Hydrogen Sulfide: An Endogenous Mediator of Resolution of Inflammation and Injury

John L. Wallace^{1,2}, Jose G.P. Ferraz,² and Marcelo N. Muscara^{2,3}

Abstract

Significance: Hydrogen sulfide is emerging as an important mediator of many aspects of inflammation, and perhaps most importantly as a factor promoting the resolution of inflammation and repair of injury. *Recent Advances:* In the gastrointestinal tract, H₂S has been shown to promote healing of ulcers and the resolution of mucosal inflammation. On the other hand, suppression of endogenous H₂S synthesis impairs mucosal defense and leads to increased granulocyte infiltration. H₂S has been exploited in the design of more effective and safe anti-inflammatory drugs. *Critical Issues:* Enteric bacteria can be a significant source of H₂S, which could affect mucosal integrity; indeed, luminal H₂S can serve as an alternative to oxygen as a metabolic substrate for mitochondrial respiration in epithelial cells. Enterocytes and colonocytes thereby represent a "metabolic barrier" to the diffusion of bacteria-derived H₂S into the subepithelial space. A compromise of this barrier could result in modulation of mucosal function and integrity by bacterial H₂S. *Future Directions:* Improvements in methods for measurement of H₂S and development of more selective inhibitors are crucial for gaining a better understanding of the pathophysiological importance of this mediator. Results from animal studies suggest that H₂S-releasing agents are promising therapeutic agents for many indications, but these compounds need to be assessed in a clinical setting. *Antioxid. Redox Signal.* 17, 58–67.

Introduction

WHILE CLINICAL BENEFITS of hydrogen sulfide, at least in the context of sulfur hot springs, have been espoused for centuries, it is only in the past 20 years that H₂S has been recognized as an important mediator of physiological processes (1, 74). Indeed, as was the case for two other gaseous mediators (nitric oxide and carbon monoxide), the physiological effects of H₂S were overshadowed by the toxicity associated with high concentrations of this substance (44). There are many similarities among H₂S, carbon monoxide, and nitric oxide. Enzymes for the synthesis of all three of these gaseous mediators have been identified and more continue to be identified, at least in the case of H_2S (50). All three mediators have very short half-lives in vivo and produce primarily beneficial effects at physiological concentrations while contributing to injury at super-physiological concentrations. All three of these mediators bind to hemoglobin (48, 63) and can influence activity and/or expression of enzymes responsible for synthesis of the other gaseous mediators (24, 39, 73–75). Also, attempts have been made to exploit the beneficial effects of each of the three gaseous mediators in designing novel therapeutic agents.

In this article, the ability of H_2S to modulate inflammation is reviewed, with a particular focus on the role of the mediator in resolution of inflammation. The use of H_2S as a therapeutic modality is also reviewed, along with the potential effects of bacteria-derived H_2S in modulating inflammation and mucosal integrity in the digestive tract and possibly in other organs.

H₂S and Inflammation

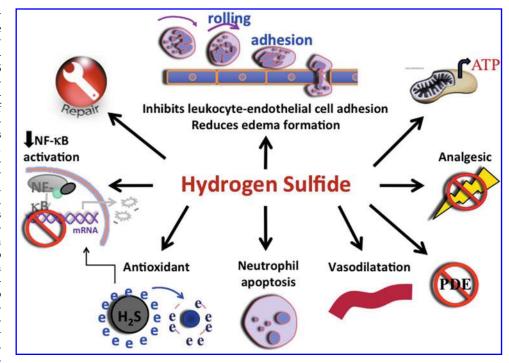
One of the first physiological effects of H_2S that was identified was its ability to relax vascular smooth muscle (73, 75), resulting in vasodilation, a hallmark of inflammation. Several studies have subsequently highlighted the importance of H_2S in inflammation (27, 30, 54, 56). As was the case for nitric oxide, the literature on H_2S in inflammation was initially contradictory, but in recent years a general pattern has emerged consistent with this mediator exerting anti-inflammatory effects, except at high concentrations (56). Moreover, there are now substantial data supportive of a role of H_2S in promoting resolution of inflammation and healing of injured tissue. Figure 1 summarizes some of the key effects of H_2S with respect to inflammation and injury. These

¹Farncombe Family Digestive Health Research Institute, McMaster University, Hamilton, Ontario, Canada.

²Division of Gastroenterology, University of Calgary, Calgary, Alberta, Canada.

³Department of Pharmacology, University of Sao Paulo, Sao Paulo, Brazil.

FIG. 1. Anti-inflammatory effects of H₂S. This figure illustrates some of the key ways in which H₂S can reduce inflammation. H_2S suppresses leukocyte adherence to the vascular endothelium and migration of leukocytes into the subendothelial space, as well as reducing plasma exudation. H₂S has been shown to reduce expression of many pro-inflammatory cytokines, chemokines, and enzymes, most likely related to its ability to suppress activation of nuclear transcription factor- κ B (NF- κ B). H₂S is also a potent anti-oxidant and can induce apoptosis in neutrophils (which can lead to their phagocytosis by macrophages). Promotion of tissue repair by H₂S is likely mediated in part by its vasodilatory actions and en-



hancement of cyclooxygenase-2 (COX-2) expression and through promotion of angiogenesis. Inhibition of phosphodiesterases (PDE) by H₂S leads to elevated cyclic AMP and/or cyclic GMP levels, which can contribute to its antiinflammatory effects. H₂S exhibits anti-nociceptive effects in the viscera, likely due, at least in part, to activation of ATPsensitive potassium channels. In many cells in the body, and in particular in gastrointestinal epithelial cells, H₂S can act as an energy source (generating ATP), substituting for oxygen in mitochondrial respiration. This appears to contribute significantly to protection and repair of tissue injury.

effects include the ability of H₂S to inhibit leukocyte adherence to the vascular endothelium and the subsequent extravasation of leukocytes (72). The impact of this effect of H₂S can be seen in various models of inflammation, in which sulfide salts or H₂S donors are able to reduce infiltration of neutrophils and lymphocytes (17, 72). These effects are likely due, at least in part, to reduced expression of endothelial and/ or leukocyte adhesion molecule expression following exposure to H₂S (18). H₂S acts as a tonic down-regulator of leukocyte adherence; thus, inhibition of H₂S synthesis leads to leukocyte adherence (72). Treatment of rats with inhibitors of H₂S synthesis resulted in a marked increase in mucosal inflammation (elevated granulocyte levels) and an increase in susceptibility to injury (18, 58, 59) in the gastrointestinal tract. This may have been in part due to reduced basal levels of cyclooxygenase-2 (COX-2) expression and a parallel reduction of mucosal prostaglandin E2 (PGE2) synthesis (61). COX-2 and PGE₂ play important roles in the maintenance of mucosal defense in the digestive tract, as well as in modulating mucosal inflammation (3, 49, 61, 70, 71).

In addition to modulating leukocyte adhesion and recruitment, H_2S can reduce plasma exudation (edema formation), while inhibitors of H_2S augment edema formation triggered by an inflammatory agent (59). These effects may contribute to the enhanced edema-reducing effects of H_2S releasing nonsteroidal anti-inflammatory drugs (NSAIDs) in rat models of acute (carrageenan) and chronic (Freund's adjuvant) paw swelling (58, 59).

The ability of H_2S to reduce inflammation has been demonstrated in a variety of animal models, including kaolin/

carrageenan-induced monoarthritis in rats (2), tobacco-smoke induced lung inflammation in mice (12, 23), synovitis in rats (17), ischemia–reperfusion injury in mice (76), and in rat and mouse models of colitis (19, 56, 61). Whiteman *et al.* (66) demonstrated that H_2S is present in synovial fluid of patients with rheumatoid arthritis and osteoarthritis, with levels correlating to disease activity, but the role of H_2S in those conditions remains unclear.

H₂S was recently shown to inhibit phosphodiesterase activity, and this may contribute to anti-inflammatory actions in some circumstances (via elevation of intracellular cyclic AMP and/or cyclic GMP) (9). Scavenging of oxidants and peroxynitrite may also contribute to the anti-inflammatory activity of H_2S (64, 65). Inhibition of nuclear transcription factor- κB (NF- κ B) has been reported in several models (27, 28, 38), and consistent with this, H₂S reduces pro-inflammatory cytokine, chemokine, and enzyme (e.g., inducible nitric oxide synthase [iNOS]) expression (17, 19, 51, 67). In addition, the antioxidant activity of H₂S can be mediated by up-regulation of enzymes such as superoxide dismutase, glutathione peroxase dismutase, and thioredoxin as assessed in rats subjected to intestinal ischemia-reperfusion injury (31) or in brain endothelial cells under methionine-induced oxidative stress in vitro (53) or even by inhibition of NADPH oxidase activity, as in the case of osteoblasts exposed to hydrogen peroxide in vitro (69). With many of these putative mechanisms of action of H₂S, it is unclear if the concentrations required for such actions are achieved in a physiological setting. Difficulties in measuring H₂S levels in vivo further complicate the assessment of the relative significance of these mechanisms in various scenarios.

H₂S has been shown to exert anti-nociceptive effects in some (15, 18, 19, 62), but not all (37) visceral pain studies. The discrepancies may be related to the different methods for measuring pain or to different doses or routes of administration of the H₂S-releasing agents. In a recent study performed in our laboratory, we found that two H₂Sgenerating substances (NaHS and Lawesson's reagent) dose-dependently reduced gastric distention-induced visceral pain (measured through cardio-autonomic responses) in the rat in an ATP-sensitive potassium channel-dependent manner (62). The ability of H₂S to exert peripheral antinociceptive effects has also been demonstrated in some, but not all studies. While anti-nociceptive effects of H₂S were reported by Cunha et al. (13) in a model of peripheral pain induced by bacterial endotoxin and by Ekundi-Valentim et al. (17) in rats with carrageenan-induced knee joint synovitis (see Fig. 2), Andruski et al. (2) observed a significant inhibition of leukocyte recruitment by an H₂S donor in a rat model of monoarthritis, but no significant effect on pain sensitivity.

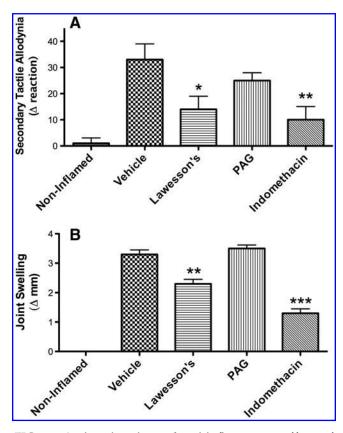


FIG. 2. Anti-nociceptive and anti-inflammatory effects of hydrogen sulfide in experimental (carrageenan-induced) synovitis in rats. (A) Pain responses and (B) joint swelling. Pretreatment with an H₂S donor (Lawesson's reagent; 3.6 μ mol intra-articularly), markedly reduced secondary tactile allodynia, while an inhibitor of H₂S synthesis (D/L-propargylglycine [PAG], 53 μ mol intra-articularly) had no significant effect. Indomethacin (6 mg/kg intraperitoneally) was used as a positive control. *p<0.05, **p<0.01, ***p<0.001 versus the vehicle-treated group. Adapted from previously published data (17).

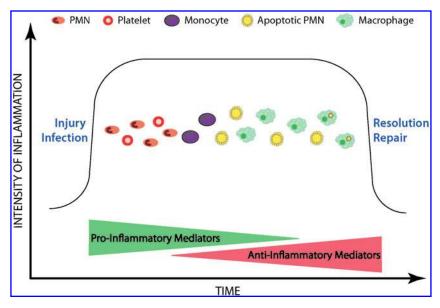
H₂S and Resolution of Inflammation/Injury

With the ability to inhibit so many elements of acute inflammation, it is not surprising that H₂S contributes significantly to the resolution of inflammation and injury. Inflammatory reactions are driven largely by soluble, proinflammatory mediators, such as leukotrienes, histamine, bradykinin, platelet-activating factor, and interleukin (IL)-1, to name just a few (49). Counteracting the effects of these pro-inflammatory mediators are a variety of soluble mediators that down-regulate inflammation, including lipoxins, certain prostaglandins, annexin-1 (AnxA-1), and IL-10 (49). An over-production of pro-inflammatory mediators or an under-production of anti-inflammatory mediators can lead to progression from acute to chronic inflammation. Resolution of inflammation occurs through removal of the triggers of the inflammatory response (e.g., a foreign body or organism), inhibition of the recruitment of neutrophils to the site of injury, and induction of apoptosis of the infiltrated neutrophils and their subsequent clearance by macrophages (Fig. 3). Macrophages undergo a phenotype shift from pro-inflammatory to anti-inflammatory during this process.

There is emerging evidence that H_2S may participate in several stages of the process of resolution of inflammation. As already mentioned, H_2S can reduce leukocyte adherence to the vascular endothelium and leukocyte migration to sites of injury (72). H_2S can also induce neutrophil apoptosis (34), and there is recent evidence that it can trigger significant changes in macrophage function consistent with a shift to a pro-resolution phenotype (16). Specifically, exposure of murine bone marrow–derived macrophages to H_2S resulted in a significant enhancement of phagocytosis of bacteria. H_2S also suppressed endotoxin-induced tumor necrosis factor α (TNF α) production by macrophages, while enhancing chemotaxis. *In vivo*, in a mouse peritonitis model, H_2S significantly reduced granulocyte infiltration while maintaining macrophage numbers (16).

H₂S also appears to interact with other pro-resolution mediators. H₂S modulates COX-2 expression in the gastrointestinal tract, which plays a crucial role in resolution of inflammation and injury (3, 49, 56, 61, 70, 71). Recently reported work from Brancaleone et al. (8) demonstrates an important interaction between H₂S and AnxA-1. AnxA-1 is a well-characterized anti-inflammatory and pro-resolution mediator that has also been shown to contribute to gastrointestinal integrity and repair (36, 41). Brancaleone et al. (8) observed that an H₂S donor (NaHS, 10-100 µM) elicited intense mobilization of AnxA-1 from the cytosol to the membrane of human neutrophils. It also markedly suppressed IL-1-induced leukocyte adhesion and emigration in mesenteric venules of wild type, but not AnxA1-deficient mice. There were also effects of endogenous AnxA1 on H₂S synthesis. Thus, mice deficient of AnxA1 displayed marked up-regulation of cystathionine β synthase (CBS) and cystathionine γ -lyase (CSE) in a variety of tissues as compared with wild-type mice. Moreover, H₂S could down-regulate other inflammatory pathways in macrophages from wild-type mice (i.e., significant suppression of iNOS and COX-2 expression in lipopolysaccharide-stimulated bone marrow-derived macrophages) but not in macrophages obtained from AnxA1-deficient mice. The authors concluded that these data demonstrate interlinks between the H₂S and AnxA1

FIG. 3. Resolution of inflammation. Inflammatory reactions to injury or infection are largely coordinated by soluble mediators (pro-inflammatory and antiinflammatory). In the early stages of an inflammatory reaction, the proinflammatory mediators predominate, resulting in the recruitment of inflammatory cells (such as neutrophils) and platelets to the site of injury/infection. Monocyte recruitment follows, with the subsequent transformation to macrophages, which are crucial for tissue repair. In part driven by anti-inflammatory mediators such as H₂S, the infiltrated neutrophils begin to undergo apoptosis, which causes macrophages to shift their phenotype from pro-inflammatory to anti-inflammatory. Engulfment of apoptotic neutrophils (polymorphonuclear leukocytes [PMN]) by macrophages is a key event in resolution of inflammation.



pathways that are likely very important in the resolution of inflammation.

The importance of H₂S in promoting resolution of inflammation and repair of injury has been clearly demonstrated in animal models of gastrointestinal inflammation and ulceration. In rat and mouse models of gastric ulcer, administration of H₂S-generating agents has been shown to significantly accelerate ulcer healing, while inhibition of endogenous H₂S synthesis was associated with impaired ulcer healing (58, 60). Administration of the precursor for H₂S synthesis, L-cysteine, promoted ulcer healing in the rat at doses that did not affect tissue glutathione levels (60). One of the significant limitations to the use of NSAIDs for treatment of inflammatory conditions is their ability to retard the healing of gastrointestinal ulcers. As shown in Fig. 3, this can be shown in a mouse model of gastric ulceration in which NSAIDs such as naproxen and celecoxib significantly impaired ulcer healing. However, an H₂S-releasing derivative of naproxen (ATB-346) not only did not impair ulcer healing, it significantly accelerated the healing process (58); thus, enhanced repair occurred despite marked suppression of COX-2 activity in the damaged tissue. Similar effects have been demonstrated previously in the rat with an H_2S -releasing derivative of mesalamine (19, 56). The mechanism underlying the enhancement of ulcer healing by H₂S is not clear, but may be related, at least in part, to its vasodilatory activity, its ability to enhance COX-2 expression, and its reported ability to promote angiogenesis (40).

Further evidence for an important role of H_2S in resolution of inflammation and repair of injury comes from studies of experimental colitis in rodents. When the colonic mucosa is inflamed, there is a marked increase in the capacity of the tissue to produce H_2S (20, 58). Most of the H_2S in this context is produced via CBS, as is the case in non-inflamed colon (35), but synthesis via CSE is also significant (61). Preliminary data suggested that there is also a marked elevation of colonic H_2S synthesis via other enzymatic pathways (i.e., non-CSE and non-CBS) when the colon is inflamed. Inhibition of H_2S synthesis once colitis is established led to a marked worsening of the inflammation, with perforation of the bowel wall and death occurring in most animals within a few days (61). In the rats that survived a week-long treatment with an inhibitor of H_2S synthesis, the colonic damage was significantly worse than that observed in vehicle-treated controls. Interestingly, there was a marked thickening of the smooth muscle in the colon in the rats treated with an inhibitor of H_2S synthesis (61).

Additional evidence that H₂S could promote resolution of colitis came from studies in which H₂S donors were administered to rats or mice with colitis (19, 61, 68). Irrespective of the H₂S donor used, a significant reduction of the severity of colitis was observed, with a marked inhibition of granulocyte infiltration into the colonic tissue (19, 61). The latter observation is important, since much of the tissue injury associated with colitis is likely produced by infiltrating granulocytes. In these studies, the H₂S-generating agents significantly reduced colonic expression of TNF α (protein and mRNA); in other studies, they also reduced expression of interferon- γ , IL-1 β , and iNOS (19, 61, 68). Consistent with these findings, allyl trisulfide, a garlic-derived substance that releases H₂S (7), was shown to suppress TNF α expression and NF- κ B activation in colonic biopsies from patients with ulcerative colitis (6).

H₂S in Novel Therapeutics

The ability of H₂S to enhance gastrointestinal resistance to injury, to promote repair of damaged tissue, and to reduce mucosal inflammation make it an attractive substance to exploit in designing novel drugs for treatment of gastrointestinal injury and inflammation (10, 54). We have synthesized and characterized the effects of an H₂S-releasing derivative of mesalamine in animal models of colitis (mesalamine is the first-line therapy for colitis) and have observed a significant enhancement of anti-inflammatory activity as compared to the parent drug (19, 56). H₂S-releasing derivatives of several NSAIDs have been assessed in animal models, with the consistent finding of greatly reduced gastrointestinal toxicity and, in some cases, enhanced anti-inflammatory activity (30, 54, 58, 59). An H₂S-releasing derivative of naproxen has been particularly well characterized. The anti-inflammatory activity of the compound, called ATB-346, is comparable to that of the parent drug (58). However, even at exceptionally high doses

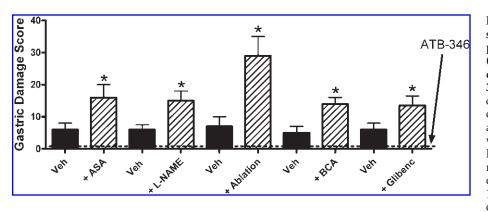


FIG. 4. Acute damage to the stomach in rats treated with naproxen or an equimolar dose ($60 \mu mol/kg$) of an H₂S-releasing derivative of naproxen (ATB-346). In rats pretreated with vehicle, naproxen induced a low level of gastric damage, while no damage was observed in rats treated with ATB-346 ($n \ge 5$ per group). Pretreatment with low-dose aspirin (10 mg/kg; ASA), an inhibitor of nitric oxide synthase (L-NAME; 15 mg/kg), ablation with capsaicin of sensory afferent nerves, pre-

treatment with an inhibitor of cystathionine β -synthase (β -cyanoalanine [BCA], 50 mg/kg) or an antagonist of ATP-sensitive K⁺ channels (glibenclamide; 10 mg/kg) each significantly increased the severity of naproxen-induced gastric damage (*p < 0.05 vs. corresponding vehicle-treated group). However, ATB-346 did not produce significant gastric damage in any of the rats receiving these treatments (dotted line). Adapted from previously published data (55).

(100 times the human dose on a per kilogram basis), ATB-346 caused negligible gastric damage in healthy rats. While impressive, studies performed in healthy rodents may not serve as a good predictor of how an anti-inflammatory drug will behave in humans with diseases such as osteoarthritis and rheumatoid arthritis alongside other co-morbidities for gastrointestinal damage. To more rigorously assess the gastric safety of ATB-346, studies were performed in rats in which gastric mucosal defense was significantly compromised (58). This was achieved by interfering with the production of mediators known to contribute to mucosal defense (e.g., nitric oxide, hydrogen sulfide), co-administering another drug that can damage the stomach (e.g., low-dose aspirin, frequently co-administered with NSAIDs in a clinical setting), blocking receptors believed to contribute to mucosal defense (e.g., glibenclamide to block ATP-sensitive K⁺ channels), or ablating sensory afferent nerves (via neonatal administration of capsaicin), which play a crucial role in mucosal responses to luminal irritants (55). As illustrated in Fig. 4, naproxen produced a low level of damage in control rats at the dose tested (60 μ mol/kg). However, with each approach to compromising mucosal defense, the extent of injury induced by naproxen was significantly increased (two- to sixfold). In contrast, ATB-346 at an equimolar dose did not produce significant damage in controls or in mucosal defense-compromised animals. Particularly interesting is the observation that ATB-346 was still gastric safe in rats pretreated with glibenclamide. Many actions of H₂S have been attributed to activation of ATPsensitive K⁺ channels (14, 62, 74). This observation suggests that the mechanism underlying the gastric tolerability of ATB-346 is unrelated to activation of ATP-sensitive K⁺ channels. It is also noteworthy that ATB-346 suppressed gastric prostaglandin synthesis as effectively as equimolar doses of naproxen (58). Thus, the compound can compensate for the reduced mucosal resistance to injury that occurs when prostaglandin synthesis is markedly reduced (presumably because H₂S can exert many of the same effects, in terms of mucosal defense, as prostaglandins do).

Another major clinical concern with respect to the use of NSAIDs is their ability to interfere with the healing of ulcers (55). This effect is likely related to suppression of COX-2 activity by the NSAIDs. COX-2–derived prostaglandin synthesis by cells at the ulcer margin is critically important for re-

epithelialization and angiogenesis (55). As shown in Fig. 5, naproxen and celecoxib (nonselective COX inhibitor and selective COX-2 inhibitor, respectively) each significantly impaired the healing of gastric ulcers in mice when administered twice daily over a 4-day period. In sharp contrast, the H₂Sreleasing derivative of naproxen (ATB-346) significantly accelerated the healing of pre-existing gastric ulcers (58). This is consistent with previous reports that L-cysteine and H₂Sgenerating agents could accelerate experimental ulcer healing, while inhibitors of H₂S synthesis impaired ulcer healing (56, 60). The beneficial effects of H₂S on ulcer healing may be attributable to the stimulatory effects of H₂S on angiogenesis (40).

From a clinical perspective, the main focus in terms of the gastrointestinal toxicity of NSAIDs is the stomach and

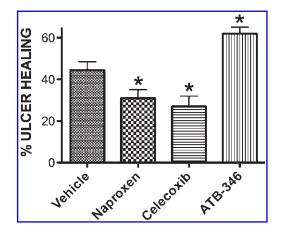


FIG. 5. Experimental gastric ulcer healing with ATB-346, an H₂S-releasing derivative of naproxen. The extent of healing of gastric ulcers in mice treated twice daily over a 4-day period with one of three NSAIDs or vehicle was examined. Gastric ulcers were induced by brief serosal application of acetic acid (55). Beginning 3 days later, the mice began treatment with naproxen, ATB-346 (each at 60 μ mol/kg), celecoxib (at 30 μ mol/kg), or vehicle. The ulcer area at the end of the treatment period was compared to that in mice euthanized 3 days after ulcer induction (no drug treatment), and the "percent ulcer healing" was calculated. *p<0.05 versus the vehicle-treated group ($n \ge 5$ per group). Adapted from previously published data (55).

H₂S AND RESOLUTION OF INFLAMMATION

proximal duodenum. This is largely due to the relative ease of endoscopically viewing these regions. However, it is becoming increasingly clear that NSAIDs frequently produce significant injury in the small intestine. Indeed, the jejunum and ileum may be the major sites of NSAID-induced bleeding (55, 57). Damage to these regions induced by NSAIDs is determined by a number of factors, but most important is the enterohepatic circulation of the NSAIDs, leading to repeated exposure of the epithelium to the NSAIDs and bile, and also to marked changes in the numbers and type of bacteria in the small intestine (57). The H₂S-releasing derivative of naproxen (ATB-346), when given twice daily to rats for 5 days, did not produce detectable small intestinal damage (58). This was in marked contrast to the parent drug, naproxen, which elicited widespread ulceration and bleeding in the small intestine.

Bacteria-Derived H₂S: A Modulator of Inflammation or Mucosal Function?

Many of the species of bacteria residing in the human gastrointestinal tract are capable of producing H₂S. Some early studies suggested that the concentrations of H₂S in the lumen of the gut were extremely high relative to levels that occur in the body (32, 33). Largely based on these data, and observations of adverse effects of such concentrations of H₂S on colonocyte function (5), roles for H_2S in the pathogenesis of inflammatory bowel disease and colon cancer were suggested (4, 45, 46). However, this hypothesis has been challenged (25, 26, 38, 42), as has the notion that there are millimolar concentrations of free H₂S in the lumen of the gut (27, 28, 38, 42, 43). Most of the H_2S that is produced in the lumen of the intestine is bound to fecal material and therefore not available to diffuse through the epithelium. The H₂S that is available, likely in micromolar concentrations, is absorbed and rapidly metabolized (51). Indeed, efficient detoxification occurs via a number of enzymes present in the mucosa, and no impairment of these detoxification systems has been detected in patients with ulcerative colitis or Crohn's disease (42). Based on studies utilizing a rat model of colitis (dextran sodium sulfate), Furne et al. (21) concluded that "excessive H₂S production" did not contribute to tissue injury.

Even before reaching the mucosa, however, there is substantial metabolism of H₂S in the mitochondria of enterocytes and colonocytes. Virtually all H₂S that crosses the apical membrane of enterocytes and colonocytes is rapidly oxidized to thiosulfate (22, 28). This is accomplished mainly by a mitochondrial enzyme, sulfide quinone reductase, which rapidly consumes H₂S, thereby providing energy to the cell and keeping H₂S concentrations at nontoxic levels (26) (Fig. 6). This ancestral capacity, predating photosynthesis, is common to organisms living in low-light, low-oxygen conditions (52). As mentioned above, efficient detoxification also occurs via a number of mucosal enzymes (52). The epithelium could therefore be viewed not only as a physical barrier, protecting organisms from potentially harmful substances in the lumen of the digestive tract, but also a metabolic barrier providing additional protection. When there is an intact, healthy epithelium, luminally produced H₂S likely has little, if any, effect on mucosal function. However, in cases in which the epithelium is dysfunctional or damaged, it remains possible that H₂S produced by bacteria could exert significant effects on several aspects of mucosal function, including secretion (via

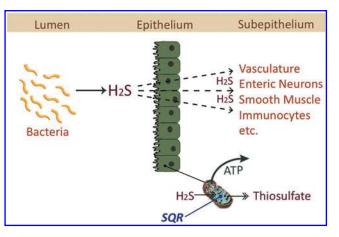


FIG. 6. The colonic epithelium as a metabolic barrier to bacteria-derived H_2S . Many species of bacteria can produce H_2S but most is bound to fecal material. The small amount that is free to diffuse across the epithelium is rapidly metabolized, primarily via mitochondrial sulfide quinone reductase (SQR) to thiosulfate, generating adenosine triphosphate (ATP) in the process. Thus, H_2S is an important energy source for colonocytes. When the epithelium is damaged or dysfunctional, more H_2S may escape metabolism and reach the subepithelial space, where it can affect many functions, including epithelial secretion, blood flow, smooth muscle contractility, and mucosal defense.

effects on enteric neurons) (47), pain sensation (14), blood flow (18, 63), and smooth muscle contractility (29, 63). Indeed, an impaired colonic "barrier" to diffusion of H_2S from the lumen may explain in part the marked beneficial effects of H_2S donors, when administered by enema, in models of colitis (19, 61).

As already reviewed, colonic production of H₂S is markedly increased when the mucosa is inflamed (20, 61). Because measurements of H₂S in this context are generally performed in vitro, and because of the capacity of some colonic bacteria to produce H₂S, we performed a series of studies to determine if some portion of what we measure as "colonic H₂S synthesis" is actually bacterial H₂S synthesis (20). These studies involved the use of germ-free mice and mice colonized with Altered Schaedler flora. There was no difference in colonic H₂S synthesis between these two groups of mice, indicating that any bacterial contribution (in the colonized mice) was negligible (Fig. 7). We also measured tissue and fecal H₂S production from healthy mice and mice with colitis induced by trinitrobenzene sulfonic acid. Colonic H₂S synthesis was markedly increased in parallel with the severity of colitis and the extent of granulocyte infiltration (Fig. 8), but neutrophils (the main infiltrating granulocytes) were not a significant source of H₂S synthesis. Taken together, these studies clearly demonstrated that the H₂S synthesis measured using the in vitro zinctrapping method was derived from the colonic tissue itself, rather than from any bacteria adherent to the tissue samples. On the other hand, certain bacteria may be able to modulate colonic H₂S synthesis. For example, butyrate, which is produced by Bifidobacteria and Faecalibacterium prausnitzii, has been shown to regulate H₂S production, at least in transformed colonocytes (WiDr cells) (11). Butyrate increased expression of CBS and CSE, the major enzymatic sources of H₂S in these cells.

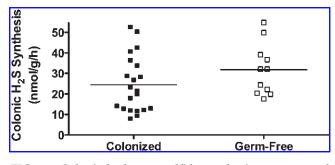


FIG. 7. Colonic hydrogen sulfide synthesis, as measured by the zinc acetate–trapping method, does not include significant "contamination" by bacterial H_2S synthesis. Mice that were colonized by Altered Schaedler Flora were compared with mice that were raised germ-free. There was considerable variation from mouse to mouse in terms of colonic H_2S synthesis, but there was no significant difference between the colonized and germ-free mice. Adapted from previously published data (20).

Conclusions and Future Directions

The contribution of H_2S as a modulator of inflammation is becoming more clear, but studies of this gaseous mediator continue to be limited by the lack of highly selective inhibitors of its synthesis and simple methods for measuring its production *in vivo*. Pathways of synthesis of H_2S other than via CSE and CBS are becoming evident and represent an important area for future research.

Novel therapeutic agents that release H_2S and exhibit significant anti-inflammatory and/or gastrointestinal mucosal protective effects look promising in preclinical studies. The mechanisms through which H_2S increases resistance to mucosal injury, promotes repair of injury, and accelerates resolution of inflammation remain incompletely understood. Evaluation of H_2S -releasing drugs in a clinical setting will provide insight as to whether or not the exploitation of H_2S as a therapeutic agent will live up to the promise.

The ability of many enteric bacteria to produce H_2S and the possibility that bacterially derived H_2S could affect mucosal function are intriguing. The intestinal epithelium appears to

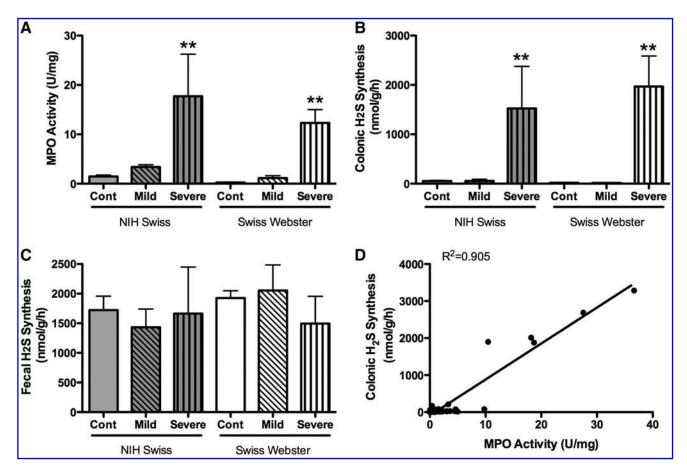


FIG. 8. Colonic hydrogen sulfide synthesis (as measured by the zinc acetate–trapping method) is markedly up-regulated during injury/inflammation, and there is negligible bacterial contribution. Colitis was induced via intrarectal administration of trinitrobenzene sulfonic acid (20). (A) Colonic myeloperoxidase (MPO) activity, a biochemical marker of granulocyte infiltration. In both strains of mice examined, MPO activity increased with the severity of the colitis. (B) Colonic H₂S synthesis, which similarly increased in parallel with the severity of colitis. (C) In contrast, fecal hydrogen sulfide synthesis did not differ between healthy mice and mice with mild or severe colitis, suggesting negligible contribution of bacteria to what was measured as "colonic" H₂S. (D) There was a strong correlation between the extent of inflammation (MPO activity) and colonic H₂S synthesis. There was no such correlation between fecal H₂S synthesis and colonic MPO activity. **p < 0.01 versus the corresponding control (cont) group. Adapted from previously published data (20).

H₂S AND RESOLUTION OF INFLAMMATION

act as a metabolic barrier to diffusion of significant concentrations of H_2S into the mucosa, at least when the epithelium is healthy. On the other hand, H_2S is an important metabolic fuel for enterocytes. Given the widespread actions that H_2S can exert on various cell types in the subepithelial compartment (e.g., blood vessels, enteric nerves, smooth muscle cells, resident and infiltrating immunocytes), it is important to determine the extent to which bacterial H_2S can "escape" epithelial and mucosal inactivation in certain circumstances and disease conditions.

Acknowledgments

This work is supported by grants from the Canadian Institutes of Health Research and the Crohn's and Colitis Foundation of Canada.

Author Disclosure Statement

Dr. Wallace holds shares in Antibe Therapeutics, a company developing H_2 S-releasing anti-inflammatory drugs. No conflicts of interest exist for the other authors.

References

- Abe K and Kimura H. The possible role of hydrogen sulfide as an endogenous neuromodulator. *J Neurosci* 16: 1066–1071, 1996.
- Andruski B, McCafferty DM, Ignacy T, Millen B, and McDougall JJ. Leukocyte trafficking and pain behavioral responses to a hydrogen sulfide donor in acute monoarthritis. *Am J Physiol Regul Integr Comp Physiol* 295: R814-R820, 2008.
- Asfaha S, MacNaughton WK, Appleyard CB, Chadee K, and Wallace JL. Persistent epithelial dysfunction and bacterial translocation after resolution of intestinal inflammation. *Am J Physiol* 281: G635-G644, 2001.
- Attene-Ramos MS, Wagner ED, Plewa MJ, et al. Evidence that hydrogen sulfide is a genotoxic agent. Mol Cancer Res 4: 9–14, 2006.
- Babidge W, Millard S, and Roediger W. Sulfides impair short chain fatty acid beta-oxidation at acyl-CoA dehydrogenase level in colonocytes: implications for ulcerative colitis. *Mol Cell Biochem* 181: 117–124, 1998.
- Bai AP, Ouyang Q, and Hu RW. Diallyl trisulfide inhibits tumor necrosis factor-alpha expression in inflamed mucosa of ulcerative colitis. *Dig Dis Sci* 50: 1426–1431, 2005.
- Benavides GA, Squadrito GL, Mills RW, et al. Hydrogen sulfide mediates the vasoactivity of garlic. Proc Natl Acad Sci USA 104: 17977–17982, 2007.
- Brancaleone V, Sampaio A, Cirino C, Flower RJ, and Perretti M. A novel cross-talk in resolution: H₂S activates the annexin A1 pathway. *Inflammation Res*, in press.
- Bucci M, Papapetropoulos A, Vellecco V, Zhou Z, Pyriochou A, Roussos C, Roviezzo F, Brancaleone V, and Cirino G. Hydrogen sulfide is an endogenous inhibitor of phosphodiesterase activity. *Arterioscler Thromb Vasc Biol* 30: 1998–2004, 2010.
- Caliendo G, Cirino G, Santagada V, and Wallace JL. Synthesis and biological effects of hydrogen sulfide (H₂S): development of H₂S-releasing drugs as pharmaceuticals. J Med Chem 53: 6275–6286, 2010.
- Cao Q, Zhang L, Yang G, Xu C, and Wang R. Butyratestimulated H₂S production in colon cancer cells. *Antioxid Redox Signal* 12: 1101–1109, 2010.
- 12. Chen YH, Wang PP, Wang XM, He YJ, Yao WZ, Qi YF, and Tang CS. Involvement of endogenous hydrogen sulfide in

cigarette smoke-induced changes in airway responsiveness and inflammation of rat lung. *Cytokine* 53: 334–341, 2011.

- Cunha TM, Dal-Secco D, Verri WA Jr, Guerrero AT, Souza GR, Vieira SM, Lotufo CM, Neto AF, Ferreira SH, and Cunha FQ. Dual role of hydrogen sulfide in mechanical inflammatory hypernociception. *Eur J Pharmacol* 590: 127–135, 2008.
- Distrutti E, Sediari L, Mencarelli A, *et al.* Evidence that hydrogen sulfide exerts antinociceptive effects in the gastrointestinal tract by activating K_{ATP} channels. *J Pharmacol Exp Ther* 316: 325–335, 2006.
- 15. Distrutti E, Sediari L, Mencarelli A, Renga B, Orlandi S, Russo G, Caliendo G, Santagada V, Cirino G, Wallace JL, and Fiorucci S. 5-Amino-2-hydroxybenzoic acid 4-(5thioxo-5H-[1,2]dithiol-3yl)-phenyl ester (ATB-429), a hydrogen sulfide-releasing derivative of mesalamine, exerts antinociceptive effects in a model of postinflammatory hypersensitivity. J Pharmacol Exp Ther 319: 447–458, 2006.
- Dufton N and Wallace JL. Phenotypic differences in hydrogen sulfide synthesis and signaling in primary macrophages. *Inflammation Res* 60: 121, 2011.
- 17. Ekundi-Valentim E, Santos KT, Camargo EA, Denadai-Souza A, Teixeira SA, Zanoni CI, Grant AD, Wallace J, Muscará MN, and Costa SK. Differing effects of exogenous and endogenous hydrogen sulphide in carrageenan-induced knee joint synovitis in the rat. *Br J Pharmacol* 159: 1463–1474, 2010.
- Fiorucci S, Antonelli E, Distrutti E, Mencarelli A, Orlandi S, Zanardo R, Renga B, Rizzo G, Morelli A, Cirino G, and Wallace JL. Inhibition of hydrogen sulfide generation contributes to gastric injury caused by anti-inflammatory nonsteroidal drugs. *Gastroenterology* 129: 1210–1224, 2005.
- Fiorucci S, Mencarelli A, Caliendo G, Santagada V, Distrutti E, Santucci L, Cirino G, and Wallace JL. Enhanced activity of a hydrogen-sulfide releasing mesalamine derivative (ATB-429) in a mouse model of colitis. *Br J Pharmacol* 150: 996–1002, 2007.
- Flannigan KL, McCoy KD, and Wallace JL. Eukaryotic and prokaryotic contributions to colonic hydrogen sulfide synthesis. *Am J Physiol Gastrointest Liver Physiol* 301: G188–G193, 2011.
- Furne JK, Suarez FL, Koenig T, DeMaster E, and Levitt MD. Oxidation of hydrogen sulfide and methanethiol to thiolsulfate by rat tissues: a specialized function of the colonic mucosa. *Biochem Pharmacol* 62: 255–259, 2001.
- Goubern M, Andriamihaja M, Nübel T, *et al.* Sulfide, the first inorganic substrate for human cells. *FASEB J* 21: 1699–1706, 2007.
- 23. Han W, Dong Z, Dimitropoulou C, Su Y. Hydrogen sulfide ameliorates tobacco smoke-induced oxidative stress and emphysema in mice. *Antioxid Redox Signal* 15: 2121–2134, 2011.
- 24. Hosoki R, Matsuki N, and Kimura H. The possible role of hydrogen sulfide as an endogenous smooth muscle relaxant in synergy with nitric oxide. *Biochem Biophys Res Commun* 237: 527–531, 1997.
- 25. Jorgensen J and Mortensen PB. Hydrogen sulfide and colonic epithelial metabolism: implications for ulcerative colitis. *Dig Dis Sci* 46: 1722–1732, 2001.
- Lagoutte E, Mimoun S, Andriamihaja M, Chaumontet C, Blachier F, and Bouillaud F. Oxidation of hydrogen sulfide remains a priority in mammalian cells and causes reverse electron transfer in colonocytes. *Biochim Biophys Acta* 1797: 1500–1511, 2010.

- Levine J, Ellis CJ, Furne JK, *et al.* Fecal hydrogen sulfide production in ulcerative colitis. *Am J Gastroenterol* 93: 83–87, 1998.
- Levitt MD, Springfield J, Furne J, et al. Physiology of sulfide in the rat colon: use of bismuth to assess colonic sulfide production. J Appl Physiol 92: 1655–1660, 2002.
- 29. Li L, Rose P, and Moore PK. Hydrogen sulfide and cell signaling. *Annu Rev Pharmacol Toxicol* 51:169–187, 2011.
- Li L, Rossoni G, Sparatore A, Lee LC, Del Soldato P, and Moore PK. Anti-inflammatory and gastrointestinal effects of a novel diclofenac derivative. *Free Radic Biol Med* 42: 706– 719, 2007.
- 31. Liu H, Bai XB, Shi S, and Cao YX. Hydrogen sulfide protects from intestinal ischaemia-reperfusion injury in rats. *J Pharm Pharmacol* 61: 207–212, 2009.
- 32. Macfarlane GT, Gibson GR, and Cummings JH. Comparison of fermentation reactions in different regions of the human colon. *J Appl Bacteriol* 72: 57–64, 1992.
- 33. Magee EA, Richardson CJ, Hughes R, *et al.* Contribution of dietary protein to sulfide production in the large intestine: an in vitro and a controlled feeding study in humans. *Am J Clin Nutr* 72: 1488–1494, 2000.
- Mariggio MA, Minunno V, Riccardi S, Santacroce R, De Rinaldis P, and Fumarulo R. Sulfide enhancement of PMN apoptosis. *Immunopharmacol Immunotoxicol* 20: 399–408, 1998.
- Martin GR, McKnight GW, Dicay M, Coffin CS, Ferraz JG, and Wallace JL. Hydrogen sulfide synthesis in the rat and mouse gastrointestinal tract. *Dig Liver Dis* 42: 103–109, 2010.
- Martin GR, Perretti M, Flower RJ, and Wallace JL. Annexin-1 modulates repair of gastric mucosal injury. *Am J Physiol Gastrointest Liver Physiol* 294: G764-G769, 2008.
- 37. Matsunami M, Tarui T, Mitani K, Nagasawa K, Fukushima O, Okubo K, Yoshida S, Takemura M, and Kawabata A. Luminal hydrogen sulfide plays a pronociceptive role in mouse colon. *Gut* 58: 751–671, 2009.
- Moore J, Babidge W, Millard S, et al. Colonic luminal hydrogen sulfide is not elevated in ulcerative colitis. Dig Dis Sci 43: 162–165, 1998.
- 39. Oh GS, Pae HO, Lee BS, Kim BN, Kim JM, Kim HR, Jeon SB, Jeon WK, Chae HJ, and Chung HT. Hydrogen sulfide inhibits nitric oxide production and nuclear factor-kB via heme oxygenase-1 expression in RAW264.7 macrophages stimulated with lipopolysaccharide. *Free Radic Biol Med* 41: 106–119, 2006.
- Papapetropoulos A, Pyriochou A, Altaany Z, Yang G, Marazioti A, Zhou Z, Jeschke MG, Branski LK, Herndon DN, Wang R, and Szabó C. Hydrogen sulfide is an endogenous stimulator of angiogenesis. *Proc Natl Acad Sci USA* 106: 21972–21977, 2009.
- 41. Perretti M and D'Acquisto F. Annexin A1 and glucocorticoids as effectors of the resolution of inflammation. *Nat Rev Immunol* 9: 62–70, 2009.
- Picton R, Eggo MC, Langman MJ, et al. Impaired detoxification of hydrogen sulfide in ulcerative colitis? *Dig Dis Sci* 52: 373–378, 2007.
- Pitcher MC, Beatty ER, and Cummings JH. The contribution of sulphate reducing bacteria and 5-aminosalicylic acid to faecal sulphide in patients with ulcerative colitis. *Gut* 46: 64– 72, 2000.
- Reiffenstein RJ, Hulbert WC, and Roth SH. Toxicology of hydrogen sulfide. *Annu Rev Pharmacol Toxicol* 32:109–134, 1992.
- 45. Roediger WE. The colonic epithelium in ulcerative colitis: an energy-deficiency disease? *Lancet* 2: 712–715, 1980.

- Roediger WE, Duncan A, Kapaniris O, *et al.* Sulphide impairment of substrate oxidation in rat colonocytes: a biochemical basis for ulcerative colitis? *Clin Sci (Lond)* 85: 623– 627, 1993.
- 47. Schicho R, Krueger D, Zeller F, *et al.* Hydrogen sulfide is a novel prosecretory neuromodulator in the guinea-pig and human colon. *Gastroenterology* 131: 1542–1552, 2006.
- Searcy DG and Lee SH. Sulfur reduction by human erythrocytes. J Exp Zool 282: 310–322, 1998.
- 49. Serhan CN, Brain SD, Buckley C, Gilroy DW, Haslett C, O'Neill LA, Perretti M, Rossi AG, and Wallace JL. Resolution of inflammation: state of the art, definitions and terms. *FASEB J* 21: 325–332, 2007.
- Shibuya N, Mikami Y, Kimura Y, Nagahara N, and Kimura H. Vascular endothelium expresses 3-mercaptopyruvate sulfurtransferase and produces hydrogen sulfide. *J Biochem* 146: 623–626, 2009.
- Suarez F, Furne J, Springfield J, et al. Production and elimination of sulfur-containing gases in the rat colon. Am J Physiol 274: G727-G733, 1998.
- 52. Theissen U, Hoffmeister M, Grieshaber M, and Martin W. Single eubacterial origin of eukaryotic sulfide:quinone oxidoreductase, a mitochondrial enzyme conserved from the early evolution of eukaryotes during anoxic and sulfidic times. *Mol Biol Evol* 20: 1564–1574, 2003.
- Tyagi N, Moshal KS, Sen U, Vacek TP, Kumar M, Hughes WM, Kundu S, and Tyagi SC. H₂S protections against methionine-induced oxidative stress in rain endothelial cells. *Antiox Redox Signal* 11: 25–33, 2009.
- 54. Wallace JL. Hydrogen sulfide-releasing anti-inflammatory drugs. *Trends Pharmacol Sci* 28: 501–505, 2007.
- 55. Wallace JL. Prostaglandins, NSAIDs and mucosal defence. Why doesn't the stomach digest itself? *Physiol Reviews* 88: 1547–1565, 2008.
- Wallace JL. Physiological and pathophysiological roles of hydrogen sulfide in the gastrointestinal tract. *Antioxid Redox Signal* 12: 1125–1133, 2010.
- 57. Wallace JL. NSAID gastropathy and enteropathy: distinct pathogenesis likely necessitates distinct prevention strategies. *Br J Pharmacol*, in press.
- Wallace JL, Caliendo G, Santagada V, and Cirino G. Markedly reduced toxicity of a hydrogen sulfide-releasing derivative of naproxen (ATB-346). *Br J Pharmacol* 159: 1236– 1246, 2010.
- Wallace JL, Caliendo G, Santagada V, Cirino G, and Fiorucci S. Gastrointestinal safety and anti-inflammatory effects of a hydrogen sulfide-releasing diclofenac derivative in the rat. *Gastroenterology* 132: 261–271, 2007.
- Wallace JL, Dicay M, McKnight W, and Martin GR. Hydrogen sulfide enhances ulcer healing. *FASEB J* 2007; 21: 4070–4076.
- Wallace JL, Vong L, McKnight W, Dicay M, and Martin GR. Endogenous and exogenous hydrogen sulphide promotes resolution of colitis in rats. *Gastroenterology* 137: 569–578, 2009.
- 62. Wang L and Wallace JL. Hydrogen sulfide reduces cardioautonomic responses to gastric distention (GD) in an ATPsensitive potassium channel (KATP)-dependent manner. *Neurogastroenterology and Motility* 23(Suppl 1): 18, 2011.
- Wang R. Two's company, three's a crowd: can H₂S be the third endogenous gaseous transmitter? *FASEB J* 16: 1792– 1798j, 2002.
- 64. Whiteman M, Armstrong JS, Chu SH, Siau JL, Wong BS, Cheung NS, Halliwell B, and Moore PK. The novel

H₂S AND RESOLUTION OF INFLAMMATION

neuromodulator hydrogen sulfide: an endogenous peroxynitrite scavenger? J Neurochem 90: 765–768, 2004.

- 65. Whiteman M, Cheung NS, Zhu YZ, Chu SH, Siau JL, Wong BS, Armstrong JS, and Moore PK. Hydrogen sulphide: a novel inhibitor of hypochlorous acid-mediated oxidative damage in the brain? *Biochem Biophys Res Commun* 326: 794–798, 2005.
- 66. Whiteman M, Haigh R, Tarr JM, Gooding KM, Shore AC, and Winyard PG. Detection of hydrogen sulfide in plasma and knee-joint synovial fluid from rheumatoid arthritis patients: relation to clinical and laboratory measures of inflammation. *Ann NY Acad Sci* 1203: 146–150, 2010.
- 67. Whiteman M, Li L, Rose P, Tan CH, Parkinson DB, and Moore PK. The effect of hydrogen sulfide donors on lipopolysaccharide-induced formation of inflammatory mediators in macrophages. *Antioxid Redox Signal* 12: 1147–1154, 2010.
- Xu XM, Yu JP, He XF, Li JH, Yu LL, and Yu HG. Effects of garlicin on apoptosis in rat model of colitis. *World J Gastroenterol* 11: 4579–4582, 2005.
- Xu ZS, Wang XY, Xiao DM, Hu LF, Lu M, Wu ZY, and Bian JS. Hydrogen sulfide protects MC3T3-E1 osteoblastic cells against H₂O₂-induced oxidative damage – implications for the treatment of osteoporosis. *Free Radic Biol Med* 15: 1314– 1323, 2011.
- Zamuner SR, Bak AW, Devchand PR, and Wallace JL. Predisposition to colorectal cancer in rats with resolved colitis: role of COX-2-derived PGD₂. Am J Pathol 167: 1293–1300, 2005.
- Zamuner SR, Warrier N, Buret AG, MacNaughton WK, and Wallace JL. Cyclooxygenase 2 mediates post-inflammatory colonic secretory and barrier dysfunction. *Gut* 52: 1714–1720, 2003.
- Zanardo RCO, Brancaleone V, Distrutti E, Fiorucci S, Cirino G, and Wallace JL. Hydrogen sulphide is an endogenous modulator of leukocyte-mediated inflammation. *FASEB J* 20: 2118–2120, 2006.
- Zhao W and Wang R. H₂S-induced vasorelaxation and underlying cellular and molecular mechanisms. *Am J Physiol* 283: H474-H480, 2002.
- Zhao W, Zhang J, Lu Y, and Wang R. The vasorelaxant effects of H₂S as a novel endogenous gaseous K_{ATP} channel opener. *EMBO J* 20: 6008–6016, 2001.

- Zhong GZ, Chen RF, Cheng YQ, Tang CS, and Du JB. The role of hydrogen sulfide generation in the pathogenesis of hypertension in rats induced by inhibition of nitric oxide synthase. J Hypertens 21: 1879–1885, 2003.
- 76. Zuidema MY, Yang Y, Wang M, Kalogeris T, Liu Y, Meininger CJ, Hill MA, Davis MJ, and Korthuis RJ. Antecedent hydrogen sulfide elicits an anti-inflammatory phenotype in postischemic murine small intestine: role of BK channels. *Am J Physiol Heart Circ Physiol* 299: H1554– H1567, 2010.

Address correspondence to: Dr. John L. Wallace Farncombe Digestive Health Research Institute McMaster University 1280 Main Street West Hamilton, Ontario, L8S 4K1 Canada

E-mail: jwalla@mcmaster.ca

Date of first submission to ARS Central, October 19, 2011; date of acceptance, October 22, 2011.

Abbreviations Used
AnxA-1 = annexin-1
AMP = adenosine monophosphate
CBS = cystathionine-beta-synthase
COX = cyclooxygenase
CSE = cystathionine-gamma-lyase
IL = interleukin
iNOS = inducible nitric oxide synthase
NF- κ B = nuclear transcription factor- κ B
NSAID = nonsteroidal anti-inflammatory drug
$PGE_2 = prostaglandin E_2$
PMN = polymorphonuclear leukocyte
TNF = tumor necrosis factor

This article has been cited by:

- 1. Qing Xiao, Lidan Xiong, Jie Tang, Li Li, Li Li. 2021. Hydrogen Sulfide in Skin Diseases: A Novel Mediator and Therapeutic Target. Oxidative Medicine and Cellular Longevity 2021, 1-11. [Crossref]
- Pinghua Ou, Yali Wang, Cong Hao, Yongbo Peng, Li-yi Zhou. 2021. Naphthalimide-based a highly selective two-photon fluorescent probe for imaging of hydrogen sulfide in living cells and inflamed tissue of mouse model. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy 245, 118886. [Crossref]
- Haonan Li, Yuxi Fang, Junjie Yan, Xiangyu Ren, Chao Zheng, Bo Wu, Siyuan Wang, Zhanlin Li, Huiming Hua, Peng Wang, Dahong Li. 2021. Small-molecule fluorescent probes for H2S detection: Advances and perspectives. *TrAC Trends in Analytical Chemistry* 134, 116117. [Crossref]
- 4. Fei Sun, Jia-Hui Luo, Tian-Tian Yue, Fa-Xi Wang, Chun-Liang Yang, Shu Zhang, Xin-Qiang Wang, Cong-Yi Wang. 2021. The role of hydrogen sulphide signalling in macrophage activation. *Immunology* **162**:1, 3-10. [Crossref]
- 5. Robert E Click. 2021. Potential alteration of tumor microenvironments by β-mercaptoethanol. *Future Oncology* 17:3, 315-331. [Crossref]
- 6. Li Theng Ng, Li Fang Ng, Richard Ming Yi Tang, Diogo Barardo, Barry Halliwell, Philip Keith Moore, Jan Gruber. 2020. Lifespan and healthspan benefits of exogenous H2S in C. elegans are independent from effects downstream of eat-2 mutation. *npj Aging and Mechanisms of Disease* **6**:1. [Crossref]
- 7. Joana Claudio Pieretti, Carolina Victoria Cruz Junho, Marcela Sorelli Carneiro-Ramos, Amedea Barozzi Seabra. 2020. H2S- and NO-releasing gasotransmitter platform: A crosstalk signaling pathway in the treatment of acute kidney injury. *Pharmacological Research* 161, 105121. [Crossref]
- 8. Mehdi Najar, Johanne Martel-Pelletier, Jean-Pierre Pelletier, Hassan Fahmi. 2020. Mesenchymal Stromal Cell Immunology for Efficient and Safe Treatment of Osteoarthritis. *Frontiers in Cell and Developmental Biology* 8. . [Crossref]
- 9. Aleksandr Birg, Henry C. Lin, Nancy Kanagy. 2020. Portal Venous Flow Is Increased by Jejunal but Not Colonic Hydrogen Sulfide in a Nitric Oxide-Dependent Fashion in Rats. *Digestive Diseases and Sciences* **312**. [Crossref]
- Ali Mujtaba Shah, Jian Ma, Zhisheng Wang, Rui Hu, Xueying Wang, Quanhui Peng, Felix Kwame Amevor, Naqash Goswami. 2020. Production of Hydrogen Sulfide by Fermentation in Rumen and Its Impact on Health and Production of Animals. *Processes* 8:9, 1169. [Crossref]
- Nayara Braga Emidio, Stuart M. Brierley, Christina I. Schroeder, Markus Muttenthaler. 2020. Structure, Function, and Therapeutic Potential of the Trefoil Factor Family in the Gastrointestinal Tract. ACS Pharmacology & Translational Science 3:4, 583-597. [Crossref]
- 12. Yaroslav Pavlovskiy, Antonina Yashchenko, Oksana Zayachkivska. 2020. H2S Donors Reverse Age-Related Gastric Malfunction Impaired Due to Fructose-Induced Injury via CBS, CSE, and TST Expression. *Frontiers in Pharmacology* 11. . [Crossref]
- 13. Gerald Cohen. 2020. Immune Dysfunction in Uremia 2020. Toxins 12:7, 439. [Crossref]
- 14. Morgan A. Perkins, Kayleigh R. Barlow, Katelyn M. Dreux, Gregory S. Tschumper. 2020. Anchoring the hydrogen sulfide dimer potential energy surface to juxtapose (H 2 S) 2 with (H 2 O) 2. *The Journal of Chemical Physics* 152:21, 214306. [Crossref]
- 15. Wuyang Hua, Jian Zhao, Shaohua Gou. 2020. A naphthalimide derivative can release COS and form H 2 S in a light-controlled manner and protect cells against ROS with real-time monitoring ability. *The Analyst* 145:11, 3878-3884. [Crossref]
- 16. Flavia Sunzini, Susanna De Stefano, Maria Sole Chimenti, Sonia Melino. 2020. Hydrogen Sulfide as Potential Regulatory Gasotransmitter in Arthritic Diseases. *International Journal of Molecular Sciences* 21:4, 1180. [Crossref]
- 17. Silvia A. Coavoy-Sánchez, Soraia K. P. Costa, Marcelo N. Muscará. 2020. Hydrogen sulfide and dermatological diseases. *British Journal of Pharmacology* 177:4, 857-865. [Crossref]
- Emanuela G. Garattini, Bruna M. Santos, Daniele P Ferrari, Camila P Capel, Heloísa D.C. Francescato, Terezila M. Coimbra, Christie R.A. Leite-Panissi, Luiz G.S. Branco, Glauce C. Nascimento. 2019. Propargylglycine decreases neuro-immune interaction inducing pain response in temporomandibular joint inflammation model. *Nitric Oxide* 93, 90-101. [Crossref]
- 19. Ghaneya S Hassan, Gehan H Hegazy, Noha M Ibrahim, Samar H Fahim. 2019. New ibuprofen derivatives as H 2 S and NO donors as safer anti-inflammatory agents. *Future Medicinal Chemistry* 11:23, 3029-3045. [Crossref]
- 20. Ivan Kushkevych, Ol'ga Leščanová, Dani Dordević, Simona Jančíková, Jan Hošek, Monika Vítězová, Leona Buňková, Lorenzo Drago. 2019. The Sulfate-Reducing Microbial Communities and Meta-Analysis of Their Occurrence during Diseases of Small-Large Intestine Axis. *Journal of Clinical Medicine* 8:10, 1656. [Crossref]
- 21. Yaqing Jiang, Xiuru Ji, Changyu Zhang, Zhen Xi, Lu Sun, Long Yi. 2019. Dual-quenching NBD-based fluorescent probes for separate detection of H 2 S and Cys/Hcy in living cells. *Organic & Biomolecular Chemistry* 17:36, 8435-8442. [Crossref]

- 22. Haonan Li, Xiang Gao, Xiaofang Huang, Xianhua Wang, Shengtao Xu, Takahiro Uchita, Ming Gao, Jinyi Xu, Huiming Hua, Dahong Li. 2019. Hydrogen sulfide donating ent-kaurane and spirolactone-type 6,7-seco-ent-kaurane derivatives: Design, synthesis and antiproliferative properties. *European Journal of Medicinal Chemistry* 178, 446-457. [Crossref]
- 23. Laura Grasa, Leticia Abecia, Ainize Peña-Cearra, Sofia Robles, Elena Layunta, Eva Latorre, José Emilio Mesonero, Raquel Forcén. 2019. TLR2 and TLR4 interact with sulfide system in the modulation of mouse colonic motility. *Neurogastroenterology* & Motility 31:9. . [Crossref]
- 24. Xiao-jing Cheng, Jing-xue Gu, Yi-peng Pang, Jiao Liu, Ting Xu, Xin-rui Li, Yu-zhou Hua, Kelly A. Newell, Xu-Feng Huang, Yinghua Yu, Yi Liu. 2019. Tacrine–Hydrogen Sulfide Donor Hybrid Ameliorates Cognitive Impairment in the Aluminum Chloride Mouse Model of Alzheimer's Disease. ACS Chemical Neuroscience 10:8, 3500-3509. [Crossref]
- Zhen Zhang, Xin Fang, Xiawen Yang, Takahiko Mitsui, Yanru Huang, Zhimin Mao, Yong Huang, Masayuki Takeda, Jian Yao. 2019. Hydrogen sulfide donor NaHS alters antibody structure and function via sulfhydration. *International Immunopharmacology* 73, 491-501. [Crossref]
- 26. Weijuan Liu, Zongchang Li, Huimin Jia, Lixia Zhang, Weiwei He, Qingbo Meng. 2019. Shell surface sulfidation mediated the plasmonic response of Au@Ag NPs for colorimetric sensing of sulfide ions and sulfur. *Applied Surface Science* **481**, 678-683. [Crossref]
- 27. Hui-Bing Na, Xian-Fa Zhang, Zhao-Peng Deng, Ying-Ming Xu, Li-Hua Huo, Shan Gao. 2019. Large-Scale Synthesis of Hierarchically Porous ZnO Hollow Tubule for Fast Response to ppb-Level H 2 S Gas. ACS Applied Materials & Interfaces 11:12, 11627-11635. [Crossref]
- Yasemin Tekşen, Emine Kadıoğlu, Cengiz Koçak, Havva Koçak. 2019. Effect of Hydrogen Sulfide on Kidney Injury in Rat Model of Crush Syndrome. *Journal of Surgical Research* 235, 470-478. [Crossref]
- 29. Changyu Zhang, Qiang-Zhe Zhang, Kun Zhang, Lu-Yuan Li, Michael D. Pluth, Long Yi, Zhen Xi. 2019. Dual-biomarkertriggered fluorescence probes for differentiating cancer cells and revealing synergistic antioxidant effects under oxidative stress. *Chemical Science* 10:7, 1945-1952. [Crossref]
- 30. Fanxing Xu, Xiang Gao, Haonan Li, Shengtao Xu, Xu Li, Xu Hu, Zhanlin Li, Jinyi Xu, Huiming Hua, Dahong Li. 2019. Hydrogen sulfide releasing enmein-type diterpenoid derivatives as apoptosis inducers through mitochondria-related pathways. *Bioorganic Chemistry* 82, 192-203. [Crossref]
- 31. Joana Viegas, Ana Filipa Esteves, Elsa M. Cardoso, Fernando A. Arosa, Marco Vitale, Luís Taborda-Barata. 2019. Biological Effects of Thermal Water-Associated Hydrogen Sulfide on Human Airways and Associated Immune Cells: Implications for Respiratory Diseases. Frontiers in Public Health 7. [Crossref]
- 32. Tian-xiao Hu, You-liang Si, Yun Ruan, Xiu-jing Wang, Jia-qi Yao, Hui-ling Wang, Yao Xu, Xin Ni, Qing-ying Tan, Jing Wang. 2019. Reduced cystathionine-γ-lyase (CSE) expression is involved in high glucose induced MMP14 expression in adipocytes and adipose tissues. *Endocrine Journal* 66:11, 1029-1037. [Crossref]
- 33. François Blachier, Ayane de Sá Resende, Geovana da Silva Fogaça Leite, Aline Vasques da Costa, Antonio Herbert Lancha Junior. 2018. Colon epithelial cells luminal environment and physiopathological consequences: impact of nutrition and exercise. *Nutrire* 43:1. . [Crossref]
- 34. Shengnan Cheng, Ying Lu, Yuanyuan Li, Luyan Gao, Huaying Shen, Kai Song. 2018. Hydrogen sulfide inhibits epithelialmesenchymal transition in peritoneal mesothelial cells. *Scientific Reports* 8:1. . [Crossref]
- 35. Wei-Chih Lin, Wen-Yu Pan, Chen-Kao Liu, Wu-Xuan Huang, Hsiang-Lin Song, Kai-Sheng Chang, Meng-Ju Li, Hsing-Wen Sung. 2018. In situ self-spray coating system that can uniformly disperse a poorly water-soluble H2S donor on the colorectal surface to treat inflammatory bowel diseases. *Biomaterials* 182, 289-298. [Crossref]
- 36. Lauren K. Wareham, Hannah M. Southam, Robert K. Poole. 2018. Do nitric oxide, carbon monoxide and hydrogen sulfide really qualify as 'gasotransmitters' in bacteria?. *Biochemical Society Transactions* 46:5, 1107-1118. [Crossref]
- 37. Jiacheng Xiong, Lili Xia, Qili Huang, Jinxin Huang, Yueqing Gu, Peng Wang. 2018. Cyanine-based NIR fluorescent probe for monitoring H2S and imaging in living cells and in vivo. *Talanta* 184, 109-114. [Crossref]
- 38. Wang Jing, Wang Wan, Li Shuangyue, Han Yi, Zhang Ping, Meng Guoliang, Xiao Yujiao, Xie Liping, Wang Xin, Sha Jiahao, Chen Qi, Moore Philip K., Wang Rui, Xiang Wenpei, Ji Yong. 2018. Hydrogen Sulfide As a Potential Target in Preventing Spermatogenic Failure and Testicular Dysfunction. *Antioxidants & Redox Signaling* 28:16, 1447-1462. [Abstract] [Full Text] [PDF] [PDF Plus] [Supplementary Material]
- Enrico Gugliandolo, Roberta Fusco, Ramona D'Amico, Angela Militi, Giacomo Oteri, John L. Wallace, Rosanna Di Paola, Salvatore Cuzzocrea. 2018. Anti-inflammatory effect of ATB-352, a H2S –releasing ketoprofen derivative, on lipopolysaccharideinduced periodontitis in rats. *Pharmacological Research* 132, 220-231. [Crossref]

- 40. Chu K. Yao, Asaf Rotbart, Jian Z. Ou, Kourosh Kalantar-Zadeh, Jane G. Muir, Peter R. Gibson. 2018. Modulation of colonic hydrogen sulfide production by diet and mesalazine utilizing a novel gas-profiling technology. *Gut Microbes* 125, 1-13. [Crossref]
- 41. Jingao Yu, Jianming Guo, Weiwei Tao, Pei Liu, Erxin Shang, Zhenhua Zhu, Xiuhe Fan, Juan Shen, Yongqing Hua, Kevin Yue Zhu, Yuping Tang, Jin-ao Duan. 2018. Gancao-Gansui combination impacts gut microbiota diversity and related metabolic functions. *Journal of Ethnopharmacology* 214, 71-82. [Crossref]
- Khosrow Kashfi. 2018. The dichotomous role of H2S in cancer cell biology? Déjà vu all over again. *Biochemical Pharmacology* 149, 205-223. [Crossref]
- 43. Ayako Ujike, Tomoki Kuraishi, Soichiro Yamaguchi, Ryota Eguchi, Taisuke Kitano, Jumpei Kamise, Shigeo Ito, Ken-ichi Otsuguro. 2018. IL-1β augments H 2 S-induced increase in intracellular Ca 2+ through polysulfides generated from H 2 S/NO interaction. *European Journal of Pharmacology* 821, 88-96. [Crossref]
- 44. John L. Wallace, Jean-Paul Motta, Andre G. Buret. 2018. Hydrogen sulfide: an agent of stability at the microbiome-mucosa interface. *American Journal of Physiology-Gastrointestinal and Liver Physiology* **314**:2, G143-G149. [Crossref]
- 45. Mariela Castelblanco, Jérôme Lugrin, Driss Ehirchiou, Sonia Nasi, Isao Ishii, Alexander So, Fabio Martinon, Nathalie Busso. 2018. Hydrogen sulfide inhibits NLRP3 inflammasome activation and reduces cytokine production both in vitro and in a mouse model of inflammation. *Journal of Biological Chemistry* 293:7, 2546-2557. [Crossref]
- 46. I. M. Kovalchuk, M. R. Gzhegotsky, Y. F. Rivis, S. M. Kovalchuk. 2018. MODIFICATION OF THE FATTY ACID COMPOSITION OF PHOSPHOLIPIDS IN LIVER, MYOCARDIUM AND PLASMA TISSUES UNDER THE INFLUENCE OF IONIZING RADIATION AND WITH THE PRIOR APPLICATION OF A HYDROGEN SULFIDE DONOR. Bulletin of Problems Biology and Medicine 1.2:144, 130. [Crossref]
- 47. Qianqian Sun, Zixi Chen, Ping He, Yuan Li, Xiaoying Ding, Ying Huang, Hang Gu, Xin Ni. 2018. Reduced Expression of Hydrogen Sulfide–Generating Enzymes Down-Regulates 15-Hydroxyprostaglandin Dehydrogenase in Chorion during Term and Preterm Labor. *The American Journal of Pathology* 188:1, 63-71. [Crossref]
- Raphael R. Fagundes, Cormac T. Taylor. 2017. Determinants of hypoxia-inducible factor activity in the intestinal mucosa. *Journal of Applied Physiology* 123:5, 1328-1334. [Crossref]
- Da Zhang, Junbao Du, Chaoshu Tang, Yaqian Huang, Hongfang Jin. 2017. H2S-Induced Sulfhydration: Biological Function and Detection Methodology. Frontiers in Pharmacology 8. [Crossref]
- Huichen Zhao, Shengxia Lu, Jiachao Chai, Yuchao Zhang, Xiaoli Ma, Jicui Chen, Qingbo Guan, Meiyan Wan, Yuantao Liu. 2017. Hydrogen sulfide improves diabetic wound healing in ob/ob mice via attenuating inflammation. *Journal of Diabetes and its Complications* 31:9, 1363-1369. [Crossref]
- 51. M Jimenez, V Gil, M Martinez-Cutillas, N Mañé, D Gallego. 2017. Hydrogen sulphide as a signalling molecule regulating physiopathological processes in gastrointestinal motility. *British Journal of Pharmacology* **174**:17, 2805-2817. [Crossref]
- 52. Xiao-Xin Tan, Kao-Qi Lian, Xiang Li, Nan Li, Wei Wang, Wei-Jun Kang, Hong-Mei Shi. 2017. Development of a derivatization method for the quantification of hydrogen sulfide and its application in vascular calcification rats. *Journal of Chromatography B* 1055-1056, 8-14. [Crossref]
- 53. Xingji You, Zixi Chen, Huina Zhao, Chen Xu, Weina Liu, Qianqian Sun, Ping He, Hang Gu, Xin Ni. 2017. Endogenous hydrogen sulfide contributes to uterine quiescence during pregnancy. *Reproduction* **153**:5, 535-543. [Crossref]
- 54. Tian-Xiao Hu, Xuejing Guo, Gang Wang, Lu Gao, Ping He, Yang Xia, Hang Gu, Xin Ni. 2017. MiR133b is involved in endogenous hydrogen sulfide suppression of sFlt-1 production in human placenta. *Placenta* 52, 33-40. [Crossref]
- Nilkantha Sen. 2017. Functional and Molecular Insights of Hydrogen Sulfide Signaling and Protein Sulfhydration. *Journal of Molecular Biology* 429:4, 543-561. [Crossref]
- Florian Huber, Sören Riegert, Manfred Madel, Klaus Thonke. 2017. H 2 S sensing in the ppb regime with zinc oxide nanowires. Sensors and Actuators B: Chemical 239, 358-363. [Crossref]
- 57. Marcin Magierowski, Katarzyna Magierowska, Magdalena Hubalewska-Mazgaj, Zbigniew Sliwowski, Robert Pajdo, Grzegorz Ginter, Slawomir Kwiecien, Tomasz Brzozowski. 2017. Exogenous and Endogenous Hydrogen Sulfide Protects Gastric Mucosa against the Formation and Time-Dependent Development of Ischemia/Reperfusion-Induced Acute Lesions Progressing into Deeper Ulcerations. *Molecules* 22:2, 295. [Crossref]
- 58. Khosrow Kashfi, Mahnoush Esmaili. NO-H 2 S-Releasing Chimeras as a Multifaceted Approach to Cancer Therapy 105-142. [Crossref]
- Bo Li, Carol Lee, Zechariah Martin, Xinpei Li, Yuhki Koike, Alison Hock, Elke Zani-Ruttenstock, Augusto Zani, Agostino Pierro. 2017. Intestinal epithelial injury induced by maternal separation is protected by hydrogen sulfide. *Journal of Pediatric* Surgery 52:1, 40-44. [Crossref]

- 60. Sung Jae Shin, Choong-Kyun Noh, Sun Gyo Lim, Kee Myung Lee, Kwang Jae Lee. 2017. Non-steroidal anti-inflammatory drug-induced enteropathy. *Intestinal Research* 15:4, 446. [Crossref]
- 61. R. Pieper, C. Villodre Tudela, M. Taciak, J. Bindelle, J. F. Pérez, J. Zentek. 2016. Health relevance of intestinal protein fermentation in young pigs. *Animal Health Research Reviews* 17:2, 137-147. [Crossref]
- 62. Akbar Ahmad, Csaba Szabo. 2016. Both the H 2 S biosynthesis inhibitor aminooxyacetic acid and the mitochondrially targeted H 2 S donor AP39 exert protective effects in a mouse model of burn injury. *Pharmacological Research* **113**, 348-355. [Crossref]
- 63. Angela Ianaro, Giuseppe Cirino, John L. Wallace. 2016. Hydrogen sulfide-releasing anti-inflammatory drugs for chemoprevention and treatment of cancer. *Pharmacological Research* 111, 652-658. [Crossref]
- 64. Xuemei Yang, Jianxiu Du, Yang Zhou. 2016. Rapid and point of care measurement of sulfide in human serum with a light emitting diode-based photometer by marriage of gas separation with paper enrichment. *Sensors and Actuators B: Chemical* 232, 738-743. [Crossref]
- 65. Akbar Ahmad, Nadiya Druzhyna, Csaba Szabo. 2016. Delayed Treatment with Sodium Hydrosulfide Improves Regional Blood Flow and Alleviates Cecal Ligation and Puncture (CLP)-Induced Septic Shock. *Shock* 46:2, 183-193. [Crossref]
- 66. Erminia Donnarumma, Murtuza J. Ali, Amanda M. Rushing, Amy L. Scarborough, Jessica M. Bradley, Chelsea L. Organ, Kazi N. Islam, David J. Polhemus, Stefano Evangelista, Giuseppe Cirino, J. Stephen Jenkins, Rajan A. G. Patel, David J. Lefer, Traci T. Goodchild. 2016. Zofenopril Protects Against Myocardial Ischemia–Reperfusion Injury by Increasing Nitric Oxide and Hydrogen Sulfide Bioavailability. *Journal of the American Heart Association* 5:7. . [Crossref]
- 67. Joseph Candela, Gopal V. Velmurugan, Carl White. 2016. Hydrogen sulfide depletion contributes to microvascular remodeling in obesity. *American Journal of Physiology-Heart and Circulatory Physiology* **310**:9, H1071-H1080. [Crossref]
- 68. Martin Beaumont, Mireille Andriamihaja, Annaïg Lan, Nadezda Khodorova, Marc Audebert, Jean-Marc Blouin, Marta Grauso, Luciana Lancha, Pierre-Henri Benetti, Robert Benamouzig, Daniel Tomé, Frédéric Bouillaud, Anne-Marie Davila, François Blachier. 2016. Detrimental effects for colonocytes of an increased exposure to luminal hydrogen sulfide: The adaptive response. *Free Radical Biology and Medicine* 93, 155-164. [Crossref]
- Tian-Xiao Hu, Gang Wang, Xue-Jing Guo, Qian-Qian Sun, Ping He, Hang Gu, Ying Huang, Lu Gao, Xin Ni. 2016. MiR 20a,-20b and -200c are involved in hydrogen sulfide stimulation of VEGF production in human placental trophoblasts. *Placenta* 39, 101-110. [Crossref]
- 70. Csaba Szabo. 2016. Gasotransmitters in cancer: from pathophysiology to experimental therapy. *Nature Reviews Drug Discovery* 15:3, 185-203. [Crossref]
- Jennifer Pichette, Jeffrey Gagnon. 2016. Implications of Hydrogen Sulfide in Glucose Regulation: How H 2 S Can Alter Glucose Homeostasis through Metabolic Hormones. Oxidative Medicine and Cellular Longevity 2016, 1-5. [Crossref]
- 72. Akhlaq A. Farooqui. Effects of Curcumin on Neuroinflammation in Animal Models and in Patients with Alzheimer Disease 259-296. [Crossref]
- John L. Wallace, Gilberto de Nucci, Oksana Sulaieva. 2015. Toward More GI-Friendly Anti-Inflammatory Medications. Current Treatment Options in Gastroenterology 13:4, 377-385. [Crossref]
- 74. Salvatore Sutti, Irene Locatelli, Stefania Bruzzì, Aastha Jindal, Marco Vacchiano, Cristina Bozzola, Emanuele Albano. 2015. CX3CR1-expressing inflammatory dendritic cells contribute to the progression of steatohepatitis. *Clinical Science* **129**:9, 797-808. [Crossref]
- 75. Chang Su Lim, Bong Rae Cho. 2015. Two-photon probes for biomedical imaging. Tetrahedron 71:43, 8219-8249. [Crossref]
- 76. Suvajit Sen, Brian Kawahara, Divya Gupta, Rebecca Tsai, Marine Khachatryan, Sinchita Roy-Chowdhuri, Shikha Bose, Alexander Yoon, Kym Faull, Robin Farias-Eisner, Gautam Chaudhuri. 2015. Role of cystathionine β-synthase in human breast Cancer. Free Radical Biology and Medicine 86, 228-238. [Crossref]
- 77. G Leoni, P-A Neumann, R Sumagin, T L Denning, A Nusrat. 2015. Wound repair: role of immune-epithelial interactions. *Mucosal Immunology* 8:5, 959-968. [Crossref]
- John L. Wallace, Angela Ianaro, Kyle L. Flannigan, Giuseppe Cirino. 2015. Gaseous mediators in resolution of inflammation. Seminars in Immunology 27:3, 227-233. [Crossref]
- 79. Jean-Paul Motta, Kyle L. Flannigan, Terence A. Agbor, Jennifer K. Beatty, Rory W. Blackler, Matthew L. Workentine, Gabriela J. Da Silva, Rui Wang, Andre G. Buret, John L. Wallace. 2015. Hydrogen Sulfide Protects from Colitis and Restores Intestinal Microbiota Biofilm and Mucus Production. *Inflammatory Bowel Diseases* 21:5, 1006-1017. [Crossref]
- Marcin Magierowski, Katarzyna Magierowska, Slawomir Kwiecien, Tomasz Brzozowski. 2015. Gaseous Mediators Nitric Oxide and Hydrogen Sulfide in the Mechanism of Gastrointestinal Integrity, Protection and Ulcer Healing. *Molecules* 20:5, 9099-9123. [Crossref]

- 81. John L. Wallace, Rui Wang. 2015. Hydrogen sulfide-based therapeutics: exploiting a unique but ubiquitous gasotransmitter. *Nature Reviews Drug Discovery* 14:5, 329-345. [Crossref]
- 82. Kyle L. Flannigan, Terence A. Agbor, Jean-Paul Motta, José G. P. Ferraz, Rui Wang, Andre G. Buret, John L. Wallace. 2015. Proresolution effects of hydrogen sulfide during colitis are mediated through hypoxia-inducible factor-1α. *The FASEB Journal* 29:4, 1591-1602. [Crossref]
- 83. Marcin Magierowski, Katarzyna Jasnos, Slawomir Kwiecien, Danuta Drozdowicz, Marcin Surmiak, Malgorzata Strzalka, Agata Ptak-Belowska, John L. Wallace, Tomasz Brzozowski. 2015. Endogenous Prostaglandins and Afferent Sensory Nerves in Gastroprotective Effect of Hydrogen Sulfide against Stress-Induced Gastric Lesions. PLOS ONE 10:3, e0118972. [Crossref]
- Minho Kang, Atsushi Hashimoto, Aravind Gade, Hamid I. Akbarali. 2015. Interaction between hydrogen sulfide-induced sulfhydration and tyrosine nitration in the K ATP channel complex. *American Journal of Physiology-Gastrointestinal and Liver Physiology* 308:6, G532-G539. [Crossref]
- Megan K. Thorson, Ryan M. Van Wagoner, Mary Kay Harper, Chris M. Ireland, Tomas Majtan, Jan P. Kraus, Amy M. Barrios. 2015. Marine natural products as inhibitors of cystathionine beta-synthase activity. *Bioorganic & Medicinal Chemistry Letters* 25:5, 1064-1066. [Crossref]
- 86. M. Martinez-Cutillas, V. Gil, N. Mañé, P. Clavé, D. Gallego, M.T. Martin, M. Jimenez. 2015. Potential role of the gaseous mediator hydrogen sulphide (H2S) in inhibition of human colonic contractility. *Pharmacological Research* **93**, 52-63. [Crossref]
- Zoltán Pálinkás, Paul G Furtmüller, Attila Nagy, Christa Jakopitsch, Katharina F Pirker, Marcin Magierowski, Katarzyna Jasnos, John L Wallace, Christian Obinger, Péter Nagy. 2015. Interactions of hydrogen sulfide with myeloperoxidase. *British Journal of Pharmacology* 172:6, 1516-1532. [Crossref]
- 88. Wallace John L., Blackler Rory W., Chan Melissa V., Da Silva Gabriela J., Elsheikh Wagdi, Flannigan Kyle L., Gamaniek Iulia, Manko Anna, Wang Lu, Motta Jean-Paul, Buret Andre G. 2015. Anti-Inflammatory and Cytoprotective Actions of Hydrogen Sulfide: Translation to Therapeutics. *Antioxidants & Redox Signaling* 22:5, 398-410. [Abstract] [Full Text] [PDF] [PDF Plus]
- Rory W Blackler, Jean-Paul Motta, Anna Manko, Matthew Workentine, Premysl Bercik, Michael G Surette, John L Wallace. 2015. Hydrogen sulphide protects against NSAID-enteropathy through modulation of bile and the microbiota. *British Journal of Pharmacology* 172:4, 992-1004. [Crossref]
- 90. Burcu Gemici, John L. Wallace. Anti-inflammatory and Cytoprotective Properties of Hydrogen Sulfide 169-193. [Crossref]
- 91. Huimin Wang, Lixuan Mu, Guangwei She, Wensheng Shi. 2015. Silicon nanowires-based fluorescent sensor for in situ detection of hydrogen sulfide in extracellular environment. *RSC Advances* 5:81, 65905-65908. [Crossref]
- 92. Daniela Sieghart, Melissa Liszt, Axel Wanivenhaus, Hans Bröll, Hans Kiener, Burkhard Klösch, Günter Steiner. 2015. Hydrogen sulphide decreases IL -1&-induced activation of fibroblast-like synoviocytes from patients with osteoarthritis. *Journal of Cellular* and Molecular Medicine 19:1, 187-197. [Crossref]
- 93. Quansheng Du, Chao Wang, Nan Zhang, Guofeng Li, Meng Zhang, Liping Li, Qingzeng Zhang, Jianxin Zhang. 2014. In vivo study of the effects of exogenous hydrogen sulfide on lung mitochondria in acute lung injury in rats. BMC Anesthesiology 14:1. . [Crossref]
- 94. Iryna Fomenko, Alexander Sklyarov, Tetyana Bondarchuk, Lilya Biletska, Natalia Panasyuk, John L. Wallace. 2014. Effects of conventional and hydrogen sulfide-releasing non-steroidal anti-inflammatory drugs in rats with stress-induced and epinephrineinduced gastric damage. Stress 17:6, 528-537. [Crossref]
- 95. Vanitha Bala, Senthilkumar Rajagopal, Divya P. Kumar, Ancy D. Nalli, Sunila Mahavadi, Arun J. Sanyal, John R. Grider, Karnam S. Murthy. 2014. Release of GLP-1 and PYY in response to the activation of G protein-coupled bile acid receptor TGR5 is mediated by Epac/PLC-ε pathway and modulated by endogenous H2S. *Frontiers in Physiology* 5. . [Crossref]
- 96. Alberto F. Donatti, Rebeca M. Araujo, Renato N. Soriano, Leopoldo U. Azevedo, Christie A. Leite-Panissi, Luiz G.S. Branco. 2014. Role of hydrogen sulfide in the formalin-induced orofacial pain in rats. *European Journal of Pharmacology* 738, 49-56. [Crossref]
- 97. Meng Teng Peh, Azzahra Binti Anwar, David S.W. Ng, Mohamed Shirhan Bin Mohamed Atan, Srinivasan Dinesh Kumar, Philip K. Moore. 2014. Effect of feeding a high fat diet on hydrogen sulfide (H2S) metabolism in the mouse. *Nitric Oxide* 41, 138-145. [Crossref]
- 98. Wagdi Elsheikh, Rory W. Blackler, Kyle L. Flannigan, John L. Wallace. 2014. Enhanced chemopreventive effects of a hydrogen sulfide-releasing anti-inflammatory drug (ATB-346) in experimental colorectal cancer. *Nitric Oxide* **41**, 131-137. [Crossref]
- 99. V. Brancaleone, E. Mitidieri, R. J. Flower, G. Cirino, M. Perretti. 2014. Annexin A1 Mediates Hydrogen Sulfide Properties in the Control of Inflammation. *Journal of Pharmacology and Experimental Therapeutics* **351**:1, 96-104. [Crossref]

- 100. Kai Song, Fen Wang, Qian Li, Yong-Bing Shi, Hui-Fen Zheng, Hanjing Peng, Hua-Ying Shen, Chun-Feng Liu, Li-Fang Hu. 2014. Hydrogen sulfide inhibits the renal fibrosis of obstructive nephropathy. *Kidney International* 85:6, 1318-1329. [Crossref]
- 101. Alicia Madurga, Ivana Mižíková, Jordi Ruiz-Camp, István Vadász, Susanne Herold, Konstantin Mayer, Heinz Fehrenbach, Werner Seeger, Rory E. Morty. 2014. Systemic hydrogen sulfide administration partially restores normal alveolarization in an experimental animal model of bronchopulmonary dysplasia. *American Journal of Physiology-Lung Cellular and Molecular Physiology* 306:7, L684-L697. [Crossref]
- 102. Péter Nagy, Zoltán Pálinkás, Attila Nagy, Barna Budai, Imre Tóth, Anita Vasas. 2014. Chemical aspects of hydrogen sulfide measurements in physiological samples. *Biochimica et Biophysica Acta (BBA) General Subjects* 1840:2, 876-891. [Crossref]
- Robert E. Click. 2014. Review: 2-Mercaptoethanol alteration of in vitro immune functions of species other than murine. *Journal of Immunological Methods* 402:1-2, 1-8. [Crossref]
- 104. Robert E. CLICK. 2014. A review: alteration of in vitro reproduction processes by thiols Emphasis on 2-mercaptoethanol. *Journal of Reproduction and Development* **60**:6, 399-405. [Crossref]
- 105. Ling Li, Philip K Moore. 2013. Could hydrogen sulfide be the next blockbuster treatment for inflammatory disease?. *Expert Review of Clinical Pharmacology* 6:6, 593-595. [Crossref]
- 106. Yijie Zheng, Naixiang Luo, Dongzhen Mu, Pei Jiang, Ronghua Liu, Haozhe Sun, Shudao Xiong, Xiaoming Liu, Luman Wang, Yiwei Chu. 2013. Lipopolysaccharide regulates biosynthesis of cystathionine γ-lyase and hydrogen sulfide through toll-like receptor-4/p38 and toll-like receptor-4/NF-κB pathways in macrophages. In Vitro Cellular & Developmental Biology - Animal 49:9, 679-688. [Crossref]
- 107. Chang Su Lim, Sajal Kumar Das, Sun Young Yang, Eun Sun Kim, Hoon Jai Chun, Bong Rae Cho. 2013. Quantitative Estimation of the Total Sulfide Concentration in Live Tissues by Two-Photon Microscopy. *Analytical Chemistry* 85:19, 9288-9295. [Crossref]
- 108. Melissa V. Chan, John L. Wallace. 2013. Hydrogen sulfide-based therapeutics and gastrointestinal diseases: translating physiology to treatments. *American Journal of Physiology-Gastrointestinal and Liver Physiology* **305**:7, G467-G473. [Crossref]
- 109. Kyle L. Flannigan, Jose G. P. Ferraz, Rui Wang, John L. Wallace. 2013. Enhanced Synthesis and Diminished Degradation of Hydrogen Sulfide in Experimental Colitis: A Site-Specific, Pro-Resolution Mechanism. *PLoS ONE* **8**:8, e71962. [Crossref]
- 110. Xinggui Shen, Mattias Carlström, Sara Borniquel, Cecilia Jädert, Christopher G. Kevil, Jon O. Lundberg. 2013. Microbial regulation of host hydrogen sulfide bioavailability and metabolism. *Free Radical Biology and Medicine* **60**, 195-200. [Crossref]
- 111. Nan Chiang, Masakazu Shinohara, Jesmond Dalli, Valbona Mirakaj, Megumi Kibi, Augustine M. K. Choi, Charles N. Serhan. 2013. Inhaled Carbon Monoxide Accelerates Resolution of Inflammation via Unique Proresolving Mediator–Heme Oxygenase-1 Circuits. *The Journal of Immunology* 190:12, 6378-6388. [Crossref]
- 112. Robert E. Click. 2013. Dietary supplemented 2-mercaptoethanol prevents spontaneous and delays virally-induced murine mammary tumorigenesis. Cancer Biology & Therapy 14:6, 521-526. [Crossref]
- 113. Huai-Dong Li, Zhao-Rui Zhang, Qing-Xiang Zhang, Zhi-Chu Qin, Deng-Ming He, Jin-Song Chen. 2013. Treatment with exogenous hydrogen sulfide attenuates hyperoxia-induced acute lung injury in mice. *European Journal of Applied Physiology* 113:6, 1555-1563. [Crossref]
- 114. Qiongqiong Wan, Yanchao Song, Zhao Li, Xinghui Gao, Huimin Ma. 2013. In vivo monitoring of hydrogen sulfide using a cresyl violet-based ratiometric fluorescence probe. *Chem. Commun.* 49:5, 502-504. [Crossref]
- 115. Akira Murakami. 2013. Modulation of protein quality control systems by food phytochemicals. *Journal of Clinical Biochemistry* and Nutrition 52:3, 215-227. [Crossref]
- 116. Robert E Click. 2013. Anticancer activity and chemoprevention of xenobiotic organosulfurs in preclinical model systems. Oncology Discovery 1:1, 4. [Crossref]
- 117. Ling Li, Mohamed Shirhan Bin Mohamed, Philip K. Moore. Multiple Roles of H2S in Inflammation: A New Class of Therapeutics? 63-82. [Crossref]
- 118. A. R. Gade, M. Kang, H. I. Akbarali. 2013. Hydrogen Sulfide as an Allosteric Modulator of ATP-Sensitive Potassium Channels in Colonic Inflammation. *Molecular Pharmacology* 83:1, 294-306. [Crossref]
- 119. Amiram Ariel, Orly Timor. 2013. Hanging in the balance: endogenous anti-inflammatory mechanisms in tissue repair and fibrosis. *The Journal of Pathology* **229**:2, 250-263. [Crossref]
- 120. Leticia R. Benetti, Daiana Campos, Sonia A. Gurgueira, Anibal E. Vercesi, Cristiane E.V. Guedes, Kleber L. Santos, John L. Wallace, Simone A. Teixeira, Juliana Florenzano, Soraia K.P. Costa, Marcelo N. Muscará, Heloisa H.A. Ferreira. 2013. Hydrogen sulfide inhibits oxidative stress in lungs from allergic mice in vivo. *European Journal of Pharmacology* 698:1-3, 463-469. [Crossref]
- 121. Neil Dufton, Jane Natividad, Elena F. Verdu, John L. Wallace. 2012. Hydrogen sulfide and resolution of acute inflammation: A comparative study utilizing a novel fluorescent probe. *Scientific Reports* **2**:1. . [Crossref]

- 122. J P Hunter, S A Hosgood, M Patel, R Rose, K Read, M L Nicholson. 2012. Effects of hydrogen sulphide in an experimental model of renal ischaemia–reperfusion injury. *British Journal of Surgery* **99**:12, 1665-1671. [Crossref]
- 123. Stefania Merighi, Stefania Gessi, Katia Varani, Debora Fazzi, Pier Andrea Borea. 2012. Hydrogen sulfide modulates the release of nitric oxide and VEGF in human keratinocytes. *Pharmacological Research* 66:5, 428-436. [Crossref]
- 124. Franck Carbonero, Ann C. Benefiel, H. Rex Gaskins. 2012. Contributions of the microbial hydrogen economy to colonic homeostasis. *Nature Reviews Gastroenterology & Hepatology* 9:9, 504-518. [Crossref]
- 125. Yi-Hong Liu, Ming Lu, Li-Fang Hu, Peter T.-H. Wong, George D. Webb, Jin-Song Bian. 2012. Hydrogen Sulfide in the Mammalian Cardiovascular System. *Antioxidants & Redox Signaling* 17:1, 141-185. [Abstract] [Full Text] [PDF] [PDF Plus]
- 126. Ammar K.H. Alshorafa, Qing Guo, Fanqin Zeng, Mingchun Chen, Guozhen Tan, Zengqi Tang, Ruofei Yin. 2012. Psoriasis Is Associated with Low Serum Levels of Hydrogen Sulfide, a Potential Anti-inflammatory Molecule. *The Tohoku Journal of Experimental Medicine* **228**:4, 325-332. [Crossref]