhydration. At physiological pH, most cysteine thiol groups in proteins are protonated (-SH) and hence display low reactivity towards H₂O₂. However, in some proteins where the cysteine residue is flanked by basic amino acids, the cysteine-SH group exists as the highly oxidizable thiolate anion (-S⁻). This introduces an element of specificity in H₂O₂-mediated signaling, suggesting that mainly proteins containing low pKa cysteine residues undergo S-sulfhydration.

In fact, ACS6 and sildenafil citrate relaxed cavernosal smooth muscle equipotently and ACS6 inhibited superoxide formation more than sildenafil citrate (175). Shukla *et al.* concluded that ACS6 not only promotes erection, but also affords effective protection from oxidative stress through upregulation of GSH synthesis. Additionally, in an investigation of the effect of NaHS on pregnant rat uterine contractility *in vitro*, Sidhu *et al.* found that this “hydrogen sulfide donor” produced significant dose-dependent decreases in uterine spontaneous contractility (176).

Life span modulation

Many lines of evidence suggest that oxidative stress plays an important role in aging. In *C. elegans* and *Drosophila melanogaster*, mutations resulting in resistance to toxic stresses, oxidative or not, tend to result in increases in longevity. In *C. elegans*, recent studies have shown that the Nrf2 homologue, SKN-1 (121), is necessary for the life span extension seen with dietary restriction, and overexpression of SKN-1 can increase life span. In *D. melanogaster*, increased Nrf2 activity correlates with oxidative stress resistance and increased life span of male flies (192). In mice, decreased Nrf2 signaling with age, and increased Nrf2 signaling with caloric restriction have been observed (111). H₂S augments the life span of *C. elegans* through a sirtuin, a process that may involve protein S-sulfhydration (130). Since sirtuins are also found in vertebrates and since H₂S signaling pathways are highly conserved, it is possible that this effect/mechanism might be found in mammals as well. According to Powolny

et al., treatment of the worm *C. elegans* with DATS increases its mean lifespan, even if the treatment is initiated during young adulthood (154). Since DATS readily yields H₂S *in vivo*, we consider it likely that this effect of DATS is mediated by H₂S.

Leiser and Miller describe a series of studies that lend support to the hypothesis that augmented Nrf2 activity contributes to several forms of stress resistance observed in long-lived Snell dwarf mice that live about 40% longer than littermate controls and show delays in the onset of many aging-related pathologies (111). Importantly, Dwarf-derived fibroblasts exhibit many of the traits associated with enhanced Nrf2/ARE activity, including higher levels of GSH and higher GSH/GSSG ratios. In a related development, Guayerbas *et al.* concluded that a 4-week treatment of mice with NAC and thioproline protected all animals against early age-associated behavioral impairment, but the improvement was more evident in prematurely aging mice (61). On the other hand, Brown-Borg and collaborators found that in long-lived Ames dwarf mice the flux of methionine through the transsulfuration pathway is enhanced (in part because of upregulation of CBS and CSE), leading to an increased reduced GSH pool, mainly in the liver (136), with heightened resistance to toxic/oxidative challenges, and 50%–64% longer lives than their wild counterparts (males and females, respectively) (207). Importantly, Ames dwarf mice have a delayed occurrence and reduced incidence of presumably fatal neoplastic disease compared with their normal siblings (78).

Protection from NSAID toxicity

Nonsteroidal anti-inflammatory drugs (NSAIDs) also possess analgesic and anti-pyretic effects. The main adverse drug reactions associated with use of NSAIDs are gastrointestinal tract irritation, inhibition of cyclooxygenase (COX)-1 and COX-2 (211), inhibition of enzymatic H₂S synthesis (211, 212), development of cardio- and cerebrovascular pathologies, and development of altered renal function. In fact, in the USA, an estimated 5% of all visits to a doctor are related to prescription of NSAIDs, and NSAID-related upper gastrointestinal adverse drug reactions are believed to result in over 100,000 hospitalizations and around 16,500 deaths yearly (204). Recent studies have shown that over 50% of patients taking NSAIDs have suffered mucosal damage to their small intestine (69). In a very recent and comprehensive meta-analysis, Sven Trelle *et al.* concluded that significantly increased CV risks are associated with taking naproxen, ibuprofen, diclofenac, celecoxib, etoricoxib, lumiracoxib, and rofecoxib (204). Since millions of persons with chronic musculoskeletal symptoms are long-term users of NSAIDs, their doubled risk of heart failure and increased risks of myocardial infarction and stroke are of the utmost concern.

Administration of NSAIDs results in a significant decrease in endogenous H₂S enzymatic production. This effect was most profound with indomethacin, but was also observed with aspirin, diclofenac, and ketoprofen (58). Since endogenous H₂S contributes significantly to mucosal defense (212), it is reasonable to expect that exogenous administration of this mediator would be effective at preventing NSAID-induced mucosal damage. Indeed, H₂S donors such as NaHS (58) and

diallyl disulfide (DADS) (212) were shown to confer mucosal protection from NSAIDs, preventing gastric damage in rodents. Furthermore, DADS prevented naproxen-induced decreases in gastric blood flow and increases in leukocyte adherence. Based on these findings, several research groups have developed NSAID derivatives that release H_2S *in vivo*. These are obtained by conjugating a molecule of an NSAID with one of an H_2S releasing compound. Typically, these H_2S -releasing NSAID derivatives are carboxylic acid esters with general formula $RCOOR'$, obtainable (at least in principle) by condensing the NSAID molecule, which bears the carboxyl moiety with a sulfur-containing phenolic molecule (Fig. 9). The sulfur-bearing H_2S releasing phenols that have been used are shown in Figure 10.

One such H_2S -releasing NSAID, S-diclofenac (ACS15, see below), showed greater anti-inflammatory activity than diclofenac at equimolar doses in several experimental models (184). Treatment with S-diclofenac, but not diclofenac, resulted in a marked reduction in severity of pancreatitis-associated lung injury. Moreover, S-diclofenac has much lower gastrointestinal toxicity than diclofenac and provides marked cardioprotection in a well-characterized experimental model of ischemia-reperfusion injury in the rabbit (164). Furthermore, S-diclofenac effects were accompanied by a significant increase in GSH, inhibition of angiogenesis and cell proliferation, and inhibition of NF- κ B and TNF- α .

Hibernation and protection against hemorrhage

In 2005, Blackstone *et al.* revealed that H_2S induces a hypometabolic state in naturally nonhibernating mice (16). When exposed to nontoxic H_2S concentrations, mice rapidly and reversibly entered a hibernation-like state, which Blackstone *et al.* designated as "suspended animation-like" (16). An 80 ppm H_2S treatment induced, within minutes, a 60% reduction in CO_2 production and oxygen consumption, which can be lowered to over 90%. Additionally, core body temperature decreases to near-ambient, and heart rate and breathing frequency are significantly lowered. Oxygen demand is so drastically diminished that H_2S -treated mice survive for over 6 hours in an atmosphere containing 5% oxygen, whereas untreated controls die within 15 min. Upon cessation of H_2S exposure, the mice awoke without displaying neurological or behavioral abnormalities.

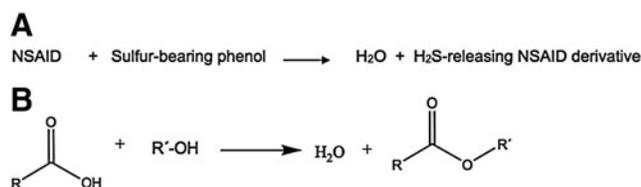


FIG. 9. Several research groups have developed NSAID derivatives that release H_2S *in vivo*. (A) These are obtained by conjugating a molecule of NSAID with one of an H_2S releasing compound. (B) Typically these H_2S -releasing NSAID derivatives are carboxylic acid esters with general formula: $RCOOR'$. These are obtainable (at least in principle) by condensing the NSAID molecule, which bears the carboxyl moiety with a sulfur-containing phenolic molecule.

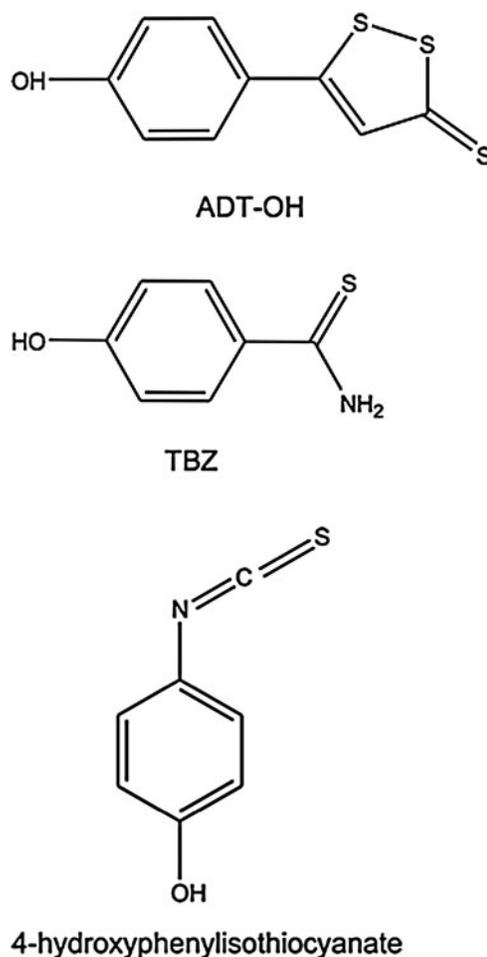


FIG. 10. Chemical structure of some of the sulfur-bearing/ H_2S releasing phenols that have been used in the literature. These phenols include: ADT-OH, TBZ, and 4-hydroxyphenylisothiocyanate.

Following up on these highly newsworthy studies, Morrison *et al.* showed that inhaled H_2S or intravenously-administered Na_2S can protect rats from lethal hemorrhage, with surviving rats free from functional or behavioral deficits (134). In the introduction to this article, the authors state that "clinicians and investigators have long hypothesized that reducing metabolic demand could buy time for patients suffering from insufficient blood supply until they can receive definitive treatment". In effect, this goal is still being actively pursued in many quarters (5, 18, 47, 50), but it is proving extremely difficult to translate the protective effects displayed by H_2S treatment of rats to larger mammals.

In fact, attempts to protect piglets and pigs from hemorrhagic shock failed (50), as well as administration of gaseous H_2S -via extracorporeal membrane lung ventilation to sheep, in an attempt to avoid the potential pulmonary toxicity of H_2S (47). It seems that these failures are related to the fact that the higher doses of H_2S required to depress metabolism in larger mammals elicit toxic effects, systemic and/or pulmonary, and the possibility that the ability of H_2S to abate metabolism depends on the specific metabolic rate of animals. H_2S may reduce metabolism when the baseline metabolic rate is high

(i.e., in awake mice), but not when metabolic rate is already depressed, for instance, in anesthetized mice or sheep (47).

Cancer prevention and treatment

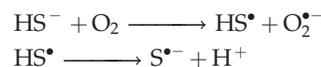
Consideration of the many differential anti-cancer effects of H₂S (collected in Table 1), shows that H₂S and H₂S prodrugs seem to be capable of inhibiting all stages in cancer development. These studies will be further expanded upon below. However, we will first discuss briefly the events that lead to the development of cancer to better understand how H₂S may be used to intervene.

Normal cellular homeostasis is maintained by a balance between the processes of cell proliferation and cell death (apoptosis). An imbalance may lead to uncontrolled cell proliferation and cancer. The causal role played by ROS/RNS in carcinogenesis is now firmly established (203) and two mechanisms are thought to operate: (i) modulation of gene expression, with numerous oncogenes and tumor-suppressor genes operating through redox mechanisms that may be amenable to pharmacological intervention (223), and (ii) induction of genetic modifications. Redox dysregulation contributes to mutations and malignant transformation/progression through mitogenic signaling and modulation of apoptotic and survival pathways. Usually, pro-oxidant deviations from redox homeostasis relate to many aspects of the cancerous phenotype including alterations in metabolism, modulation of the cell cycle, upregulation of anti-apoptotic survival signaling, and upregulation of pro-angiogenic signaling.

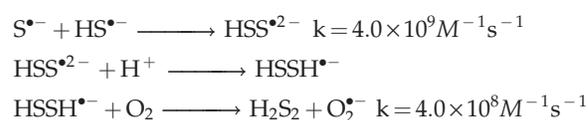
According to the “differential redox set points” hypothesis, pro-oxidant-induced upregulation of intracellular ROS/disulfide stress specifically targets cancer cells, the therapeutic index being determined by the redox differential between the set points of normal and malignant cells. Wondrak (223) uses a highly descriptive analogy of this process with the operation of a car engine, where the red bar displayed on car tachometers denotes the maximum speed at which the car’s engine is designed to operate without being damaged. In cancer cells, with a high set point of oxidative stress, pro-oxidant manipulation induces a redox shift that “redlines” and ruins the cancer cell’s proliferative machinery. In contrast, normal cells

tolerate the same pro-oxidant shift. It is important to note that many redox-targeted cancer drugs (including H₂S donors) have been shown to potentiate the effect of other anticancer agents and radiation, which is consistent with preferential sensitization of cancer cells to the cytotoxicity of the non-redox-directed agent.

H₂S is a Janus-faced molecule that can also behave as a pro-oxidant (6, 7, 189) via its interaction with dioxygen and/or the superoxide ion to generate sulfur-centered and oxygen-centered free radicals as well as higher sulfides H₂S_n (1 < n < 8). A likely mechanism for H₂S₂ formation from NaHS and O₂ in aqueous solution at pH close to 7 is:



Please note that K_a for HS[•] is greater than K_{a1} for H₂S by a factor of about 1000 (118).



The first step would be the slowest in the sequence, but it is efficiently catalyzed by transition metal ions (117).

In turn, inorganic polysulfides (H₂S_n) and organic hydro-polysulfides (RS_nH, n > 1) are known to possess a high tendency to undergo homolysis and generate perthiyl radicals, RS_n[•] and HS_n[•] (137). RS_n[•] and HS_n[•] are highly reactive and easily generate ROS, and react rapidly with oxidants, such as dioxygen and oxyhemoglobin, to form ROS through a pseudocatalytic redox cycle (31, 137). In short, the ability of H₂S to act as pro-oxidant and the high reactivity as both oxidants and reductants (81) of inorganic polysulfides lead us to consider the possibility that, in many kinds of cancer cells, H₂S treatment has pro-oxidant effects that may lead to malignant cell death through redlining.

There is evidence in favor of pro-oxidant redlining of cancer cells by treatment with H₂S or its prodrugs. Treatment of human neuroblastoma SH-SY5Y cells with NAC and ribose-

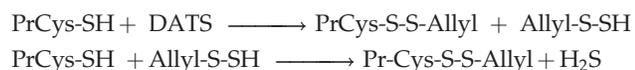
TABLE 1. “DIRECT” ANTICANCER EFFECTS OF H₂S

Effect	Mediator (s)	Affected stage(s)*				References
		1	2	3	4	
1 Increased immunocompetence	GSH (↑), Taurine (↑)	X	X	X	X	(9, 20, 53, 60, 178, 246, 247)
2 Inhibition of procarcinogen activation by oxidases (Cyp-450, etc)	Nrf2 (↑)	X	X	X		(11)
3 Inhibition of NfkB nuclear translocation		X	X	X		(112, 222)
4 Epigenetic silencing of protooncogenes	SAM (↑)	X	X	X		(39)
5 Epigenetic reactivation of tumor suppressor genes	HDAC (↓)	X	X	X		(142)
6 DNA protection/repair	GSH (↑), Trx(↑)	X	X	X		(43, 52, 77, 93, 149)
7 Abolishment of chronic inflammation	GSH (↑), CAMs in Leukocytes (↓)	X	X	X		(9, 143, 151, 196, 201)
8 Prooxidant/proapoptotic “redlining”	Sulfane sulfur (↑) ROS (↑)			X		(56, 104, 166, 223, 235)
9 Antiangiogenesis (at “high levels” of H ₂ S)				X		(79, 184)
10 Antimetastatic effect	E-Cadherin (↑)				X	(74, 120)

*1, Initiation; 2, Promotion; 3, Progression; 4, Metastasis.

cysteine resulted in elevation of sulfane sulfur level and inhibition of their proliferation (85). Diallyl disulfide (which releases H₂S *in vivo*), was found by Filomeni *et al.* to induce neuroblastoma cell death (56). These researchers presented evidence that supports mediation of cytotoxicity by a ROS-dependent c-Jun NH₂-terminal kinase/c-Jun signaling cascade (56). An extensive series of publications by Singh *et al.* (4, 66–68, 92, 154, 231–237), Seki *et al.* (72, 73, 170), Das *et al.* (42), and Lee *et al.* (104) demonstrate that DATS selectively targets DU145 and PC-3 cells in prostate cancer models, amazingly without damaging a normal prostate cancer cell line (237), kills cells of human gastric cancer cell lines, arrests the cell cycle in human cancer cell lines, and is cytotoxic towards a human breast cancer cell line (104), lung adenocarcinoma (228), prostate (231), colon (72), and human glioblastoma cells. This differential effect of DATS has been attributed to induction of intracellular oxidative stress through ROS generation. In a recent report, Lee *et al.* postulate that mitochondria are the main source of ROS generation and that DATS-induced oxidative stress is detected through glutaredoxin (GRX) (104).

DATS contains sulfane sulfur and is an excellent source of H₂S *in vivo* (see section Diabetes and Metabolism), as evidenced by its ability to increase the intracellular GSH level and enhance the antioxidant and detoxification capabilities of rat primary hepatocytes (226). While we consider it likely that the effect of DATS on cancer cells is mediated by H₂S, it has yet to be definitively demonstrated. Although the anticancer effects have been attributed in some cases to reversible covalent modification of specific proteins (73), it is estimated that this may be just an epiphenomenon (81, 137). In fact, Hosono's demonstration that cysteine residues Cys-12 and Cys-354 of beta tubulin are oxidized by DATS to S-allylmercaptocysteine residues constitutes indirect proof of H₂S formation in this system (73):



DATS has been intensively studied in China during the last 25 years. However, most research results were published in obscure Chinese periodicals. Validation of the anticancer and cancer chemopreventative activities of DATS is found in that body of literature, of which the following studies are worth mentioning:

- DATS inhibits mouse colon tumors in mouse CT-26 cells allograft model *in vivo* (227). The authors conclude that DATS may represent a colon cancer-preventing agent.
- A double blind intervention study was performed on 2526 experimental subjects and 2507 persons in the control group, with those in the first group taking doses of 200 mg synthetic DATS daily plus 100 µg of selenium every other day for one month of each year from November 1989 to December 1991. After a 5-year follow-up, it was concluded that the DATS+Se treatment had the effect of decreasing the incidence of digestive cancer by over 50% (64, 251).
- DATS enhances the antitumor function of macrophages either by priming macrophages alone or by synergic action, meanwhile increasing the susceptibility of some tumor cells to macrophage cytotoxicity (247).
- DATS augments the activation of T lymphocytes. This effect is related to inhibition of NO production by

macrophages. In addition, DATS can antagonize the inhibition of tumor-derived immunosuppressive factors produced by S180 cells and Ehrlich ascitic cancer cells on the activation of T lymphocytes and reduce the inhibitory rate significantly. The authors state that DATS is potentially useful in tumor therapy (246).

- Apoptosis of human cholangiocarcinoma FRH-0201 cells can be induced by DATS *in vitro* in a dose-dependent manner (38).
- DATS can induce mitotic arrest in human gastric cell line SGC-7901 (35).
- DATS can cause gastric cancer cell (MGC803 and SGC7901 cell lines) arrest in the M-phase, and this may be one of the mechanisms for inhibiting cell proliferation (63).
- DATS induces apoptosis in human gastric cancer cell line BGC-823 through downregulation of Bcl-2 and increased caspase-3 expression and activity (101, 102).

There are many studies from several laboratories on the cytotoxic effects of organic isothiocyanates (*e.g.*, sulforaphane and benzyl isothiocyanate) derived from cruciferous vegetables (36, 168, 180, 188, 203, 229, 249). These authors report that organic isothiocyanates selectively kill cancer cells (human prostate, human pancreatic, etc.) in culture through ROS-mediated mechanisms. Since hydrolysis of organic isothiocyanates under physiologic conditions may generate H₂S, it is likely that these selective cytotoxic effects are H₂S-mediated (238).

Allyl sodium thiosulfate, also known as 2-propenyl thiosulphate (2-PTS) and also found in garlic, has been shown by two research groups to behave similarly to DATS in many respects (28–30, 166). These authors found that 2-PTS reacts with GSH, under physiologically relevant conditions, generating H₂S. *In vitro*, 2-PTS inhibits proliferation of human tumor cell lines WiDR, 293, HL-60, and HuT78 (human T-lymphoblastoid cell line) in a dose-dependent manner, and caused oxidative damage and apoptosis to HL-60 and HuT 78 cells. Cytotoxicity of 2-PTS is related to a blockage in the G₂/M phase of the cell cycle, which was linked to an early increment in ROS flux, and to inactivation of rhodanese, with concomitant thiolation to yield a protein disulfide highly sensitive to proteolytic degradation.

Moore, Deng and co-workers provided further evidence on the anticancer effects of H₂S (108). They studied the interaction of two H₂S donors (NaHS and GYY4137) with cancer cells *in vitro* and the effect on mice tumors of intraperitoneal injection of 100–300 mg/kg/day of GYY4137, a slow-releasing H₂S donor that persists in the culture medium for up to 7 days, versus only 2 hours for NaHS. GYY4137 (but not NaHS) is cytotoxic to human cancer cells in a concentration-dependent manner. The two H₂S donors studied did not affect the survival of normal human lung fibroblasts (IMR-90 and WI-38), and GYY4137 promoted cancer cell (MCF-7), but not normal cell (IMR-90) apoptosis. It also induced cell cycle arrest of cancer cells in the G₂/M phase. Daily administration of GYY4137 to immunodeficient mice for 14 days caused a dose-dependent reduction in the growth of tumors induced by prior injection of a human leukemia cell line (HL-60 or MB4-11).

Chattopadhyay *et al.* show, in a recent series of articles, that H₂S-releasing-NSAIDs are effective at inhibiting the growth of a variety of cancer cells (32–34). H₂S-releasing-NSAIDs inhibited cell proliferation, promoted apoptosis, and caused G₀/G₁ cell cycle block of eleven different cancer cell lines (33).

The H₂S-releasing-NSAIDs had potencies of 28- over 3,000-fold compared to their NSAID counterparts in these effects, and H₂S-releasing-aspirin (HS-ASA) was consistently more potent than the other H₂S-releasing-NSAIDs tested (33). HS-ASA not only inhibits the growth of HT-29 human colon and Hepa 1c1c7 mouse liver adenocarcinoma cells in culture, it also induces Nrf2 expression and Phase-II detoxifying enzymes *in vivo* (34). HS-ASA also shows promise as a therapeutic agent in estrogen receptor negative breast cancer (32).

Conclusion

H₂S has come to the forefront of some very exciting and promising research to treat a variety of diseases, and several H₂S-releasing prodrugs are currently under development by the pharmaceutical industry. H₂S possesses a very diverse biological profile that includes: potent antioxidant, anti-apoptotic, anti-inflammatory, metabolic, vasoactive, and cytoprotective actions on normal cells that could potentially be harnessed to treat a number of pathological states. The robust antioxidant actions of H₂S involving direct scavenging of toxic reactive oxygen species combined with the effects on antioxidant enzyme expression and function are highly attractive features of this gaseous signaling molecule. It is possible that H₂S prodrugs and novel agents that modulate H₂S bioavailability might be efficacious for acute myocardial infarction, stroke, diabetes, arthritis, peripheral artery disease (PAD), metabolic syndrome, organ transplantation, erectile dysfunction, diabetes, inflammatory bowel disease and pulmonary hypertension. Contrastingly, H₂S appears to exert powerful prooxidant and proapoptotic actions on cancer cells of different origins, which suggests that H₂S prodrugs might be developed into effective anticancer agents capable of achieving high specificity and broad efficacy across different cancer types. However, it is important to fully consider the highly toxic actions of supraphysiological levels of hydrogen sulfide, and great care must be taken during the development of H₂S-based therapeutic agents. This is true of any agent that exerts potent actions on the redox status in both normal cells and cells undergoing oxidative stress during pathological states.

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Disclosure Statement

DJL is a co-founder and consultant for Sulfagenix. GG is a founder and consultant for Sulfagenix. GG is an inventor of several United States patents for the use of hydrogen sulfide-based therapeutics for a number of disease states.

References

- Aggarwal BB and Gehlot P. Inflammation and cancer: How friendly is the relationship for cancer patients? *Curr Opin Pharmacol* 9: 351–369, 2009.
- Aminzadeh MA and Vaziri ND. Downregulation of the renal and hepatic hydrogen sulfide (H₂S)-producing enzymes and capacity in chronic kidney disease. *Nephrol Dial Transplant* 27: 498–504, 2012.
- Anderson MF and Sims NR. The effects of focal ischemia and reperfusion on the glutathione content of mitochondria from rat brain subregions. *J Neurochem* 81: 541–549, 2002.
- Antosiewicz J, Herman-Antosiewicz A, Marynowski SW, and Singh SV. c-Jun NH(2)-terminal kinase signaling axis regulates diallyl trisulfide-induced generation of reactive oxygen species and cell cycle arrest in human prostate cancer cells. *Cancer Res* 66: 5379–5386, 2006.
- Aslami H and Juffermans NP. Induction of a hypometabolic state during critical illness. A new concept in the ICU? *Neth J Med* 68: 190–198, 2010.
- Attene-Ramos MS, Wagner ED, Gaskins HR, and Plewa MJ. Hydrogen sulfide induces direct radical-associated DNA damage. *Mol Cancer Res* 5: 455–459, 2007.
- Attene-Ramos MS, Wagner ED, Plewa MJ, and Gaskins HR. Evidence that hydrogen sulfide is a genotoxic agent. *Mol Cancer Res* 4: 9–14, 2006.
- Baker DH, Bryant KI, Dilger RN, and Parsons CM. Dietary L-homoserine spares threonine in chicks. *J Nutr* 139: 1298–1302, 2009.
- Ballatori N, Krance SM, Notenboom S, Shi S, Tieu K, and Hammond CL. Glutathione dysregulation and the etiology and progression of human diseases. *Biol Chem* 390: 191–214, 2009.
- Banerjee R and Kabil O. Redox biochemistry of hydrogen sulfide. *J Biol Chem* 285: 21903–21907, 2010.
- Bass SE, Sienkiewicz P, Macdonald CJ, Cheng RY, Sparatore A, Del Soldato P, Roberts DD, Moody TW, Wink DA, and Yeh GC. Novel dithiolethione-modified nonsteroidal anti-inflammatory drugs in human hepatoma HepG2 and colon LS180 cells. *Clin Cancer Res* 15: 1964–1972, 2009.
- Beinert H. A tribute to sulfur. *Eur J Biochem* 267: 5657–5664, 2000.
- Benavides GA, Squadrito GL, Mills RW, Patel HD, Isbell TS, Patel RP, Darley-Usmar VM, Doeller JE, and Kraus DW. Hydrogen sulfide mediates the vasoactivity of garlic. *Proc Natl Acad Sci USA* 104: 17977–17982, 2007.
- Biermann J, Lagreze WA, Schallner N, Schwer CI, and Goebel U. Inhalative preconditioning with hydrogen sulfide attenuated apoptosis after retinal ischemia/reperfusion injury. *Mol Vis* 17: 1275–1286, 2011.
- Bivalacqua TJ, Usta MF, Kendirci M, Pradhan L, Alvarez X, Champion HC, Kadowitz PJ, and Hellstrom WJ. Superoxide anion production in the rat penis impairs erectile function in diabetes: Influence of *in vivo* extracellular superoxide dismutase gene therapy. *J Sex Med* 2: 187–197; discussion 197–198, 2005.
- Blackstone E, Morrison M, and Roth MB. H₂S induces a suspended animation-like state in mice. *Science* 308: 518–518, 2005.
- Blanco RA, Ziegler TR, Carlson BA, Cheng PY, Park Y, Cotsonis GA, Accardi CJ, and Jones DP. Diurnal variation in glutathione and cysteine redox states in human plasma. *Am J Clin Nutr* 86: 1016–1023, 2007.
- Bouma HR, Verhaag EM, Otis JP, Heldmaier G, Swoap SJ, Strijkstra AM, Henning RH, and Carey HV. Induction of torpor: Mimicking natural metabolic suppression for biomedical applications. *J Cell Physiol* 227: 1285–1290, 2012.
- Brancaleone V, Roviezzo F, Vellecco V, De Gruttola L, Bucci M, and Cirino G. Biosynthesis of H₂S is impaired in

- non-obese diabetic (NOD) mice. *Br J Pharmacol* 155: 673–680, 2008.
20. Breitkreutz R, Pittack N, Nebe CT, Schuster D, Brust J, Beichert M, Hack V, Daniel V, Edler L, and Droge W. Improvement of immune functions in HIV infection by sulfur supplementation: Two randomized trials. *J Mol Med* 78: 55–62, 2000.
 21. Brosnan JT and Brosnan ME. The sulfur-containing amino acids: An overview. *J Nutr* 136: 1636S–1640S, 2006.
 22. Bucci M, Papapetropoulos A, Vellecco V, Zhou Z, Pyrriochou A, Roussos C, Roviezzo F, Brancaleone V, and Cirino G. Hydrogen sulfide is an endogenous inhibitor of phosphodiesterase activity. *Arterioscler Thromb Vasc Biol* 30: 1998–2004, 2010.
 23. Budde MW and Roth MB. Hydrogen sulfide increases hypoxia-inducible factor-1 activity independently of von Hippel-Lindau tumor suppressor-1 in *C. elegans*. *Mol Biol Cell* 21: 212–217, 2010.
 24. Cai WJ, Wang MJ, Moore PK, Jin HM, Yao T, and Zhu YC. The novel proangiogenic effect of hydrogen sulfide is dependent on Akt phosphorylation. *Cardiovasc Res* 76: 29–40, 2007.
 25. Calvert JW, Elston M, Nicholson CK, Gundewar S, Jha S, Elrod JW, Ramachandran A, and Lefer DJ. Genetic and pharmacologic hydrogen sulfide therapy attenuates ischemia-induced heart failure in mice. *Circulation* 122: 11–19, 2010.
 26. Calvert JW, Jha S, Gundewar S, Elrod JW, Ramachandran A, Pattillo CB, Kevil CG, and Lefer DJ. Hydrogen sulfide mediates cardioprotection through Nrf2 signaling. *Circ Res* 105: 365–374, 2009.
 27. Chan EC, Peshavariya H, Datla SR, Dusting GJ, and Jiang F. Redox regulation of angiogenesis: NADPH oxidase as a drug target. *Anti-angiogen Drug Disc Devel* 1: 86–115, 2011.
 28. Chang HS, Ko M, Ishizuka M, Fujita S, Yabuki A, Hossain MA, and Yamato O. Sodium 2-propenyl thiosulfate derived from garlic induces phase II detoxification enzymes in rat hepatoma H4IIE cells. *Nutr Res* 30: 435–440, 2010.
 29. Chang HS, Yamato O, Yamasaki M, Ko M, and Maede Y. Growth inhibitory effect of alk(en)yl thiosulfates derived from onion and garlic in human immortalized and tumor cell lines. *Cancer Lett* 223: 47–55, 2005.
 30. Chang HS, Yamato O, Yamasaki M, and Maede Y. Modulatory influence of sodium 2-propenyl thiosulfate from garlic on cyclooxygenase activity in canine platelets: Possible mechanism for the anti-aggregatory effect. *Prostaglandins Leukot Essent Fatty Acids* 72: 351–355, 2005.
 31. Chatterji T, Keerthi K, and Gates KS. Generation of reactive oxygen species by a persulfide (BnSSH). *Bioorg Med Chem Lett* 15: 3921–3924, 2005.
 32. Chattopadhyay M, Kodela R, Nath N, Barsegian A, Boring D, and Kashfi K. Hydrogen sulfide-releasing aspirin suppresses NF-kappaB signaling in estrogen receptor negative breast cancer cells *in vitro* and *in vivo*. *Biochem Pharmacol* 83: 723–732, 2012.
 33. Chattopadhyay M, Kodela R, Nath N, Dastagirzada YM, Velazquez-Martinez CA, Boring D, and Kashfi K. Hydrogen sulfide-releasing NSAIDs inhibit the growth of human cancer cells: A general property and evidence of a tissue type-independent effect. *Biochem Pharmacol* 83: 715–722, 2012.
 34. Chattopadhyay M, Kodela R, Nath N, Street CR, Velazquez-Martinez CA, Boring D, and Kashfi K. Hydrogen sulfide-releasing aspirin modulates xenobiotic metabolizing enzymes *in vitro* and *in vivo*. *Biochem Pharmacol* 83: 733–740, 2012.
 35. Chen C-M and Yin M-C. S-allylcysteine, S-ethyl cysteine and S-propyl cysteine alleviate oxidative stress-induced damage within PC-12 cells. *J Sci Food Agric* 88: 2493–2498, 2008.
 36. Chen YR, Wang WF, Kong ANT, and Tan TH. Molecular mechanisms of c-Jun N-terminal kinase-mediated apoptosis induced by anticarcinogenic isothiocyanates. *J Biol Chem* 273: 1769–1775, 1998.
 37. Cheng X, Siow RC, and Mann GE. Impaired redox signaling and antioxidant gene expression in endothelial cells in diabetes: A role for mitochondria and the nuclear factor-E2-related factor 2-Kelch-like ECH-associated protein 1 defense pathway. *Antioxid Redox Signal* 14: 469–487, 2011.
 38. Cong C, Chen H, and Sun J. Apoptosis of FRH-0201 cells in human cholangiocarcinoma induced by allitridi. *Hebei Medicine*, 2008; DOI: CNKI:SUN:HICYX.0.2008-06-000.
 39. Cyr AR and Domann FE. The redox basis of epigenetic modifications: From mechanisms to functional consequences. *Antioxid Redox Signal* 15: 551–589, 2011.
 40. d'Emmanuele di Villa Bianca R, Sorrentino R, Maffia P, Mirone V, Imbimbo C, Fusco F, De Palma R, Ignarro LJ, and Cirino G. Hydrogen sulfide as a mediator of human corpus cavernosum smooth-muscle relaxation. *Proc Natl Acad Sci USA* 106: 4513–4518, 2009.
 41. d'Emmanuele di Villa Bianca R, Sorrentino R, Mirone V, and Cirino G. Hydrogen sulfide and erectile function: A novel therapeutic target. *Nat Rev Urol* 8: 286–289, 2011.
 42. Das A, Banik NL, and Ray SK. Garlic compounds generate reactive oxygen species leading to activation of stress kinases and cysteine proteases for apoptosis in human glioblastoma T98G and U87MG cells. *Cancer* 110: 1083–1095, 2007.
 43. de la Asuncion JG, Millan A, Pla R, Bruseghini L, Esteras A, Pallardo FV, Sastre J, and Vina J. Mitochondrial glutathione oxidation correlates with age-associated oxidative damage to mitochondrial DNA. *FASEB J* 10: 333–338, 1996.
 44. Deng W, Bivalacqua TJ, Champion HC, Hellstrom WJ, Murthy SN, and Kadowitz PJ. Superoxide dismutase. A target for gene therapeutic approach to reduce oxidative stress in erectile dysfunction. *Methods Mol Biol* 610: 213–227, 2010.
 45. Denisov ET and Afanas'ev IB. *Oxidation and Antioxidants in Organic Chemistry and Biology*, Boca Raton, LA; 2005.
 46. Denizalti M, Bozkurt TE, Akpulat U, Sahin-Erdemli I, and Abacioglu N. The vasorelaxant effect of hydrogen sulfide is enhanced in streptozotocin-induced diabetic rats. *Naunyn Schmiedebergs Arch Pharmacol* 383: 509–517, 2011.
 47. Derwall M, Francis RC, Kida K, Bougaki M, Crimi E, Adrie C, Zapol WM, and Ichinose F. Administration of hydrogen sulfide via extracorporeal membrane lung ventilation in sheep with partial cardiopulmonary bypass perfusion: A proof of concept study on metabolic and vasomotor effects. *Crit Care* 15: R51, 2011.
 48. Desai KM, Chang T, Untereiner A, and Wu L. Hydrogen sulfide and the metabolic syndrome. *Expert Rev Clin Pharmacol* 4: 63–73, 2011.
 49. Dombkowski RA, Russell MJ, Schulman AA, Doellman MM, and Olson KR. Vertebrate phylogeny of hydrogen sulfide vasoactivity. *Am J Physiol Regul Integr Comp Physiol* 288: R243–52, 2005.
 50. Drabek T, Kochanek PM, Stezoski J, Wu XR, Bayir H, Morhard RC, Stezoski SW, and Tisherman SA. Intravenous hydrogen sulfide does not induce hypothermia or improve survival from hemorrhagic shock in pigs. *Shock* 35: 67–73, 2011.
 51. Droge W. Oxidative stress and ageing: Is ageing a cysteine deficiency syndrome? *Philos Trans R Soc Lond B Biol Sci* 360: 2355–2372, 2005.

52. Droge W and Kinscherf R. Aberrant insulin receptor signaling and amino acid homeostasis as a major cause of oxidative stress in aging. *Antioxid Redox Signal* 10: 661–678, 2008.
53. Droge W, Schulze-Osthoff K, Mihm S, Galter D, Schenk H, Eck HP, Roth S, and Gmunder H. Functions of glutathione and glutathione disulfide in immunology and immunopathology. *FASEB J* 8: 1131–1138, 1994.
54. Duan Y, Wang B. Influence of allitridi on oxidative stress reaction in rats with type 2 diabetes mellitus. *Mod J Integ Trad Chinese Western Med* 2010; DOI: CNKI:SUN:XDJH.0.2010-18-017.
55. Elrod JW, Calvert JW, Morrison J, Doeller JE, Kraus DW, Tao L, Jiao X, Scalia R, Kiss L, Szabo C, Kimura H, Chow CW, and Lefer DJ. Hydrogen sulfide attenuates myocardial ischemia-reperfusion injury by preservation of mitochondrial function. *Proc Natl Acad Sci USA* 104: 15560–15565, 2007.
56. Filomeni G, Aquilano K, Rotilio G, and Ciriolo MR. Reactive oxygen species-dependent c-Jun NH2-terminal kinase/c-Jun signaling cascade mediates neuroblastoma cell death induced by diallyl disulfide. *Cancer Res* 63: 5940–5949, 2003.
57. Fiorucci S, Antonelli E, Distrutti E, Rizzo G, Mencarelli A, Orlandi S, Zanardo R, Renga B, Di Sante M, Morelli A, Cirino G, and Wallace JL. Inhibition of hydrogen sulfide generation contributes to gastric injury caused by anti-inflammatory nonsteroidal drugs. *Gastroenterology* 129: 1210–1224, 2005.
58. Fiorucci S, Distrutti E, Cirino G, and Wallace JL. The emerging roles of hydrogen sulfide in the gastrointestinal tract and liver. *Gastroenterology* 131: 259–271, 2006.
59. Giustarini D, Del Soldato P, Sparatore A, and Rossi R. Modulation of thiol homeostasis induced by H2S-releasing aspirin. *Free Radic Biol Med* 48: 1263–1272, 2010.
60. Grimble R. Sulfur amino acids, glutathione, and immune function. In: *Glutathione and Sulfur Amino Acids in Human Health and Disease*, edited by Roberta M, Giuseppe M. Hoboken, NJ: John Wiley and Sons, Inc.; 2009. pp. 273–288.
61. Guayerbas N, Puerto M, Hernanz A, Miquel J, and De la Fuente M. Thiolic antioxidant supplementation of the diet reverses age-related behavioural dysfunction in prematurely ageing mice. *Pharmacol Biochem Behav* 80: 45–51, 2005.
62. Gupta S, Kuhnisch J, Mustafa A, Lhotak S, Schlachterman A, Slifker MJ, Klein-Szanto A, High KA, Austin RC, and Kruger WD. Mouse models of cystathionine beta-synthase deficiency reveal significant threshold effects of hyperhomocysteinemia. *FASEB J* 23: 883–893, 2009.
63. Ha MW, Ma R, Shun LP, Gong YH, and Yuan Y. Effects of allitridi on cell cycle arrest of human gastric cancer cells. *World J Gastroenterol* 11: 5433–5437, 2005.
64. Hao L, Hui-quing L, Yun W, Hai-xiu X, Wan-teng F, Mei-ling W, Pei-hong S, and Xio-yan X. An intervention study to prevent gastric cancer by micro-selenium and large dose of allitridum. *Chin Med J (Engl)* 117: 1155–1160, 2004.
65. Henderson PW, Singh SP, Weinstein AL, Nagineni V, Rafii DC, Kadouch D, Krijgh DD, and Spector JA. Therapeutic metabolic inhibition: Hydrogen sulfide significantly mitigates skeletal muscle ischemia reperfusion injury *in vitro* and *in vivo*. *Plast Reconstr Surg* 126: 1890–1898, 2010.
66. Herman-Antosiewicz A, Powolny AA, and Singh SV. Molecular targets of cancer chemoprevention by garlic-derived organosulfides. *Acta Pharmacol Sin* 28: 1355–1364, 2007.
67. Herman-Antosiewicz A and Singh SV. Checkpoint kinase 1 regulates diallyl trisulfide-induced mitotic arrest in human prostate cancer cells. *J Biol Chem* 280: 28519–28528, 2005.
68. Herman-Antosiewicz A, Stan SD, Hahm ER, Xiao D, and Singh SV. Activation of a novel ataxia-telangiectasia mutated and Rad3 related/checkpoint kinase 1-dependent prometaphase checkpoint in cancer cells by diallyl trisulfide, a promising cancer chemopreventive constituent of processed garlic. *Mol Cancer Ther* 6: 1249–1261, 2007.
69. Higuchi K, Umegaki E, Watanabe T, Yoda Y, Morita E, Murano M, Tokioka S, and Arakawa T. Present status and strategy of NSAIDs-induced small bowel injury. *J Gastroenterol* 44: 879–888, 2009.
70. Hildebrandt TM and Grieshaber MK. Redox regulation of mitochondrial sulfide oxidation in the lugworm, *Arenicola marina*. *J Exp Biol* 211: 2617–2623, 2008.
71. Hildebrandt TM and Grieshaber MK. Three enzymatic activities catalyze the oxidation of sulfide to thiosulfate in mammalian and invertebrate mitochondria. *FEBS J* 275: 3352–3361, 2008.
72. Hosono T, Fukao T, Ogihara J, Ito Y, Shiba H, Seki T, and Ariga T. Diallyl trisulfide suppresses the proliferation and induces apoptosis of human colon cancer cells through oxidative modification of beta-tubulin. *J Biol Chem* 280: 41487–41493, 2005.
73. Hosono T, Hosono-Fukao T, Inada K, Tanaka R, Yamada H, Iitsuka Y, Seki T, Hasegawa I, and Ariga T. Alkenyl group is responsible for the disruption of microtubule network formation in human colon cancer cell line HT-29 cells. *Carcinogenesis* 29: 1400–1406, 2008.
74. Howard EW, Ling MT, Chua CW, Cheung HW, Wang X, and Wong YC. Garlic-derived S-allylmercaptocysteine is a novel *in vivo* antimetastatic agent for androgen-independent prostate cancer. *Clin Cancer Res* 13: 1847–1856, 2007.
75. Hsu CC, Huang CN, Hung YC, and Yin MC. Five cysteine-containing compounds have antioxidative activity in Balb/cA mice. *J Nutr* 134: 149–152, 2004.
76. Huovinen JA and Gustafsson BE. Inorganic sulphate, sulphite and sulphide as sulphur donors in the biosynthesis of sulphur amino acids in germ-free and conventional rats. *Biochim Biophys Acta* 136: 441–447, 1967.
77. Huseby N-E, Sundkvist E, and Svineng G. Glutathione and sulfur containing amino acids: antioxidant and conjugation activities. In: *Glutathione and Sulfur Amino Acids in Human Health and Disease*, eds. Masella R, Mazza G. Hoboken, NJ: John Wiley and Sons, Inc.; 2009. pp. 93–120.
78. Ikeno Y, Bronson RT, Hubbard GB, Lee S, and Bartke A. Delayed occurrence of fatal neoplastic diseases in Ames dwarf mice: Correlation to extended longevity. *J Gerontol A Biol Sci Med Sci* 58: 291–296, 2003.
79. Isenberg JS, Jia Y, Field L, Ridnour LA, Sparatore A, Del Soldato P, Sowers AL, Yeh GC, Moody TW, Wink DA, Ramchandran R, and Roberts DD. Modulation of angiogenesis by dithiolethione-modified NSAIDs and valproic acid. *Br J Pharmacol* 151: 63–72, 2007.
80. Ishigami M, Hiraki K, Umemura K, Ogasawara Y, Ishii K, and Kimura H. A source of hydrogen sulfide and a mechanism of its release in the brain. *Antioxid Redox Signal* 11: 205–214, 2009.
81. Jacob C, Anwar A, and Burkholz T. Perspective on recent developments on sulfur-containing agents and hydrogen sulfide signaling. *Planta Med* 74: 1580–1592, 2008.

82. Jha S, Calvert JW, Duranski MR, Ramachandran A, and Lefer DJ. Hydrogen sulfide attenuates hepatic ischemia-reperfusion injury: Role of antioxidant and antiapoptotic signaling. *Am J Physiol Heart Circ Physiol* 295: H801–806, 2008.
83. Jones DP. Radical-free biology of oxidative stress. *Am J Physiol Cell Physiol* 295: C849–868, 2008.
84. Jung KA and Kwak MK. The Nrf2 system as a potential target for the development of indirect antioxidants. *Molecules* 15: 7266–7291, 2010.
85. Jurkowska H and Wrobel M. N-acetyl-L-cysteine as a source of sulfane sulfur in astrocytoma and astrocyte cultures: Correlations with cell proliferation. *Amino Acids* 34: 231–237, 2008.
86. Kabil O and Banerjee R. Redox biochemistry of hydrogen sulfide. *J Biol Chem* 285: 21903–21907, 2010.
87. Kamoun P. Endogenous production of hydrogen sulfide in mammals. *Amino Acids* 26: 243–254, 2004.
88. Kamyshny A, Ekeltchik I, Gun J, and Lev O. Method for the determination of inorganic polysulfide distribution in aquatic systems. *Anal Chem* 78: 2631–2639, 2006.
89. Kamyshny A, Zilberbrand M, Ekeltchik I, Voitsekovski T, Gun J, and Lev O. Speciation of polysulfides and zerovalent sulfur in sulfide-rich water wells in southern and central Israel. *Aqua Geochem* 14: 171–192, 2008.
90. Kaneko Y, Kimura T, Taniguchi S, Souma M, Kojima Y, Kimura Y, Kimura H, and Niki I. Glucose-induced production of hydrogen sulfide may protect the pancreatic beta-cells from apoptotic cell death by high glucose. *FEBS Lett* 583: 377–382, 2009.
91. Kida K, Yamada M, Tokuda K, Marutani E, Kakinohana M, Kaneki M, and Ichinose F. Inhaled hydrogen sulfide prevents neurodegeneration and movement disorder in a mouse model of Parkinson's disease. *Antioxid Redox Signal* 15: 343–352, 2011.
92. Kim YA, Xiao D, Xiao H, Powolny AA, Lew KL, Reilly ML, Zeng Y, Wang Z, and Singh SV. Mitochondria-mediated apoptosis by diallyl trisulfide in human prostate cancer cells is associated with generation of reactive oxygen species and regulated by Bax/Bak. *Mol Cancer Ther* 6: 1599–1609, 2007.
93. Kimura Y, Goto Y, and Kimura H. Hydrogen sulfide increases glutathione production and suppresses oxidative stress in mitochondria. *Antioxid Redox Signal* 12: 1–13, 2010.
94. Kimura Y and Kimura H. Hydrogen sulfide protects neurons from oxidative stress. *FASEB J* 18: 1165–1167, 2004.
95. Kitteringham NR, Abdullah A, Walsh J, Randle L, Jenkins RE, Sison R, Goldring CE, Powell H, Sanderson C, Williams S, Higgins L, Yamamoto M, Hayes J, and Park BK. Proteomic analysis of Nrf2 deficient transgenic mice reveals cellular defence and lipid metabolism as primary Nrf2-dependent pathways in the liver. *J Proteom* 73: 1612–1631, 2010.
96. Klaassen CD and Reisman SA. Nrf2 the rescue: Effects of the antioxidative/electrophilic response on the liver. *Toxicol Appl Pharmacol* 244: 57–65, 2010.
97. Klatt P, Molina EP, De Lacoba MG, Padilla CA, Martinez-Galesteo E, Barcena JA, and Lamas S. Redox regulation of c-Jun DNA binding by reversible S-glutathiolation. *FASEB J* 13: 1481–1490, 1999.
98. Kloesch B, Liszt M, Steiner G, and Broll J. Inhibitors of p38 and ERK1/2 MAPkinase and hydrogen sulphide block constitutive and IL-1beta-induced IL-6 and IL-8 expression in the human chondrocyte cell line C-28/I2. *Rheumatol Int* 32: 729–736, 2012.
99. Kraus J, Packman S, Fowler B, and Rosenberg LE. Purification and properties of cystathionine beta-synthase from human liver. Evidence for identical subunits. *J Biol Chem* 253: 6523–6528, 1978.
100. Laggner H, Muellner MK, Schreier S, Sturm B, Hermann M, Exner M, Gmeiner BM, and Kapiotis S. Hydrogen sulphide: A novel physiological inhibitor of LDL atherogenic modification by HOCl. *Free Radic Res* 41: 741–747, 2007.
101. Lan H and Lu YY. [Effect of allitridi on cyclin D1 and p27(Kip1) protein expression in gastric carcinoma BGC823 cells]. *Ai Zheng* 22: 1268–1271, 2003.
102. Lan H and Lu YY. Allitridi induces apoptosis by affecting Bcl-2 expression and caspase-3 activity in human gastric cancer cells. *Acta Pharmacol Sin* 25: 219–225, 2004.
103. Lands LC, Grey VL, and Smountas AA. Effect of supplementation with a cysteine donor on muscular performance. *J Appl Physiol* 87: 1381–1385, 1999.
104. Lee BC, Park BH, Kim SY, and Lee YJ. Role of Bim in diallyl trisulfide-induced cytotoxicity in human cancer cells. *J Cell Biochem* 112: 118–127, 2011.
105. Lee M, Sparatore A, Del Soldato P, McGeer E, and McGeer PL. Hydrogen sulfide-releasing NSAIDs attenuate neuroinflammation induced by microglial and astrocytic activation. *Glia* 58: 103–113, 2010.
106. Lee M, Tazzari V, Giustarini D, Rossi R, Sparatore A, Del Soldato P, McGeer E, and McGeer PL. Effects of hydrogen sulfide-releasing L-DOPA derivatives on glial activation: Potential for treating Parkinson disease. *J Biol Chem* 285: 17318–17328, 2010.
107. Lee S, Park Y, Zuidema MY, Hannink M, and Zhang C. Effects of interventions on oxidative stress and inflammation of cardiovascular diseases. *World J Cardiol* 3: 18–24, 2011.
108. Lee ZW, Zhou J, Chen CS, Zhao Y, Tan CH, Li L, Moore PK, and Deng LW. The slow-releasing hydrogen sulfide donor, GYY4137, exhibits novel anti-cancer effects *in vitro* and *in vivo*. *PLoS One* 6: e21077, 2011.
109. Lefer DJ. Potential importance of alterations in hydrogen sulphide (H₂S) bioavailability in diabetes. *Br J Pharmacol* 155: 617–619, 2008.
110. Lefer DJ, Scalia R, Campbell B, Nossuli T, Hayward R, Salamon M, Grayson J, and Lefer AM. Peroxynitrite inhibits leukocyte-endothelial cell interactions and protects against ischemia-reperfusion injury in rats. *J Clin Invest* 99: 684–691, 1997.
111. Leiser SF and Miller RA. Nrf2 signaling, a mechanism for cellular stress resistance in long-lived mice. *Mol Cell Biol* 30: 871–884, 2010.
112. Li L, Rose P, and Moore PK. Hydrogen sulfide and cell signaling. *Annu Rev Pharmacol Toxicol* 51: 169–187, 2011.
113. Li L, Salto-Tellez M, Tan CH, Whiteman M, and Moore PK. GYY4137, a novel hydrogen sulfide-releasing molecule, protects against endotoxic shock in the rat. *Free Radic Biol Med* 47: 103–113, 2009.
114. Limon-Pacheco JH, Hernandez NA, Fanjul-Moles ML, and Gonshebat ME. Glutathione depletion activates mitogen-activated protein kinase (MAPK) pathways that display organ-specific responses and brain protection in mice. *Free Radic Biol Med* 43: 1335–1347, 2007.
115. Lin CC and Yin MC. Antiglycative and anti-VEGF effects of S-ethyl cysteine and S-propyl cysteine in kidney of diabetic mice. *Mol Nutr Food Res* 52: 1358–1364, 2008.

116. Loffredo L, Marcoccia A, Pignatelli P, Andreozzi P, Borgia MC, Cangemi R, Chiarotti F, and Violi F. Oxidative-stress-mediated arterial dysfunction in patients with peripheral arterial disease. *Eur Heart J* 28: 608–612, 2007.
117. Luther GW, Findlay AJ, MacDonald DJ, Owings SM, Hanson TE, Beinart RA, and Girguis PR. Thermodynamics and kinetics of sulfide oxidation by oxygen: A look at inorganically controlled reactions and biologically mediated processes in the environment. *Front Microbiol* 2: 62, 2011.
118. Lykakis IN, Ferreri C, and Chatgililoglu C. The sulfhydryl radical (HS(·)/S(-·)): A contender for the isomerization of double bonds in membrane lipids. *Angew Chem Int Ed Engl* 46: 1914–1916, 2007.
119. Lynn EG and Austin RC. Hydrogen sulfide in the pathogenesis of atherosclerosis and its therapeutic potential. *Expert Rev Clin Pharmacol* 4: 97–108, 2010.
120. Ma K, Liu Y, Zhu Q, Liu CH, Duan JL, Tan BK, and Zhu YZ. H₂S donor, S-propargyl-cysteine, increases CSE in SGC-7901 and cancer-induced mice: Evidence for a novel anti-cancer effect of endogenous H₂S? *PLoS One* 6: e20525, 2011.
121. Maher J and Yamamoto M. The rise of antioxidant signaling—The evolution and hormetic actions of Nrf2. *Toxicol Appl Pharmacol* 244: 4–15, 2010.
122. Mantovani G, Madeddu C, Maccio A, Gramignano G, Lusso MR, Massa E, Astaro G, and Serpe R. Cancer-related anorexia/cachexia syndrome and oxidative stress: An innovative approach beyond current treatment. *Cancer Epidemiol Biomarkers Prev* 13: 1651–1659, 2004.
123. Mari M, Morales A, Colell A, Garcia-Ruiz C, and Fernandez-Checa JC. Mitochondrial glutathione, a key survival antioxidant. *Antioxid Redox Signal* 11: 2685–2700, 2009.
124. Marsden PA. Low-molecular-weight S-nitrosothiols and blood vessel injury. *J Clin Invest* 117: 2377–2380, 2007.
125. Matteucci E and Giampietro O. Thiol signalling network with an eye to diabetes. *Molecules* 15: 8890–8903, 2010.
126. Medeiros JV, Bezerra VH, Gomes AS, Barbosa AL, Lima-Junior RC, Soares PM, Brito GA, Ribeiro RA, Cunha FQ, and Souza MH. Hydrogen sulfide prevents ethanol-induced gastric damage in mice: Role of ATP-sensitive potassium channels and capsaicin-sensitive primary afferent neurons. *J Pharmacol Exp Ther* 330: 764–770, 2009.
127. Meng JL, Mei WY, Dong YF, Wang JH, Zhao CM, Lan AP, Yang CT, Chen PX, Feng JQ, and Hu CH. Heat shock protein 90 mediates cytoprotection by HS against chemical hypoxia-induced injury in PC12 cells. *Clin Exp Pharmacol Physiol* 38: 42–49, 2011.
128. Mikami Y, Shibuya N, Kimura Y, Nagahara N, Ogasawara Y, and Kimura H. Thioredoxin and dihydrolipoic acid are required for 3-mercaptopyruvate sulfurtransferase to produce hydrogen sulfide. *Biochem J* 439: 479–485, 2011.
129. Mikami Y, Shibuya N, Kimura Y, Nagahara N, Yamada M, and Kimura H. Hydrogen sulfide protects the retina from light-induced degeneration by the modulation of Ca²⁺ influx. *J Biol Chem* 286: 39379–39386, 2011.
130. Miller DL and Roth MB. Hydrogen sulfide increases thermotolerance and lifespan in *Caenorhabditis elegans*. *Proc Natl Acad Sci USA* 104: 20618–20622, 2007.
131. Minamishima S, Bougaki M, Sips PY, Yu JD, Minamishima YA, Elrod JW, Lefer DJ, Bloch KD, and Ichinose F. Hydrogen sulfide improves survival after cardiac arrest and cardiopulmonary resuscitation via a nitric oxide synthase 3-dependent mechanism in mice. *Circulation* 120: 888–896, 2009.
132. Mitsuhashi H, Yamashita S, Ikeuchi H, Kuroiwa T, Kaneko Y, Hiromura K, Ueki K, and Nojima Y. Oxidative stress-dependent conversion of hydrogen sulfide to sulfite by activated neutrophils. *Shock* 24: 529–534, 2005.
133. Miyoshi N, Takabayashi S, Osawa T, and Nakamura Y. Benzyl isothiocyanate inhibits excessive superoxide generation in inflammatory leukocytes: Implication for prevention against inflammation-related carcinogenesis. *Carcinogenesis* 25: 567–575, 2004.
134. Morrison ML, Blackwood JE, Lockett SL, Iwata A, Winn RK, and Roth MB. Surviving blood loss using hydrogen sulfide. *J Trauma* 65: 183–188, 2008.
135. Morsy MA, Ibrahim SA, Abdelwahab SA, Zedan MZ, and Elbitar HI. Curative effects of hydrogen sulfide against acetaminophen-induced hepatotoxicity in mice. *Life Sci* 87: 692–698, 2010.
136. Mosharov E, Cranford MR, and Banerjee R. The quantitatively important relationship between homocysteine metabolism and glutathione synthesis by the transsulfuration pathway and its regulation by redox changes. *Biochemistry* 39: 13005–13011, 2000.
137. Munchberg U, Anwar A, Mecklenburg S, and Jacob C. Polysulfides as biologically active ingredients of garlic. *Organic Biomol Chem* 5: 1505–1518, 2007.
138. Mustafa AK, Gadalla MM, Sen N, Kim S, Mu W, Gazi SK, Barrow RK, Yang G, Wang R, and Snyder SH. H₂S signals through protein S-sulfhydration. *Sci Signal* 2: ra72, 2009.
139. Nagy P and Winterbourn CC. Rapid reaction of hydrogen sulfide with the neutrophil oxidant hypochlorous acid to generate polysulfides. *Chem Res Toxicol* 23: 1541–1543, 2010.
140. Nagy P and Winterbourn CC. Chapter 6: Redox chemistry of biological thiols. In: *Advances in Molecular Toxicology*, ed. James CF. Elsevier; 2010. pp. 183–222.
141. Nelson KC, Armstrong JS, Moriarty S, Cai J, Wu MW, Sternberg P, Jr., and Jones DP. Protection of retinal pigment epithelial cells from oxidative damage by oltipraz, a cancer chemopreventive agent. *Invest Ophthalmol Vis Sci* 43: 3550–3554, 2002.
142. Nian H, Delage B, Ho E, and Dashwood RH. Modulation of histone deacetylase activity by dietary isothiocyanates and allyl sulfides: Studies with sulforaphane and garlic organosulfur compounds. *Environ Mol Mutagen* 50: 213–221, 2009.
143. Nimni ME, Han B, and Cordoba F. Are we getting enough sulfur in our diet? *Nutr Metab (Lond)* 4: 24, 2007.
144. Oh GS, Pae HO, Lee BS, Kim BN, Kim JM, Kim HR, Jeon SB, Jeon WK, Chae HJ, and Chung HT. Hydrogen sulfide inhibits nitric oxide production and nuclear factor-kappaB via heme oxygenase-1 expression in RAW264.7 macrophages stimulated with lipopolysaccharide. *Free Radic Biol Med* 41: 106–119, 2006.
145. Olson KR. Is hydrogen sulfide a circulating “gasotransmitter” in vertebrate blood? *Biochim Biophys Acta* 1787: 856–863, 2009.
146. Olson KR and Whitfield NL. Hydrogen sulfide and oxygen sensing in the cardiovascular system. *Antioxid Redox Signal* 12: 1219–1234, 2010.
147. Osborne NN, Ji D, Abdul Majid AS, Fawcett RJ, Sparatore A, and Del Soldato P. ACS67, a hydrogen sulfide-releasing derivative of latanoprost acid, attenuates retinal ischemia and oxidative stress to RGC-5 cells in culture. *Invest Ophthalmol Vis Sci* 51: 284–294, 2010.
148. Otani H. Oxidative stress as pathogenesis of cardiovascular risk associated with metabolic syndrome. *Antioxid Redox Signal* 15: 1911–1926, 2011.

149. Pallardo FV, Markovich J, Garcia JL, and Viña F. Role of nuclear glutathione as a key regulator of cell proliferation. *Molecular Aspects of Medicine* 30: 77–85, 2009.
150. Palmer LA, Doctor A, Chhabra P, Sheram ML, Laubach VE, Karlinsey MZ, Forbes MS, Macdonald T, and Gaston B. S-nitrosothiols signal hypoxia-mimetic vascular pathology. *J Clin Invest* 117: 2592–2601, 2007.
151. Pan LL, Liu XH, Gong QH, Wu D, and Zhu YZ. Hydrogen sulfide attenuated tumor necrosis factor- α -induced inflammatory signaling and dysfunction in vascular endothelial cells. *PLoS One* 6: e19766, 2011.
152. Pinto JT, Krasnikov BF, and Cooper AJ. Redox-sensitive proteins are potential targets of garlic-derived mercaptocysteine derivatives. *J Nutr* 136: 835S–841S, 2006.
153. Porter PN, Grishaver MS, and Jones OW. Characterization of human cystathionine beta-synthase. Evidence for the identity of human L-serine dehydratase and cystathionine beta-synthase. *Biochim Biophys Acta* 364: 128–139, 1974.
154. Powolny AA and Singh SV. Multitargeted prevention and therapy of cancer by diallyl trisulfide and related Allium vegetable-derived organosulfur compounds. *Cancer Lett* 269: 305–314, 2008.
155. Predmore BL and Lefer DJ. Development of hydrogen sulfide-based therapeutics for cardiovascular disease. *J Cardiovasc Transl Res* 3: 487–498, 2010.
156. Predmore BL and Lefer DJ. Hydrogen sulfide-mediated myocardial pre- and post-conditioning. *Expert Rev Clin Pharmacol* 4: 83–96, 2011.
157. Pryor WA, Gojon G, and Church DF. Relative rate constants for hydrogen atom abstraction by the cyclohexanethiyl and benzenethiyl radicals. *J Organic Chem* 43: 793–800, 1978.
158. Pryor WA, Houk KN, Foote CS, Fukuto JM, Ignarro LJ, Squadrito GL, and Davies KJ. Free radical biology and medicine: It's a gas, man! *Am J Physiol Regul Integr Comp Physiol* 291: R491–511, 2006.
159. Qiu X, Villalta J, Lin G, and Lue TF. Role of hydrogen sulfide in the physiology of penile erection. *J Androl* 2011; [Epub ahead of Print]; DOI: 10.2164/jandrol.111.014936.
160. Rabai G, Orban M, and Epstein IR. Systematic design of chemical oscillators. A model for the pH-regulated oscillatory reaction between hydrogen peroxide and sulfide ion. *J Phys Chem* 96: 5414–5419, 1992.
161. Rahangdale S, Yeh SY, Malhotra A, and Veves A. Therapeutic interventions and oxidative stress in diabetes. *Front Biosci* 14: 192–209, 2009.
162. Reisman SA, Buckley DB, Tanaka Y, and Klaassen CD. CDDO-Im protects from acetaminophen hepatotoxicity through induction of Nrf2-dependent genes. *Toxicol Appl Pharmacol* 236: 109–114, 2009.
163. Rosado JO, Salvador M, and Bonatto D. Importance of the trans-sulfuration pathway in cancer prevention and promotion. *Mol Cell Biochem* 301: 1–12, 2007.
164. Rossoni G, Sparatore A, Tazzari V, Manfredi B, Del Soldato P, and Berti F. The hydrogen sulphide-releasing derivative of diclofenac protects against ischaemia-reperfusion injury in the isolated rabbit heart. *Br J Pharmacol* 153: 100–109, 2008.
165. Ryter SW, Alam J, and Choi AMK. Heme oxygenase-1/carbon monoxide: From basic science to therapeutic applications. *Physiol Rev* 86: 583–650, 2006.
166. Sabelli R, Iorio E, De Martino A, Podo F, Ricci A, Viticchie G, Rotilio G, Paci M, and Melino S. Rhodanese-thioredoxin system and allyl sulfur compounds. *FEBS J* 275: 3884–3899, 2008.
167. Saez G, Thornalley PJ, Hill HA, Hems R, and Bannister JV. The production of free radicals during the autoxidation of cysteine and their effect on isolated rat hepatocytes. *Biochim Biophys Acta* 719: 24–31, 1982.
168. Sahu RP, Zhang R, Batra S, Shi Y, and Srivastava SK. Benzyl isothiocyanate-mediated generation of reactive oxygen species causes cell cycle arrest and induces apoptosis via activation of MAPK in human pancreatic cancer cells. *Carcinogenesis* 30: 1744–1753, 2009.
169. Schalinske KL. Hepatic sulfur amino acid metabolism. In: *Glutathione and Sulfur Amino Acids in Human Health and Disease*, eds. Masella R, Mazza G. Hoboken, NJ: John Wiley and Sons, Inc.; 2009. pp. 73–90.
170. Seki T, Hosono T, Hosono-Fukao T, Inada K, Tanaka R, Ogihara J, and Ariga T. Anticancer effects of diallyl trisulfide derived from garlic. *Asia Pac J Clin Nutr* 17: 249–252, 2008.
171. Sen N, Paul BD, Gadalla MM, Mustafa AK, Sen T, Xu R, Kim S, and Snyder SH. Hydrogen sulfide-linked sulfhydration of NF- κ B mediates its antiapoptotic actions. *Mol Cell* 45: 13–24, 2012.
172. Sen N and Snyder SH. Protein modifications involved in neurotransmitter and gasotransmitter signaling. *Trends Neurosci* 33: 493–502, 2010.
173. Sen U, Basu P, Abe OA, Givvimani S, Tyagi N, Metreveli N, Shah KS, Passmore JC, and Tyagi SC. Hydrogen sulfide ameliorates hyperhomocysteinemia-associated chronic renal failure. *Am J Physiol Renal Physiol* 297: F410–419, 2009.
174. Shin S, Wakabayashi J, Yates MS, Wakabayashi N, Dolan PM, Aja S, Liby KT, Sporn MB, Yamamoto M, and Kensler TW. Role of Nrf2 in prevention of high-fat diet-induced obesity by synthetic triterpenoid CDDO-imidazolide. *Eur J Pharmacol* 620: 138–144, 2009.
175. Shukla N, Rossoni G, Hotston M, Sparatore A, Del Soldato P, Tazzari V, Persad R, Angelini GD, and Jeremy JY. Effect of hydrogen sulphide-donating sildenafil (ACS6) on erectile function and oxidative stress in rabbit isolated corpus cavernosum and in hypertensive rats. *Bju Int* 103: 1522–1529, 2009.
176. Sidhu R, Singh M, Samir G, and Carson RJ. L-cysteine and sodium hydrosulphide inhibit spontaneous contractility in isolated pregnant rat uterine strips *in vitro*. *Pharmacol Toxicol* 88: 198–203, 2001.
177. Sies H. Preface. In: *Antioxidants in Disease Mechanisms and Therapy*, ed. Sies H. San Diego, CA: Academic Press; 1996. pp. xxiii.
178. Sies H. Glutathione and its role in cellular functions. *Free Radic Biol Med* 27: 916–921, 1999.
179. Simon F, Giudici R, Duy CN, Schelzig H, Oter S, Groger M, Wachter U, Vogt J, Speit G, Szabo C, Radermacher P, and Calzia E. Hemodynamic and metabolic effects of hydrogen sulfide during porcine ischemia/reperfusion injury. *Shock* 30: 359–364, 2008.
180. Singh SV, Srivastava SK, Choi S, Lew KL, Antosiewicz J, Xiao D, Zeng Y, Watkins SC, Johnson CS, Trump DL, Lee YJ, Xiao H, and Herman-Antosiewicz A. Sulforaphane-induced cell death in human prostate cancer cells is initiated by reactive oxygen species. *J Biol Chem* 280: 19911–19924, 2005.
181. Sodha NR, Clements RT, Feng J, Liu Y, Bianchi C, Horvath EM, Szabo C, Stahl GL, and Sellke FW. Hydrogen sulfide therapy attenuates the inflammatory response in a porcine model of myocardial ischemia/reperfusion injury. *J Thorac Cardiovasc Surg* 138: 977–984, 2009.

182. Sommer F, Klotz T, Steinritz D, and Bloch W. Evaluation of tetrahydrobiopterin (BH4) as a potential therapeutic agent to treat erectile dysfunction. *Asian J Androl* 8: 159–167, 2006.
183. Sparatore A, Perrino E, Tazzari V, Giustarini D, Rossi R, Rossoni G, Erdman K, Schroder H, and Soldato PD. Pharmacological profile of a novel H₂S-releasing aspirin. *Free Radical Biol Med* 46: 586–592, 2009.
184. Sparatore A, Santus G, Giustarini D, Rossi R, and Del Soldato P. Therapeutic potential of new hydrogen sulfide-releasing hybrids. *Expert Rev Clin Pharmacol* 4: 109–121, 2011.
185. Srilatha B, Adaikan PG, Li L, and Moore PK. Hydrogen sulphide: A novel endogenous gasotransmitter facilitates erectile function. *J Sex Med* 4: 1304–1311, 2007.
186. Srilatha B, Adaikan PG, and Moore PK. Possible role for the novel gasotransmitter hydrogen sulphide in erectile dysfunction—A pilot study. *Eur J Pharmacol* 535: 280–282, 2006.
187. Srilatha B, Hu L, Adaikan GP, and Moore PK. Initial characterization of hydrogen sulfide effects in female sexual function. *J Sex Med* 6: 1875–1884, 2009.
188. Srivastava SK and Singh SV. Cell cycle arrest, apoptosis induction and inhibition of nuclear factor kappa B activation in anti-proliferative activity of benzyl isothiocyanate against human pancreatic cancer cells. *Carcinogenesis* 25: 1701–1709, 2004.
189. Stasko A, Brezova V, Zalibera M, Biskupic S, and Ondrias K. Electron transfer: A primary step in the reactions of sodium hydrosulphide, an H₂S/HS(-) donor. *Free Radic Res* 43: 581–593, 2009.
190. Stipanuk MH. Sulfur amino acid metabolism: Pathways for production and removal of homocysteine and cysteine. *Annu Rev Nutr* 24: 539–577, 2004.
191. Stoyanovsky DA, Maeda A, Atkins JL, and Kagan VE. Assessments of thyl radicals in biosystems: Difficulties and new applications. *Anal Chem* 83: 6432–6438, 2011.
192. Sykietis GP, Habeos IG, Samuelson AV, and Bohmann D. The role of the antioxidant and longevity-promoting Nrf2 pathway in metabolic regulation. *Curr Opin Clin Nutr Metab Care* 14: 41–48, 2011.
193. Takeuchi K, Kita K, Hayashi S, and Aihara E. Regulatory mechanism of duodenal bicarbonate secretion. Roles of endogenous prostaglandins and nitric oxide. *Pharmacol Therapeut* 130: 59–70, 2011.
194. Tan BH, Wong PT, and Bian JS. Hydrogen sulfide: A novel signaling molecule in the central nervous system. *Neurochem Int* 56: 3–10, 2010.
195. Tan Y, Ichikawa T, Li J, Si Q, Yang H, Chen X, Goldblatt CS, Meyer CJ, Li X, Cai L, and Cui T. Diabetic down-regulation of Nrf2 activity via ERK contributes to oxidative stress-induced insulin resistance in cardiac cells *in vitro* and *in vivo*. *Diabetes* 60: 625–633, 2011.
196. Tang G, Wu L, and Wang R. Interaction of hydrogen sulfide with ion channels. *Clin Exp Pharmacol Physiol* 37: 753–763, 2010.
197. Taniguchi S, Kang L, Kimura T, and Niki I. Hydrogen sulphide protects mouse pancreatic beta-cells from cell death induced by oxidative stress, but not by endoplasmic reticulum stress. *Br J Pharmacol* 162: 1171–1178, 2011.
198. Taniguchi S and Niki I. Significance of hydrogen sulfide production in the pancreatic beta-cell. *J Pharmacol Sci* 116: 1–5, 2011.
199. Tay AS, Hu LF, Lu M, Wong PT, and Bian JS. Hydrogen sulfide protects neurons against hypoxic injury via stimulation of ATP-sensitive potassium channel/protein kinase C/extracellular signal-regulated kinase/heat shock protein 90 pathway. *Neuroscience* 167: 277–286, 2010.
200. Tiranti V, Viscomi C, Hildebrandt T, Di Meo I, Minerì R, Tiveron C, Levitt MD, Prella A, Fagiolarì G, Rimoldi M, and Zeviani M. Loss of ETHE1, a mitochondrial dioxxygenase, causes fatal sulfide toxicity in ethylmalonic encephalopathy. *Nat Med* 15: 200–205, 2009.
201. Tirouvanziam R, Conrad CK, Bottiglieri T, Herzenberg LA, and Moss RB. High-dose oral N-acetylcysteine, a glutathione prodrug, modulates inflammation in cystic fibrosis. *Proc Natl Acad Sci USA* 103: 4628–4633, 2006.
202. Tomaskova Z, Cacanyiova S, Benco A, Kristek F, Dugovicova L, Hrbac J, and Ondrias K. Lipids modulate H₂S/HS(-) induced NO release from S-nitrosoglutathione. *Biochem Biophys Res Commun* 390: 1241–1244, 2009.
203. Trachootham D, Lu WQ, Ogasawara MA, Valle NRD, and Huang P. Redox regulation of cell survival. *Antioxid Redox Signal* 10: 1343–1374, 2008.
204. Trelle S, Reichenbach S, Wandel S, Hildebrand P, Tschanen B, Villiger PM, Egger M, and Juni P. Cardiovascular safety of non-steroidal anti-inflammatory drugs: Network meta-analysis. *Br Med J* 342: c7086, 2011.
205. Tyagi N, Givvimani S, Qipshidze N, Kundu S, Kapoor S, Vacek JC, and Tyagi SC. Hydrogen sulfide mitigates matrix metalloproteinase-9 activity and neurovascular permeability in hyperhomocysteinemic mice. *Neurochem Int* 56: 301–307, 2010.
206. Ubuka T. Assay methods and biological roles of labile sulfur in animal tissues. *J Chromatogr B Analyt Technol Biomed Life Sci* 781: 227–249, 2002.
207. Uthus EO and Brown-Borg HM. Methionine flux to trans-sulfuration is enhanced in the long living Ames dwarf mouse. *Mech Ageing Dev* 127: 444–450, 2006.
208. Viscomi C, Burlina AB, Dweikat I, Savoirdo M, Lamperti C, Hildebrandt T, Tiranti V, and Zeviani M. Combined treatment with oral metronidazole and N-acetylcysteine is effective in ethylmalonic encephalopathy. *Nat Med* 16: 869–871, 2010.
209. Wakabayashi N, Slocum SL, Skoko JJ, Shin S, and Kensler TW. When NRF2 talks, who's listening? *Antioxid Redox Signal* 13: 1649–1663, 2010.
210. Wallace JL. Hydrogen sulfide-releasing anti-inflammatory drugs. *Trends Pharmacol Sci* 28: 501–505, 2007.
211. Wallace JL. Prostaglandins, NSAIDs, and gastric mucosal protection: Why doesn't the stomach digest itself? *Physiol Rev* 88: 1547–1565, 2008.
212. Wallace JL. Physiological and pathophysiological roles of hydrogen sulfide in the gastrointestinal tract. *Antioxid Redox Signal* 12: 1125–1133, 2010.
213. Wallace JL, Vong L, McKnight W, Dickey M, and Martin GR. Endogenous and exogenous hydrogen sulfide promotes resolution of colitis in rats. *Gastroenterology* 137: 569–578, 578 e1, 2009.
214. Wang M, Li K, Zhu RR, Cheng LL, Wu QS, and Wang SL. The protective function of hydrogen sulfide for lysozyme against riboflavin-sensitized photo-oxidation. *J Photochem Photobiol B-Biol* 103: 186–191, 2011.
215. Wang MJ, Cai WJ, Li N, Ding YJ, Chen Y, and Zhu YC. The hydrogen sulfide donor NaHS promotes angiogenesis in a rat model of hind limb ischemia. *Antioxid Redox Signal* 12: 1065–1077, 2010.
216. Wang R. Two's company, three's a crowd: Can H₂S be the third endogenous gaseous transmitter? *FASEB J* 16: 1792–1798, 2002.

217. Watanabe A, Okada K, Shimizu Y, Wakabayashi H, Higuchi K, Niiya K, Kuwabara Y, Yasuyama T, Ito H, Tsukishiro T, Kondoh Y, Emi N, and Kohri H. Nutritional therapy of chronic hepatitis by whey protein (non-heated). *J Med* 31: 283–302, 2000.
218. Westrop GD, Georg I, and Coombs GH. The mercaptopyruvate sulfurtransferase of *Trichomonas vaginalis* links cysteine catabolism to the production of thioredoxin persulfide. *J Biol Chem* 284: 33485–33494, 2009.
219. Whiteman M, Armstrong JS, Chu SH, Jia-Ling S, Wong BS, Cheung NS, Halliwell B, and Moore PK. The novel neuromodulator hydrogen sulfide: An endogenous peroxynitrite 'scavenger'? *J Neurochem* 90: 765–768, 2004.
220. Whiteman M, Cheung NS, Zhu YZ, Chu SH, Siau JL, Wong BS, Armstrong JS, and Moore PK. Hydrogen sulphide: A novel inhibitor of hypochlorous acid-mediated oxidative damage in the brain? *Biochem Biophys Res Commun* 326: 794–798, 2005.
221. Whiteman M, Gooding KM, Whatmore JL, Ball CI, Mawson D, Skinner K, Tooke JE, and Shore AC. Adiposity is a major determinant of plasma levels of the novel vasodilator hydrogen sulphide. *Diabetologia* 53: 1722–1726, 2010.
222. Whiteman M, Li L, Rose P, Tan CH, Parkinson DB, and Moore PK. The effect of hydrogen sulfide donors on lipopolysaccharide-induced formation of inflammatory mediators in macrophages. *Antioxid Redox Signal* 12: 1147–1154, 2010.
223. Wondrak GT. Redox-directed cancer therapeutics: Molecular mechanisms and opportunities. *Antioxid Redox Signal* 11: 3013–3069, 2009.
224. Wood JL. Sulfane sulfur. *Methods Enzymol* 143: 25–29, 1987.
225. Wrobel M, Lewandowska I, Bronowicka-Adamska P, and Paszewski A. The level of sulfane sulfur in the fungus *Aspergillus nidulans* wild type and mutant strains. *Amino Acids* 37: 565–571, 2009.
226. Wu CC, Lii CK, Tsai SJ, and Sheen LY. Diallyl trisulfide modulates cell viability and the antioxidation and detoxification systems of rat primary hepatocytes. *J Nutr* 134: 724–728, 2004.
227. Wu PP, Liu KC, Huang WW, Chueh FS, Ko YC, Chiu TH, Lin JP, Kuo JH, Yang JS, and Chung JG. Diallyl trisulfide (DATS) inhibits mouse colon tumor in mouse CT-26 cells allograft model *in vivo*. *Phytomedicine* 18: 672–676, 2011.
228. Wu XJ, Hu Y, Lamy E, and Mersch-Sundermann V. Apoptosis induction in human lung adenocarcinoma cells by oil-soluble allyl sulfides: Triggers, pathways, and modulators. *Environ Mol Mutagen* 50: 266–275, 2009.
229. Wu XJ and Hua X. Targeting ROS: Selective killing of cancer cells by a cruciferous vegetable derived pro-oxidant compound. *Cancer Biol Ther* 6: 646–647, 2007.
230. Xia M, Chen L, Muh RW, Li PL, and Li NJ. Production and actions of hydrogen sulfide, a novel gaseous bioactive substance, in the kidneys. *J Pharmacol Exp Therap* 329: 1056–1062, 2009.
231. Xiao D, Choi S, Johnson DE, Vogel VG, Johnson CS, Trump DL, Lee YJ, and Singh SV. Diallyl trisulfide-induced apoptosis in human prostate cancer cells involves c-Jun N-terminal kinase and extracellular-signal regulated kinase-mediated phosphorylation of Bcl-2. *Oncogene* 23: 5594–5606, 2004.
232. Xiao D, Lew KL, Kim YA, Zeng Y, Hahm ER, Dhir R, and Singh SV. Diallyl trisulfide suppresses growth of PC-3 human prostate cancer xenograft *in vivo* in association with Bax and Bak induction. *Clin Cancer Res* 12: 6836–6843, 2006.
233. Xiao D, Pinto JT, Gundersen GG, and Weinstein IB. Effects of a series of organosulfur compounds on mitotic arrest and induction of apoptosis in colon cancer cells. *Mol Cancer Ther* 4: 1388–1398, 2005.
234. Xiao D, Pinto JT, Soh JW, Deguchi A, Gundersen GG, Pallazzo AF, Yoon JT, Shirin H, and Weinstein IB. Induction of apoptosis by the garlic-derived compound S-allylmercaptocysteine (SAMC) is associated with microtubule depolymerization and c-Jun NH(2)-terminal kinase 1 activation. *Cancer Res* 63: 6825–6837, 2003.
235. Xiao D, Powolny AA, Antosiewicz J, Hahm ER, Bommarreddy A, Zeng Y, Desai D, Amin S, Herman-Antosiewicz A, and Singh SV. Cellular responses to cancer chemopreventive agent D,L-sulforaphane in human prostate cancer cells are initiated by mitochondrial reactive oxygen species. *Pharm Res* 26: 1729–1738, 2009.
236. Xiao D and Singh SV. Diallyl trisulfide, a constituent of processed garlic, inactivates Akt to trigger mitochondrial translocation of BAD and caspase-mediated apoptosis in human prostate cancer cells. *Carcinogenesis* 27: 533–540, 2006.
237. Xiao D, Zeng Y, Hahm ER, Kim YA, Ramalingam S, and Singh SV. Diallyl trisulfide selectively causes Bax- and Bak-mediated apoptosis in human lung cancer cells. *Environ Mol Mutagen* 50: 201–212, 2009.
238. Yang G. Hydrogen sulfide in cell survival: A double-edged sword. *Expert Rev Clin Pharmacol* 4: 33–47, 2011.
239. Yang G, Wu L, Jiang B, Yang W, Qi J, Cao K, Meng Q, Mustafa AK, Mu W, Zhang S, Snyder SH, and Wang R. H₂S as a physiologic vasorelaxant: Hypertension in mice with deletion of cystathionine gamma-lyase. *Science* 322: 587–590, 2008.
240. Yonezawa D, Sekiguchi F, Miyamoto M, Taniguchi E, Honjo M, Masuko T, Nishikawa H, Kawabata A. A protective role of hydrogen sulfide against oxidative stress in rat gastric mucosal epithelium. *Toxicology* 241: 11–18, 2007.
241. Yong QC, Lee SW, Foo CS, Neo KL, Chen X, and Bian JS. Endogenous hydrogen sulphide mediates the cardioprotection induced by ischemic postconditioning. *Am J Physiol Heart Circ Physiol* 295: H1330–H1340, 2008.
242. Yusof M, Kamada K, Kalogeris T, Gaskin FS, and Korthuis RJ. Hydrogen sulfide triggers late-phase preconditioning in postischemic small intestine by an NO- and p38 MAPK-dependent mechanism. *Am J Physiol Heart Circ Physiol* 296: H868–876, 2009.
243. Zanardo RC, Brancalone V, Distrutti E, Fiorucci S, Cirino G, and Wallace JL. Hydrogen sulfide is an endogenous modulator of leukocyte-mediated inflammation. *FASEB J* 20: 2118–2120, 2006.
244. Zeng T, Zhang CL, Zhu ZP, Yu LH, Zhao XL, and Xie KQ. Diallyl trisulfide (DATS) effectively attenuated oxidative stress-mediated liver injury and hepatic mitochondrial dysfunction in acute ethanol-exposed mice. *Toxicology* 252: 86–91, 2008.
245. Zhang DD. The Nrf2-Keap1-ARE signaling pathway: The regulation and dual function of Nrf2 in cancer. *Antioxid Redox Signal* 13: 1623–1626, 2010.
246. Zhang G, Feng Z, Hao T, Zhang H, and Jiang Z. Effect of allitridum on the activation of T-lymphocytes. *Pharmacol Clinics Chinese Materia Medica*, 1995; DOI: CNKI:SUN:ZYLL.0.1995-01-008.
247. Zhang G, Feng Z, Hao T, Zhang H, and Jiang Z. Effect of allitridum on macrophage-mediated cytotoxicity. *China J Chinese Materia Medica* 21: 45, 1996.

248. Zhang H, Gao Y, Zhao F, Dai Z, Meng T, Tu S, and Yan Y. Hydrogen sulfide reduces mRNA and protein levels of beta-site amyloid precursor protein cleaving enzyme 1 in PC12 cells. *Neurochem Int* 58: 169–175, 2011.
249. Zhang R, Loganathan S, Humphreys I, and Srivastava SK. Benzyl isothiocyanate-induced DNA damage causes G2/M cell cycle arrest and apoptosis in human pancreatic cancer cells. *J Nutr* 136: 2728–2734, 2006.
250. Zhao ZZ, Wang Z, Li GH, Wang R, Tan JM, Cao X, Suo R, and Jiang ZS. Hydrogen sulfide inhibits macrophage-derived foam cell formation. *Exp Biol Med (Maywood)* 236: 169–176, 2011.
251. Zheng GH, Li H, Fan WT, and Li HQ. [Study on the long-time effect on allitridum and selenium in prevention of digestive system cancers]. *Zhonghua Liu Xing Bing Xue Za Zhi* 26: 110–112, 2005.
252. Zhu X, Gu H, and Ni X. Hydrogen sulfide in the endocrine and reproductive systems. *Expert Rev Clin Pharmacol* 4: 75–82, 2011.

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Abbreviations Used

2-PTS = 2-propenyl thiosulfate
 3-MST = 3-mercaptopyruvate sulfurtransferase
 ACAT1 = acyl-coenzyme A:cholesterol
 acyltransferase-1
 Akt = protein kinase B
 Apaf-1 = apoptotic protease activating factor-1
 AREs = antioxidant responsive elements
 ATP = adenosine triphosphate
 cAMP = cyclic adenosine monophosphate
 CAMs = cell adhesion molecules
 CBS = cystathionine beta-synthase
 cGMP = cyclic guanosine monophosphate
 CO = carbon monoxide
 COX = cyclooxygenase
 CSE = cystathionine gamma-lyase
 CV = cardiovascular
 DATS = diallyl trisulfide
 DTT = dithiolethione
 ED = erectile dysfunction
 EE = ethylmalonic encephalopathy
 ERK1/2 = p44/42 MAPK
 FMD = flow-mediated dilation

GAPDH = glyceraldehyde 3-phosphate
 dehydrogenase
 GDOPs = garlic-derived organic polysulfides
 GRX = glutaredoxin
 GSH = glutathione
 GSSH = glutathione persulfide
 Hcy = homocysteine
 HIF = hypoxia inducible factor
 HO-1 = heme oxygenase-1
 H₂S = hydrogen sulfide
 HS-ASA = H₂S-releasing-aspirin
 HSP = heat shock protein
 IBD = inflammatory bowel disease
 IL = interleukin
 INF-gamma = interferon gamma
 I/R = ischemia-reperfusion
 I-R-I = ischemia-reperfusion injury
 K_{ATP} = ATP-sensitive K⁺ channels
 LDL = low density lipoprotein
 L-DOPA = L-3,4-dihydroxyphenylalanine
 LPS = lipopolysaccharide
 MAPK = mitogen-activated protein kinase
 MPTP = 1-methyl-4-phenyl-1,2,3,6-
 tetrahydropyridine
 mtGSH = mitochondrial glutathione
 NAC = N-acetylcysteine
 NADPH = nicotinamide adenine dinucleotide
 phosphate
 NF-κB = nuclear factor kappa-light-chain-
 enhancer of activated B cells
 NMDA = N-methyl-D-aspartic acid
 NO = nitric oxide
 Nrf2 = nuclear factor erythroid 2-related factor 2
 NSAIDs = nonsteroidal anti-inflammatory drugs
 OTC = L-2-oxothiazolidine-4-carboxylate
 PAD = peripheral arterial disease
 PAPs = adenosine-3'-phosphate-5'-
 phosphosulfate
 PD = Parkinson's disease
 RBC = red blood cells
 RGC = retinal ganglion cells
 RISK = reperfusion injury salvage kinase
 RNS = reactive nitrogen species
 ROS = reactive oxygen species
 RSSH = hydroperosulfide
 SAAs = sulfur containing amino acids
 SHY = S-sulfhydration
 S-NSAIDs = dithiolethione-modified nonsteroidal
 anti-inflammatory drugs
 SR-A = scavenger receptor A
 -SSH = hydroperosulfide group
 STAT-3 = signal transducer and activator of
 transcription-3
 TGF = tumor growth factor
 TNF-α = tumor necrosis factor alpha
 Trx = thioredoxin
 VEGF = vascular endothelial growth factor
 VEGFR2 = VEGF receptor 2